

**EPICARDIAL AUTOFLUORESCENCE NAD(P)H KINETICS IN THE ISCHEMICALLY PRECONDITIONED LANGERDORFF RAT HEART. EFFECTS OF CAPSAICIN. PART 1.**

LUIGI ROSSINI<sup>+</sup>, BOZENA KUZIO, ROXANNE DESLAURIERS AND CRAIG W. JOHNSON<sup>°</sup>

*Institute of Biodiagnostics, National Research Council Canada, 435 Ellice Avenue, Winnipeg, Manitoba, Canada; \*I.M.O., Section of Human Pharmacology and Toxicology, the Polytechnic University of Marche, Department of Neuroscience, Faculty of Medicine, Via Tronto 10A, Ancona, Italy and °Department of Statistics and Epidemiology, Health Science, University of Texas Medical Center, Houston, U.S.A.*

KEY WORDS OR PHRASES: Ischemic preconditioning; paradoxical contracture; autofluorescence; Langerdorff rat heart; capsaicin

SHORTENED RUNNING TITLE: Autofluorescence, capsaicin, ischemic preconditioning and paradoxical contracture in Langendorff rat heart

-----  
+ Corresponding author. On leave of absence from I.M.O - Pharmacotoxicology, the Polytechnic University of Marche, Ancona, Via Tronto 10 A, 60020 Ancona, Italy. E-mail [l.rossini@univpm.it](mailto:l.rossini@univpm.it); Tel. n. 0039-071-2181028; Fax n. 0039-071-2206037. L.R. is recipient of a grant from the Medical Research Council Canada and the Centro Nazionale delle Ricerche Italy. The work represents a contribution to celebrate the teaching and research activities of Professor Britton Chance.

### SUMMARY

The data presented here are preceded by a review of the conventional *ex vivo* model of conditioned ischemic learning. The fluorescence kinetics of indistinct pools of reduced pyridinnucleotides, together with the conventional functional parameters observed in short, long and preconditioning preperfusion conditions, as well as in the common phase of protracted ischemia (30 min) and in the phase of reperfusion monitored over 60 min, demonstrate significant trends in the context of the more recent integrated metabolic observations of heart perfusion after capsaicin pretreatment. The present paper reports the mean kinetic values of the metabolic parameters and those from functional sampling, with emphasis on protracted ischemia following short (15 min) and long (1 h) perfusion with and without short preconditioning ischemic insults in capsaicin-treated and untreated specimens. We also describe some trends observed in single cases, where the phases of preischemic conditioning, protracted ischemia, and reperfusion demonstrated both the expected protection effect and the prevalent damage. The paper examines the significance of this experimental model in the biological and pharmacotoxicological integrated context.

## 1. INTRODUCTION

### 1.1. LITERATURE OVERVIEW

Oxygen use and short-term adaptation to its deprivation have been analyzed in a number of experimental models. Cardiovascular observations have mainly focused on ischemic/anoxic and reperfusion damage and protection. Some of the most recent studies have been conducted not only *in vivo* and *ex vivo* on native, supposedly intact organs and tissues perfused with blood or crystalloid buffered solutions, but also on the cellular and subcellular samples. Different phenotypes and/or molecular dynamics and kinetic behaviours have been observed with cultured (24-48 h) cardiomyocytes vs freshly isolated myocytes and isolated perfused hearts, as well as within immature vs mature isolated cells and hearts from young vs aged rodents and other non human widely used experimental preparations [1-11]. Regional or global ischemia has been obtained *in vivo*, *in situ* or *in vitro* by occlusion of the coronary flow or by stopping the perfusion. The perfusate was mostly Krebs-Henseleit (KH) bicarbonate buffer, reequilibrated with a 95% O<sub>2</sub> and 5% CO<sub>2</sub> mixture, previously equilibrated with a mixture of 95% N<sub>2</sub> and CO<sub>2</sub> at pH 7.40. The preparations were maintained at constant temperature during observations and measurements also under occlusion, to avoid hypothermia- induced cardioprotection.

The Langerdorff rodent heart preparation is a widely used model because the most advanced metabolic properties have been monitored associated with the evaluation of traditional functional parameters. In the isolated rat heart (r.h.), transient ischemic endogenous preconditioning (IPC) has previously been shown firstly mediated via a subfamily of protein kinase C (PKC) activation and translocation coupled to  $\alpha_1$ -adrenoceptor and  $B_2$  associated bradykinin receptors [12-13], or partly through endothelial function and  $B_1$  - not  $B_2$  - receptors [14-15]. IPC is not affected by depletion of endogenous catecholamines resulting from reserpine or 6-hydroxydopamine treatment [16]. Activation of the  $\alpha_1$  adrenergic receptor has been shown to confer protection against the lethal injury from  $Ca^{2+}$  preconditioning (via the protein kinase C signaling pathway) [17]. Hearse and Sutherland [18] have more recently observed paradoxical exacerbation of contracture [19-22] followed by enhanced post-anoxic recovery both under ischemic and l-nor-epinephrine preconditioning (PC), and Hearse, Ferrari and Sutherland [23] observed PC, but not paradoxical contracture in blood perfused r.h. during ventricular fibrillation and/or rapid pacing.

In the transient, early energy imbalance of IPC protocols, a small population of  $\alpha$ -G, s or i subunit proteins appears to be involved as coupled to muscarinic  $M_2$  receptors and  $A_1$  adenosine receptors [24-26].

Adenosine does not mediate improvement of functional recovery after PC in globally ischemic, isolated Langerdorff r.h. [27]. Increased adenosine formation through  $\beta$ -adrenergic receptors and noradrenaline release protects ischemic rat heart after hypoxic PC [27, 28-30]. Adenosine mediates persistent adrenergic desensitization in the r.h. via activation of iso-PKCs [31]. Although targeted deletion of the  $A_3$  Adenosine receptor confers resistance against myocardial ischemic/reperfusion injury,  $A_3$ ARs are not required for the development of the early phase of IPC [32]. Additional references related to the open selectivity of the nucleoside receptors and transporters are mentioned below [133-136, 145, 155, 161].

Upregulation of cardiac uptake1 carrier and related loss of extra-tissutal norepinephrine increase under ischemia and thereafter - except in the 1st minute - up to more than 20 min through reperfusion [33].

Activation and translocation of iso-PKCs appear to be key events in r.h. ischemic and reperfusion damage, as well as in IPC [34-36]. In the same Langerdorff r.h. preparation, inhibition of some iso-PKC, which limits ischemic injury and eliminates the effect of IPC on stunning during reflow, is not related to PC attenuation of acidification [37-39]. In different cardiomyocytes from transgenic or normal *in* or *ex vivo* heart, not only the protein kinase C  $\epsilon$  and  $\delta$  isoforms, which

have been seen to have opposite effects [40-41], but also protein kinase A has been found to be independently associated to IPC (see also [42-47]).

RNA expression of the Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1 is rapidly regulated in acutely ischemic rat myocardium [48].

Inhibition of Na<sup>+</sup>/H<sup>+</sup> exchange adds to the protective effect of IPC [49-51], but the same [pH]<sub>i</sub> decrease attenuation does not appear to be tightly coupled to Na<sup>+</sup>/H<sup>+</sup> turnover [52]. In reperfusion injury, exchange of accumulated Na<sup>+</sup> with Ca<sup>2+</sup> is detrimental to function [53-57]. Dietary cariporide, a Na<sup>+</sup>/H<sup>+</sup> exchange inhibitor, as well as treatment with an inhibitor of the reverse mode of the Na<sup>+</sup>/Ca<sup>2+</sup> exchanger, confer cardioprotection following coronary occlusion and reperfusion [58-60]. Sodium and calcium overloads [61-63] and protection by metabolic uncoupling in reperfusion [64-66] may contribute to the understanding of our present data (see also [67]).

Some observations may be related to methodological conditions: in the case of myocardial stunning, an important functional impairment parameter in the evaluation of hypoxia-ischemia and reperfusion dynamics, the isovolumic preparation, has sometimes been subjected to changes in systolic and diastolic pressure by collapse and reinflation of the left ventricular balloon in order to counteract the no-reflow phenomenon [20-21, 52, 68]. Pacing at 2 Hz at 35 °C showed delay of ischemic contracture [50] instead of exacerbation. In the heart not immersed and overdrive paced (300-

330 bpm), earlier contracture development was found only after repetitive PC [69].

Modulation of stunning by glycolysis, glyconeogenesis, glycogenolysis, and associated balance of proton production and cytosolic (coupled) export, are still debated (see [70-79] and below).

The role of endogenous NO in monophosphoryl lipid A acute cardioprotection in the working isolated perfused r.h. [80] has not been confirmed in the isolated retrogradely perfused isovolumic preparation - either under constant flow or pressure perfusion - as a mediator of early IPC [81], whereas in a feline study NO-peroxynitrite exchange has been confirmed to be cardioprotective [82-83], and in mice cardiac myocyte IPC has been found to contribute integratively by both inducible and constitutive NOSs [84-85], a topic under that is rapidly evolving: NOSs "imported" by rapidly IPC-recruited endothelial progenitor cells mediate a protective myocardial effect [86]. Although oxygen-derived free radicals are not believed to contribute to PC in the r.h. [87], they are held to play a role both in reperfusion injury [88] and in IPC [89], as they affect myocardial stunning [90-91].

Downregulation of the Na<sup>+</sup>-creatine cotransporter has been considered an important feature of the failing myocardium [92]. The dual regulation of muscle AMP-activated protein kinase, which inhibits the creatine kinase-phosphocreatine system and is inhibited

by phosphocreatine, while creatine antagonizes this inhibition [93], may be active in the heart.

IPC has been reported to be independent of r.h. mitochondrial  $F_1F_0$ -ATPase inhibition [94], whereas other researchers consider its integrity essential, and not only in this rodent assay (i.e.: rat [95]; dog [96]). In the same perfused r.h., mitochondrial vs glycolytic phosphate and redox potential sensitive mechanisms have been shown to be involved in the protection afforded by IPC ([68]. See the Discussion section). The sulphhydryl redox potential modulates sarcoplasmic reticulum  $Ca^{2+}$  release in PC [97], even though glutathione depletion has not been found to be essential to ventricular reperfusion arrhythmias [98]. The large production of oxygen radicals from ischemic mitochondria in the Langerdorff r.h. has been measured in 1991 [99], and the importance of mitochondrial/cytosolic couplings in acute short-term ischemic/reoxygenation cycles, particularly in IPC transients, indirect pyridine redox potential [100] and sarcoplasmic reticulum  $Ca^{2+}$  turnover, have recently been restated in rat heart by Dhalla and Brandes et al. [101-102], after Zucchi et al. [103], as previously observed in ischemic heart failure in the guinea pig [104]. The initial phases of ischemia are associated with a time-dependent positive imbalance in mitochondrial oxyphosphorylation reactions [105-106]. Downregulation of oxygen demand, and altered mechanisms of energy transfer have also been reported in acute hypoxia [107].



PC mechanisms activated by stretching tissues (injection of salts into the myocardium [108]), those elicited by pharmacological substances such as ethanol [109] and general anesthetics (i.e. [110]), or phosphodiesterase-5 and/or -6 inhibitors [111] occurring in endothelia [Cf.: 84, 112-113], or associated with the heat stress to cytoprotection, finally appear to develop through  $K_{ATP}$  channels [114-118], and are not related to enhanced action potential duration in a dog model [119]. Use of K channel openers, such as cromakalim and bimakalim, pinacidil and microrantil, and antagonists such as glibenclamide, glyburide and 5-hydroxydecanoate, has confirmed the role of these channels in ischemia and reperfusion phenomena [120-122]. In the r.h., cardioprotection, but not PC, is related to the special  $K_{IR}$ , the inward-rectifier potassium channel assayed by dofetilide and terikalant [123]. Nevertheless, in the Langendorff r.h. calcium PC, but not IPC, bypasses the  $K_{ATP}$  channel, a model condition that may explain why patients chronically exposed to sulphonylurea hypoglycemics remain protected [124]. Mitochondrial  $K_{ATP}$  channels [125], as proven with diazoxide [36, 126-127], have been shown to be an (i.e. one) end-point receptor/effector contributing to triggering and mediation of cardioprotective effects in r.h., not only in acute and chronic ischemia/reperfusion, but also in both early and delayed PC [128] (the same holds true in rabbit ventricular myocytes, e.g.: [129]).

As the cross-talks among  $K^+$  specific channels, NO and interchanged reactive oxygen species, noradrenaline, mostly  $\beta$  [130-132], and adenosine subfamilies of receptors and transporters [31,

133-136] are studied, naloxone sensitive  $\delta$ 1 opioid receptors - not  $\mu$  or  $\kappa$ , which appear to be related to delayed cardioprotection [137-143; i.e.: 144] -, and other peptidergic extended modulations, continue to be evaluated. Mitochondrial vs cytosolic phosphorylation and redox subcellular control networks are the focus of present research, both in r.h. and cardiac myocytes; the same problems are also being studied, for example, in rabbit [145] and chick cardiomyocytes [146-148]. G-Protein-coupled receptor internalization and primary triggering vs secondary processing signaling pathways, even in the immediate phase of protection against ischemia reperfusion injury - which consists both of irreversible necrosis and apoptosis by induction of phosphatidyl inositol 3-OH, P13-kinase, but not the p42/p44 cascade [149] -, are actually required to act together on mitochondria for IPC cardioprotection [150-162]. However, more integrated approaches extend the analyses to the responsive transcription factors [i.e.: 163].

### *1.2. THE CAPSAICIN-VANILLOID TOOL*

In the same way as reserpine and 6-hydroxydopamine [16] have been used to prevent catechol-dopaminergic and enteraminergic/serotonergic sympathetic neurotransmission, capsaicin and other vanilloids have been used (after Jánóso and Jánóso-Gabór [164]) to activate by release nonadrenergic, noncholinergic, mostly peptidergic modulators, and produce a long-lasting refractory state referred to as desensitization. A cloned subset

of capsaicin activated cation channel receptors has been associated with thermal and proton sensitive neuronal functions [165], and multiple iso-receptor groups have been characterized [166] by use of specific agonists and antagonists both in newborn [167-168] and adult mammals [169-171]. In the r.h., capsaicin targets and mechanisms of action have mostly been related to specific primary Ad-1 small myelinated centrally signaling afferent fibers that selectively contribute to the short local efferent circuits, which are activated by transient hypoxia and by anoxia/ischemia, underpinning chronic neuro-inflammatory disorders.

These structures release endogenous bradykinin, substance P and other tachykinins, atriopeptin(s) and  $\alpha$ -calcitonin-gene-related peptide (CGRP), which has in turn been correlated to oxygen deprivation/redistribution insults and even to IPC adaptation [172-174].

Epoxy eicosatrienoid acid products of cytochrome P450 epoxygenases - like the CYP2J2 human cloned isoform [175-176], contribute to the endogenously activated anandamide reactive cannabinoid receptors on peripheral sensory nerves, showing selectivity to capsaicin-vanilloid receptors accompanied by release of CGRP [177]. While capsaicin induces a reversible stimulus length dependent negative staircase inotropic effect in the rat ventricle, without inhibition of its calcium handling [178-179], its anti-arrhythmic and anti-ischemic activity has been postulated to act by blocking

some K<sup>+</sup> and/or Ca<sup>2+</sup> channels [180-181], possibly through the release of neuropeptides, especially CGRP. Modulation of coronary circulation [182], an interplay (in guinea pig heart) between NO and CGRP in capsaicin induced increase in coronary flow and heart rate [183], and capsaicin related r.h. PC [184] associated with oxygen radicals and NO contributions (i.e.: [185]) or pacing induced [186], have been reported. (Our preparatory work on cannabinoids vs vanilloids, and a contribution on some O<sub>2</sub>-NO-redox dependent structurally covalent post-translational cellular issues are reported in [187, 188]).

### 1.3. GENERAL AIM OF THE STUDY

We present our first paper, divided into two parts, on autofluorescence, and the second and third contributions on near infrared and NMR spectrometry studies. In this 1<sup>st</sup> paper, the adult r.h. spontaneously beating Langerdorff preparation was used while submerged and infused at a constant standard temperature and pressure with the widely used crystalloid buffered solution. Three sets of control conditions (short preinfusion, long preperfusion, and a commonly applied preconditioning protocol) were established in a total of 41 hearts, 18 of which were acutely pretreated *in vitro* at one capsaicin saturating dose. All hearts were thereafter subjected to 30 min global ischemia/anoxia followed by at least 60 min oxygenated

reperfusion, while conventional functional parameters were continuously monitored. For the fluorescence observations (2<sup>nd</sup> part), 3 other control groups and 3 groups of capsaicin-pretreated hearts were monitored in the same previously standardized conditions. All hearts of all animals sacrificed were used in the experiments.

The work first assessed the suitability of the most commonly monitored functional parameters to characterize *ex vivo*, in the preconditioned rat heart, early amelioration, protection or delay in recovery, following the *in vivo* original studies (i.e.: [189]) and the most recent hypotheses (i.e.: [190]) and contributions, as briefly reviewed above.

Noninvasive technique(s) were applied to analyze the kinetics of the interrelations of the most relevant redox markers - pyridine nucleotide fluorescence signals in this first paper. Their unique properties to express mito-cytosolic dynamic equilibria will help - it is our aim and basic hypothesis - to clarify the feedback interrelations among different organ/tissue/cell functional compartments. In particular, the optical techniques applied are held to be sufficiently fast to identify in a *peripheral* network of coupled metabolic vs functional adaptations, memory acquisition and maintenance processes, substrate/oxygen use vs deprivation precursor-product relationships and signaling. Our aim is thus to characterize matching of energy demand with respect to supply and the related damage vs protection features in the *in vitro*

*preparation*, after having analyzed some of them in our previous studies [191-198]. So, we would like to contribute to a description of those steps which, in the native *in vitro preparation*, are basic to modulation of energy availability vs oxygen deprivation processes associated with repeated short time insults, particularly in the frequency domain control of the metabolic machinery. Last but not least, capsaicin specific mechanisms will help clarify some *peripheral* residual short memory acquired adaptations in a model free of other neurohormonal and vascular factors [i.e.: 187]. The other noninvasive measurements of the metabolic parameters, analyzed by near-infrared and NMR spectrometric techniques, both *in vivo*, as well as *in vitro* capsaicin treatments, will be presented In the next two papers, and their modulation in ischemic and reperfusion injuries and interference on acute adaptation/attenuation throughout early IPC will be more comprehensively elaborated (work in progress).

#### 1.4. TOPICS NOT ANALYZED

The second window of protection, i.e. the delayed effects of preconditioning (for r.h.: [199-203], for mouse heart [204], and for conscious rabbit heart, with different mechanisms shown at 24 vs 72 hours, [205]), the effects of remote and transferable preconditioning ([201, 204-208]), and those of the form of modified reperfusion called post-conditioning ([209-210] were excluded from the study.

Some preliminary considerations and single heart-data slides have been presented in a local Academy Seminar [211].

## **2. MATERIAL AND METHODS**

### *2.1. HEART PREPARATION*

All experiments met the guidelines of the Canadian Council on Animal Care regarding the care and use of experimental animals, and were approved by the local Animal Committee of the National Research Council of Canada.

Sprague Dawley rats of both sexes, weighing  $250 \pm 15$  (S.D.) g, obtained from Charles River and acclimatized to animal facilities were submitted to 12 hour cycles of artificial light at constant temperature and relative humidity for at least one week prior to use, standard food and water being allowed ad libitum. The rats were anesthetized with sodium pentobarbital (120 mg/kg ip), and the hearts removed as soon as the toe reflex disappeared (within 3 min), immediately immersed in ice-cold buffer and perfused according to Langerdorff at  $36.5 \pm 0.1$  °C in less than 30 sec at a constant pressure of 80 mm Hg. The Krebs-Henseleit (KH) buffer contained (mM) NaCl 118, KCl 4.7, CaCl<sub>2</sub> 1.75 (free Ca<sup>2+</sup>  $\approx$  1.1), MgSO<sub>4</sub> 1.2, EDTA 0.5, NaHCO<sub>3</sub> 25 and glucose 11, and was equilibrated at pH of 7.4 with a 95% N<sub>2</sub> and 5% CO<sub>2</sub> gas mixture prior to the 95% O<sub>2</sub>/5% CO<sub>2</sub> gas mixture. An apical drain was inserted via the mitral valve in the left ventricle to vent the drainage from the thebesian veins, and a

water-filled compliant balloon was placed into the same ventricle. The balloon was connected to a Statham P23Db, or to WPI BLPR5326 (Sarasota, FL, USA) pressure transducers to monitor left ventricular pressure and heart rate. The left ventricular end diastolic pressure was adjusted to the averaged initial % of any maximum systolic pressure of 7.5 mm Hg by inflating the balloon, its volume being kept constant throughout all experiments. Functional parameters were monitored with a Digi Med<sup>R</sup> Instantaneous Data-capture and Analysis System (model 200, Micro-Med Inc., Louisville, Ky, USA), by sampling at 600 Hz and monitoring the successive 120 sec arithmetic averages.

## **2.2. FUNCTIONAL PARAMETERS**

The first parameter, coronary flow (CF), was followed with an ultrasonic blood flow meter (model T101, Transonic Systems Inc., Ithaca, New York) standardized by repeated collection of the effluent from the heart. Hearts were subjected to periods of global ischemia by clamping the perfusion line to the aortic cannula; reperfusion was achieved by releasing the clamp; the dead volume of fluid up to the aorta was maintained constant and equal to 13.50 ml. Mechanical



function was assessed as frequency (BPM), maximum systolic left ventricular pressure (MSLVP), and end diastolic left ventricular pressure (EDLVP). These parameters were used to obtain: rate pressure product; heart rate times left ventricular developed pressure (systolic minus diastolic pressure) (RPP); and RPP divided by the coronary flow (RPP/CF). The interleaved lengths of unspecified arrhythmia were taken into account. The internal heart temperature was monitored continuously using a thermocouple (model 39641-T Atkins Technology Inc., Gainesville, Florida, USA) placed into the pulmonary artery.

### 2.3. PROTOCOLS

The three standard sets of assays consisted of *controls* and *in vitro* capsaicin-pretreated preparations.

#### 2.3.1. Short perfusion (SP)

All hearts were observed for 15 min after the start of the perfusion. In treated specimens, after oxygenated KH perfusion and monitoring of all parameters for 5 min, a capsaicin/DMSO (see below) solution was infused through a collateral line at the top of the Langerdorff cannula for 5 min; the bathing fluid external to the heart was then substituted with control oxygenated, 36.5 °C KH Ringer, whose infusion was protracted up to the end of the *first* step of the SP protocol. The *second step* consisted of 30 min global ischemia and the *third step* of 60 min constant pressure oxygenated reperfusion. At the end of the

protocol, the heart was removed from the fluid and weighed or frozen immediately with Wollenberger clamps precooled with liquid nitrogen.

### *2.3.2. Long Perfusion (LP)*

After the *first* 15 min step, perfusion was continued for an additional 45 min before ischemia and reperfusion (*second* and *third* steps unchanged). Long perfusion, from 30 to 60 min, in the isolated r.h. perfused with glucose as the only external substrate, has been shown to correlate with O<sub>2</sub> uptake and decreased mechanical activity [212-213]. Osmotic swelling, a key feature of ischemic/reperfusion injury, is attenuated by activation of volume regulated chloride channels, a candidate for the final step of ischemic preconditioning, which is the subject of debate due to contradictory results obtained in isolated perfused rabbit heart and isolated cardiomyocytes [1-6].

### *2.3.3. Preconditioning (PC)*

After the first step, three cycles of 6 min global ischemia/anoxia, each following the first and the last by 10 min, and the middle by 8 min reperfusion, were repeated before the final long ischemia and reperfusion. The PC protocol replaced the LP, which acted as the most appropriate control.

A series of different conditions (i.e.: insults of varying ischemia and reperfusion times, from 1 to 10 PC), and final ischemia from 15 up to 45 min, which may encompass distinct mechanisms of regulation of iso-PKCs [214], were also assayed in different groups of rats (not included in the presentation) to assess the effect of PC on a roughly 50% recovery as evaluated through RPP. Another set of

hearts was treated with rising concentrations of DMSO, which is known to interfere with myocardial contractility and ischemic transients [215]. Capsaicin from 0.1 to 30  $\mu$ molar was also assayed for one up to 10 min perfusion.

#### **2.4. AUTOFLUORESCENCE**

The direct fluorimetric technique for recording intracellular oxidation-reduction states (i.e.: [193-194, 217]) was performed using a commercial instrument (Ratiometer and Quantimeter Photon Technology Int. Inc., S. Brunswick, NJ, USA). The OC-4000 optical chopper and the shutter controller were used with a 100 W LPS-220 Xenon lamp power supply and a 710 photon multiplier system, interfaced with Felix software. The 340 nm peak FS10-25, AM28470-03, and 430 nm peak FS10-25, AM28128-01 (Andover Corporation, Salem, NH, USA) excitation and emission filters were used. Even though the flavin-ox 436 - 460 excitation vs the 570 - 580 emission nm peaks was a possible second channel for time-shared observation (i.e.: [194, 217-221]), the second interleaved channel was used to monitor the light scattering (at 550 nm), which did not show any coherent optical changes. At the wavelength used, in the glucose-enriched KH perfusate, the surface fluorescence of the intact organ has been confirmed to originate from reduced nicotinamide nucleotides in mitochondria [222], with a contribution from cytosolic exchanges mostly pertaining to modulation by glyceraldehyde 3-

phosphate- and lactate dehydrogenases (i.e.: [223]), and the NADH shuttle system (i.e.: [224]).

A custom bifurcated fiber optic bundle (Ceram Optics, Enfield, CT, USA) that delivered the UV excitation light to the heart and also collected the emitted light was used. The common end of the optic fiber was a stainless steel cylinder 5 mm in external diameter at the polished optical end. This was placed through a hole in the water bath such that the left ventricular wall of the totally submerged heart focused at 4 mm distance, which had been adjusted to the best signal to noise ratio. The end of the fibers and the surface of the ventricle were maintained at a fixed distance with a lucite chamber empty of fluid by adjustment of the maximum energy emission.

Calibration with NADH in a KH solution confirmed the linearity of fluorescence over a wide range of concentrations, including those observed in the heart. After initial monitoring of quenching (less than 10% when observations were performed for 30 sec any 3 min), the fluorescence emission was found to be stable (less than  $\pm 5\%$  variation) in each heart for as long as 3 hours, the maximum allowed for any experiment when the irradiation chopped 3 times/sec. For comparisons with the functional parameters, fluorescence emissions were averaged up to 120 sec intervals (abscissae), and their values (ordinatae) standardized as % between the zero, almost steady, initial level and the maximum value, taken as 100%, reached in the 30 min ischemia.

It should be noted that our approach could not monitor the turnover and magnitude kinetics of heterogeneous ischemic areas in the perfused r.h., which can be evaluated using advanced imaging technologies [225-235].

### **2.5. STATISTICAL ANALYSIS**

Fluorescence data and single cardiac functional parameter kinetics were averaged to achieve coincident steps at 120 sec intervals, and their trends were evaluated in the 6 groups. The absolute values of the four functional parameters of each heart evaluated as independent - CF, ml/min; BPM, Hz or beats/min; MSLVP and EDLVP, mm Hg – were averaged as monitored in the first 5 min of the standard protocols and normalized as 100%. Differences between each percentually transformed variable in control and treated preparations were assessed as averages of the subsequent 2 min kinetic steps using the t test (unpaired, two tailed) applied to each next repeated measurement. All data sets, averages, standard errors of the means (S.E.) and probabilities (P) for each parameter, control vs treated groups, were calculated using the Microsoft Excel 2000 and SPSS 13.0 statistical packages. Additional evaluation were performed with the SPSS 13.0 for Windows full package, and the HTM (Microsoft word editable) & PDF formats, as well as the proprietary SPSS.SPO (editable with SPSS) format. All data and evaluations are presented in the attached files; a few, selected final Figures are included into the Results section.

## 2.6. PRODUCTS

Capsaicin synthetic analog (N-vanillylnonamide RBI, Natick, MA, USA; m.w. 293.41, lot VPR-396A) was prepared 60x of the corresponding measured CF final 10  $\mu$ mole, diluted in dimethyl sulfoxide (DMSO) (Merck & Co., Inc., Rahway, NJ, USA), final 0.05%, maintained under nitrogen. All other chemicals were Sigma Chem. Co. (St. Louis, MI, USA) reagents.

## 3. RESULTS

The means and S.E. of the absolute values of the four functional parameters assumed as independent in the 41 hearts of the 6 groups are presented in Table I. Figures 1 - 4 show the percentually normalized, t test evaluated, related time courses, means and S.E. of control vs capsaicin pretreated SP, LP and PC groups (1<sup>st</sup> attachment). Figures 5 and 6 show the time courses of the means of the two calculated, dependent parameters from same attachment.

Figure 7 reports the specific kinetics of the single measurable parameter of the 30 min ischemia applied to all 41 hearts.

Table 1

Absolute values of the 4 functional parameters measured in each rat taken as the average of the first 5 min of *ex vivo* KH Langendorff perfusion. Means and S.E. of the 6 groups and P values of the t test (unpaired, two tailed) between control and capsaicin-treated specimens subjected to preconditioning (PC), long perfusion (LP) or short perfusion (SP).

n total	Groups	Coronary Flow (CF; ml/min)				Frequency (BPM; Hz)				Maximum Systolic Pressure (MSLVP; mmHg)				End Diastolic Pressure (EDLVP; mmHg)			
		n	Control	n	Treated	n	Control	n	Treated	n	Control	n	Treated	n	Control	n	Treated
	Preconditioned (PC)																
2	2	1	16.3	1	19.4	1	245.8	1	246.	1	134.7	1	98.0	1	5.6	1	7.7
4	4	2	16.3	2	21.6	2	237.	2	284.2	2	131.5	2	111.9	2	5.7	2	6.7
6	6	3	17.7	3	20.4	3	231.8	3	313.9	3	116.1	3	82.0	3	6.5	3	9.2
8	8	4	17.	4	18.	4	252.1	4	243.2	4	94.3	4	99.2	4	8.0	4	7.3
10	10	5	13.5	5	15.8	5	267.2	5	273.4	5	114.5	5	94.9	5	6.5	5	7.6
12	12	6	16.3	6	17.7	6	256.8	6	258.7	6	94.7	6	124.7	6	7.9	6	5.1
13	13	7	17.1			7	305.2			7	104.			7	7.2		
	Means		16.31		18.8		255.1		269.9		112.8		101.8		6.8		7.2
	S.E.		0.51		0.84		9.1		10.9		6.2		6.		0.4		1.2
	P		0.024				0.32				0.2				0.46		
	Long Perfusion (LP)																
15	2	1	12.5	1	19.	1	222.1	1	261.5	1	134.2	1	113.2	1	5.6	1	6.6
17	4	2	13.1	2	20.	2	252.1	2	272.8	2	141.6	2	91.8	2	5.3	2	8.3
19	6	3	13.2	3	15.	3	269.8	3	221.3	3	124.2	3	95.5	3	6.0	3	7.9
21	8	4	13.9	4	21.	4	184.7	4	268.3	4	85.6	4	102.9	4	8.8	4	11.0
23	10	5	13.8	5	14.7	5	238.3	5	222.	5	155.3	5	108.3	5	4.8	5	7.5
25	12	6	12.9	6	16.1	6	247.7	6	284.5	6	121.2	6	89.	6	6.2	6	10.1
26	13	7	12.6			7	250.4			7	88.6			7	8.5		
27	14	8	13.3			8	208.			8	106.5			8	6.9		
28	15	9	13.9			9	267.8			9	101.1			9	7.4		
	Means		13.2		17.6		255.1		255.1		117.8		100.1		6.6		8.5
	S.E.		0.2		1.11		11.		11.		7.95		3.92		0.5		0.7
	P		0.00034				0.26				0.11				0.05		
	Short Perfusion (LP)																
30	2	1	19.4	1	22.8	1	235.5	1	251.1	1	103.5	1	95.8	1	7.3	1	7.8
32	4	2	21.5	2	23.5	2	219.4	2	286	2	112.8	2	81.6	2	6.7	2	9.2
34	6	3	19.2	3	22.2	3	225.3	3	268.3	3	103.5	3	101.9	3	7.3	3	7.4
36	8	4	18.7	4	19.2	4	201.9	4	237.9	4	109.1	4	104.2	4	6.9	4	7.2
38	10	5	22.3	5	19.7	5	301.7	5	278.1	5	99.3	5	96.7	5	7.5	5	7.7
40	12	6	19.4	6	22.1	6	280.2	6	309.8	6	96.5	6	91.5	6	7.8	6	8.3
41	13	7	19.4			7	273.3			7	102.4			7	7.4		
	Means		19.99		21.58		248.2		271.9		103.9		95.3		7.3		7.9
	S.E.		0.51		0.71		14.		10.4		2.1		3.3		0.1		0.3
	P		0.089				0.21				0.04				0.05		

Figure 1

Coronary flow (CF; ml/min). Control (◆) vs capsaicin-pretreated (■) time courses of the means of the percent values and of their S.E. Data from 1° attachment.

X axis: consecutive measurement at 2 min intervals.

Y axis: means of the percent values and their S.E. (vertical bars).

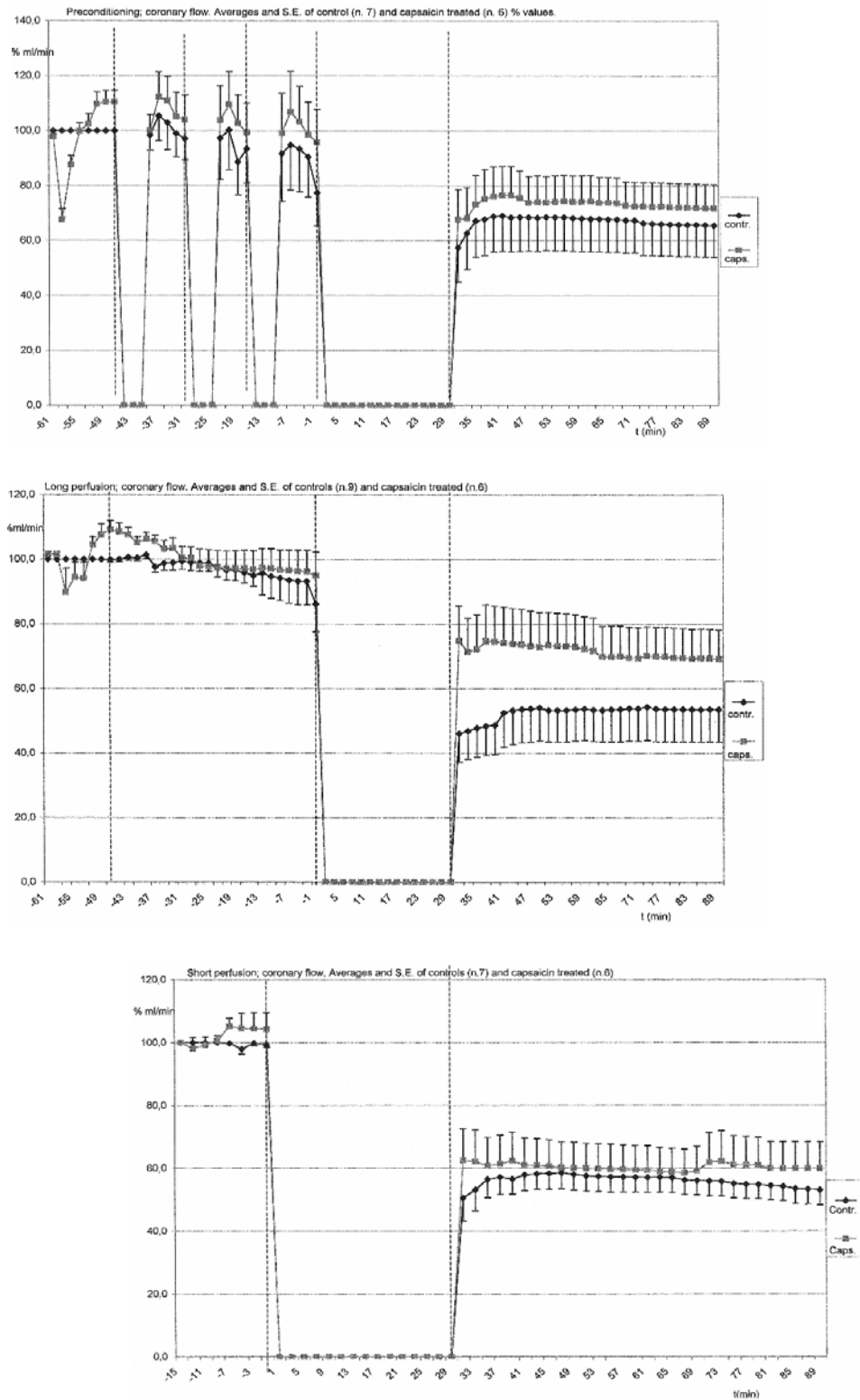




Figure 2

Frequency (BPM; Hz). Control (◆) vs capsaicin-pretreated (■) time courses of the means of the percent values and of their S.E. Data from 1° attachment.

X axis: consecutive measurement at 2 min intervals.

Y axis: means of the percent values and their S.E. (vertical bars).

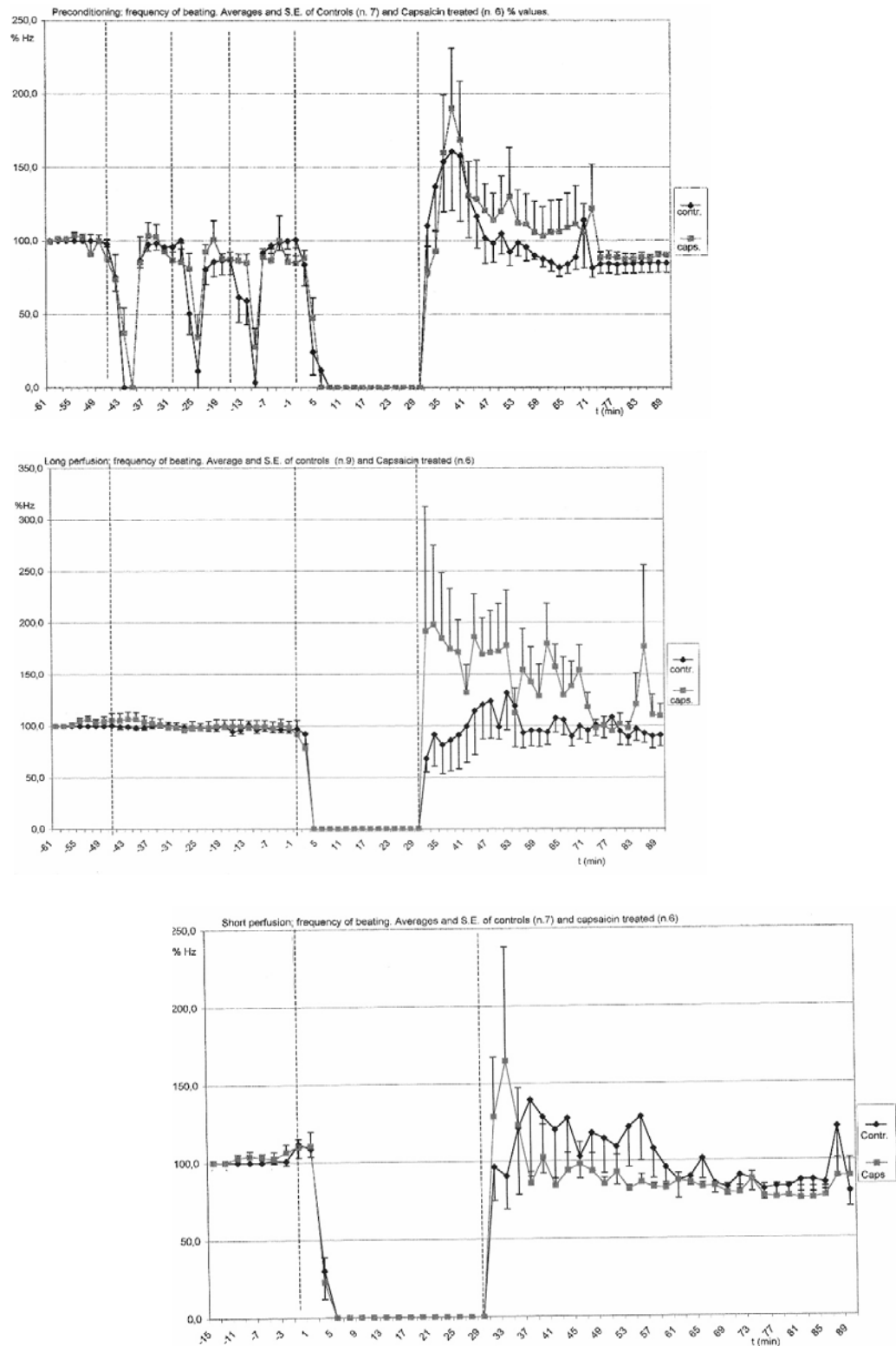


Figure 3

Maximum systolic left ventricular pressure (MSLVP; mmHg). Control (◆) vs capsaicin-pretreated (■) time courses of the means of the percent values and of their S.E. X axis: consecutive measurement at 2 min intervals. Y axis: means of the percent values and their S.E. (vertical bars).

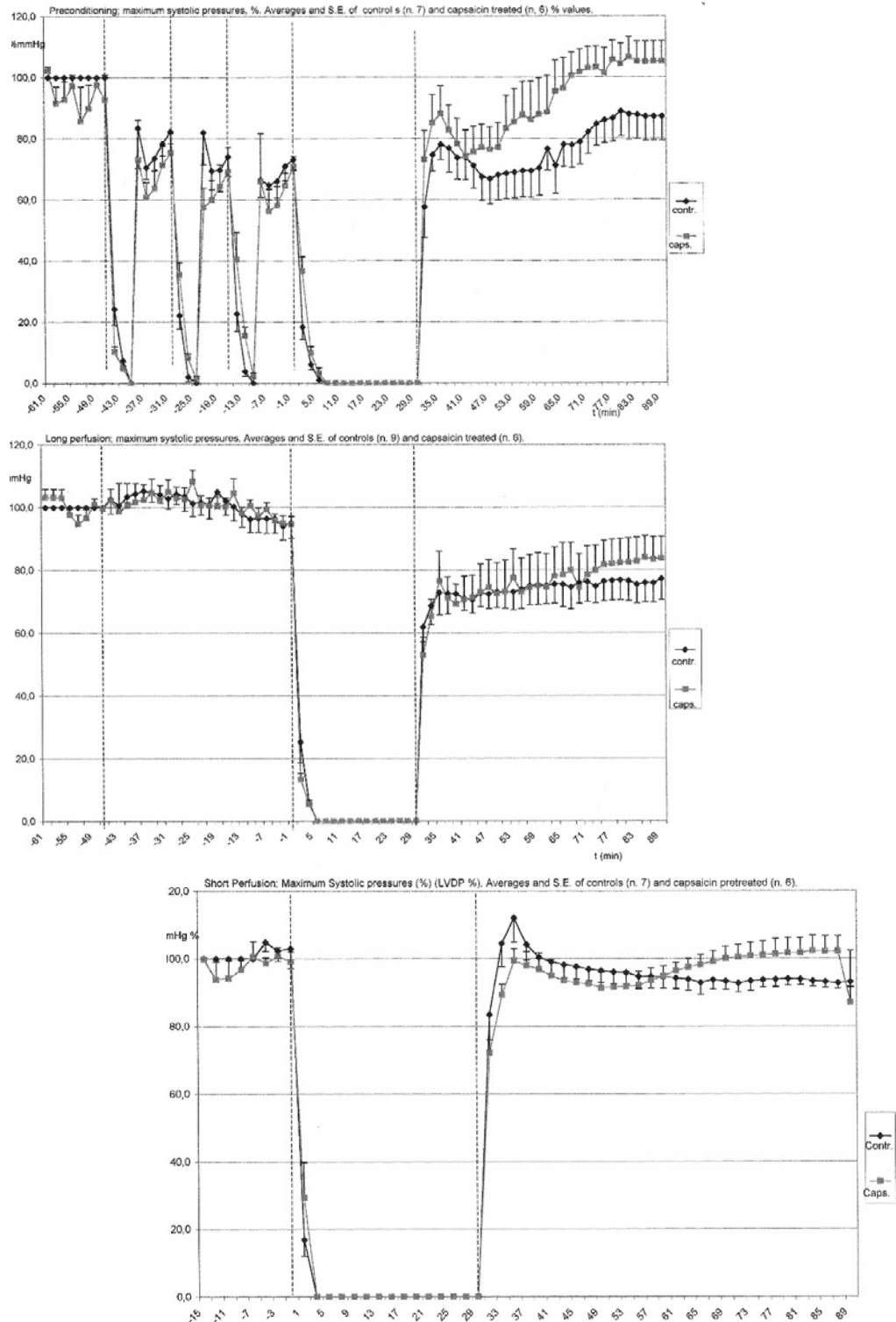


Figure 4

End diastolic left ventricular pressure (EDLVP; mmHg). Control (♦) vs capsaicin-pretreated (■) time courses of the means of the percent values and of their S.E. X axis: consecutive measurement at 2 min intervals. Y axis: means of the percent values and their S.E. (vertical bars).

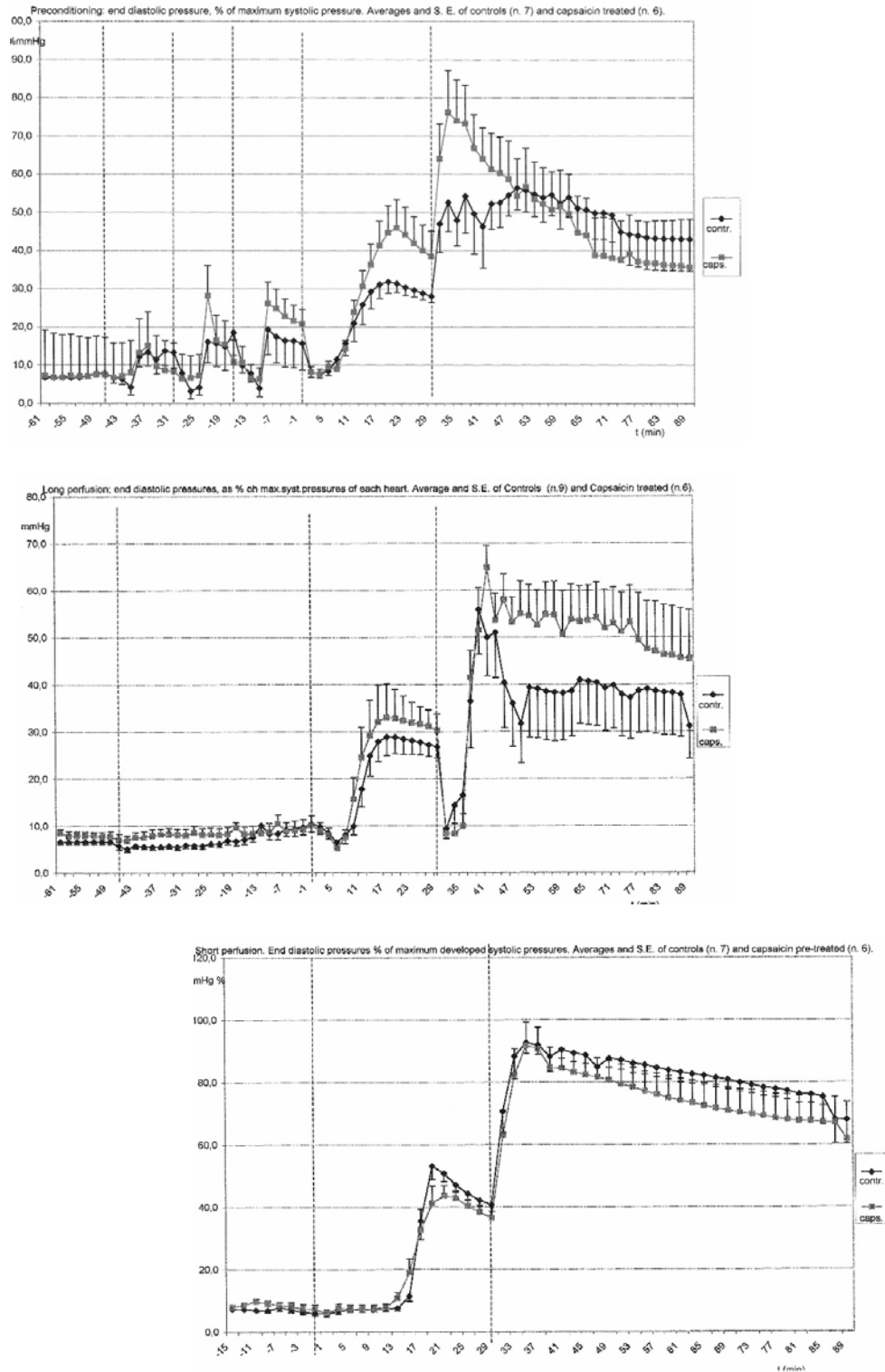
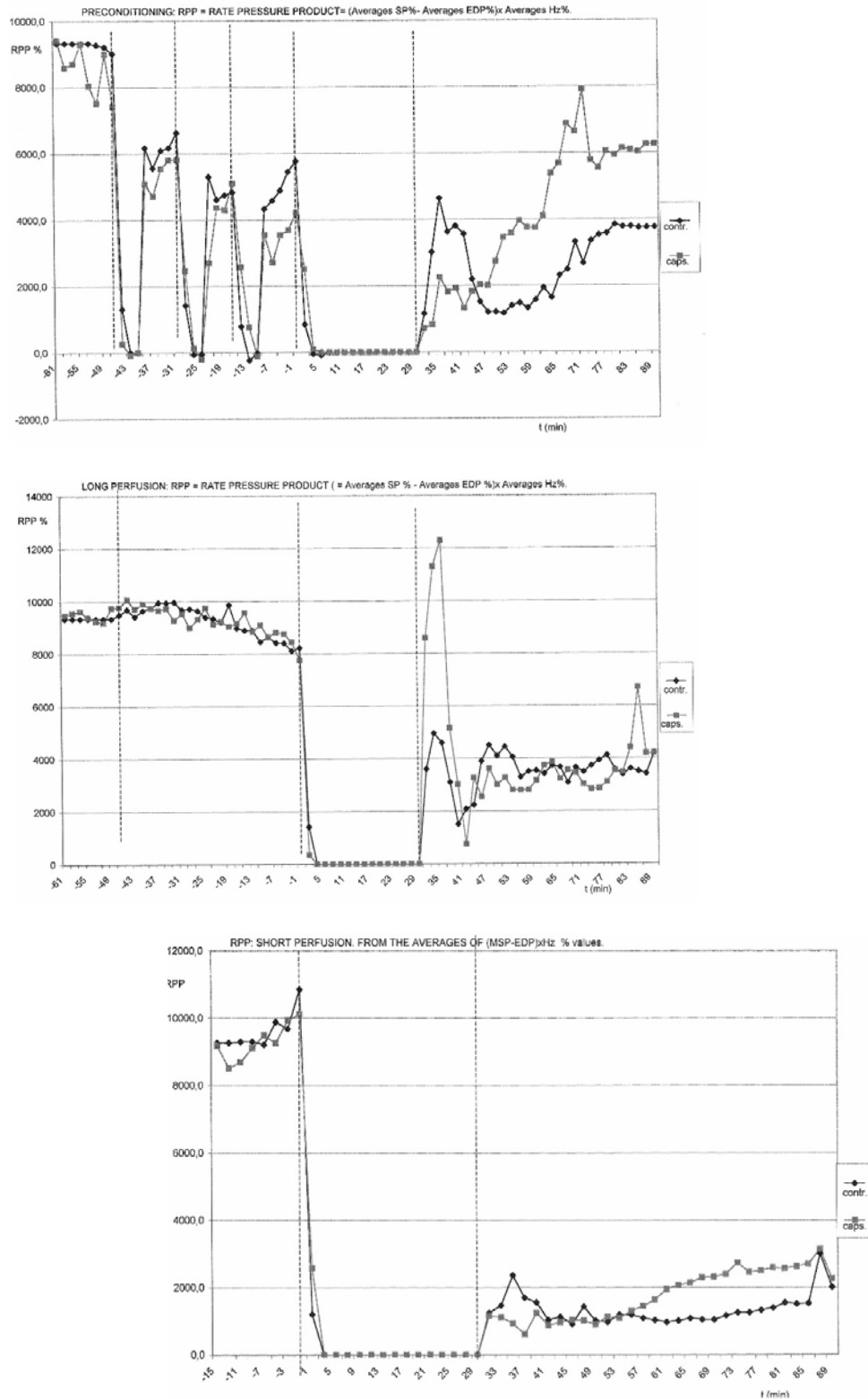


Figure 5

Rate pressure product (RPP). Control (◆) vs capsaicin-pretreated (■) time courses of the means of the percent values.

X axis: consecutive measurements at 2 min intervals.

Y axis: means of the percent values.



**Figure 6**

Rate pressure product divided by the coronary flow (RPP/CF). Control (◆) vs capsaicin-pretreated (■) time courses of the means of the percent values.  
 X axis: consecutive measurements at 2 min intervals.  
 Y axis: means of the percent values.

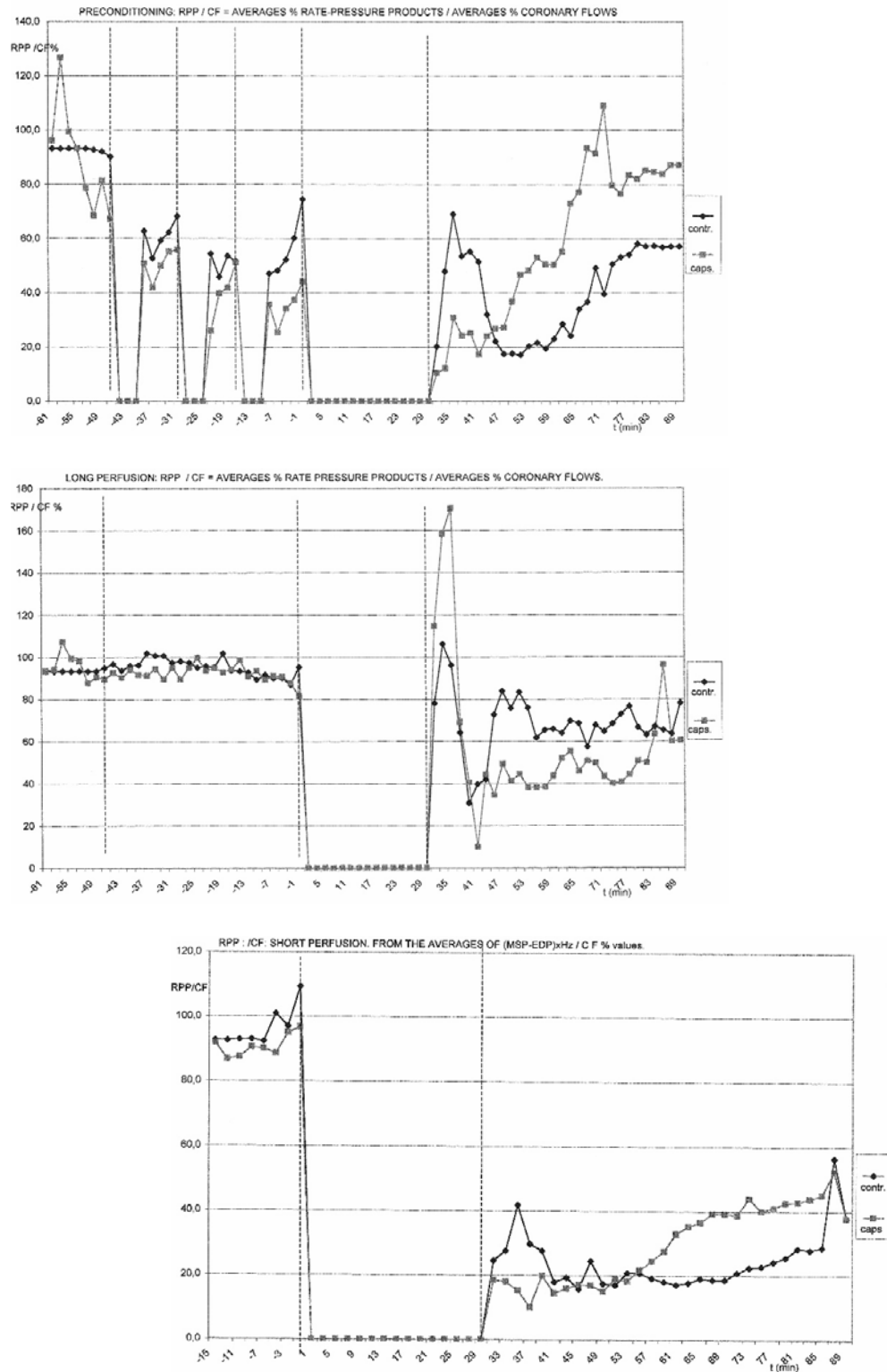
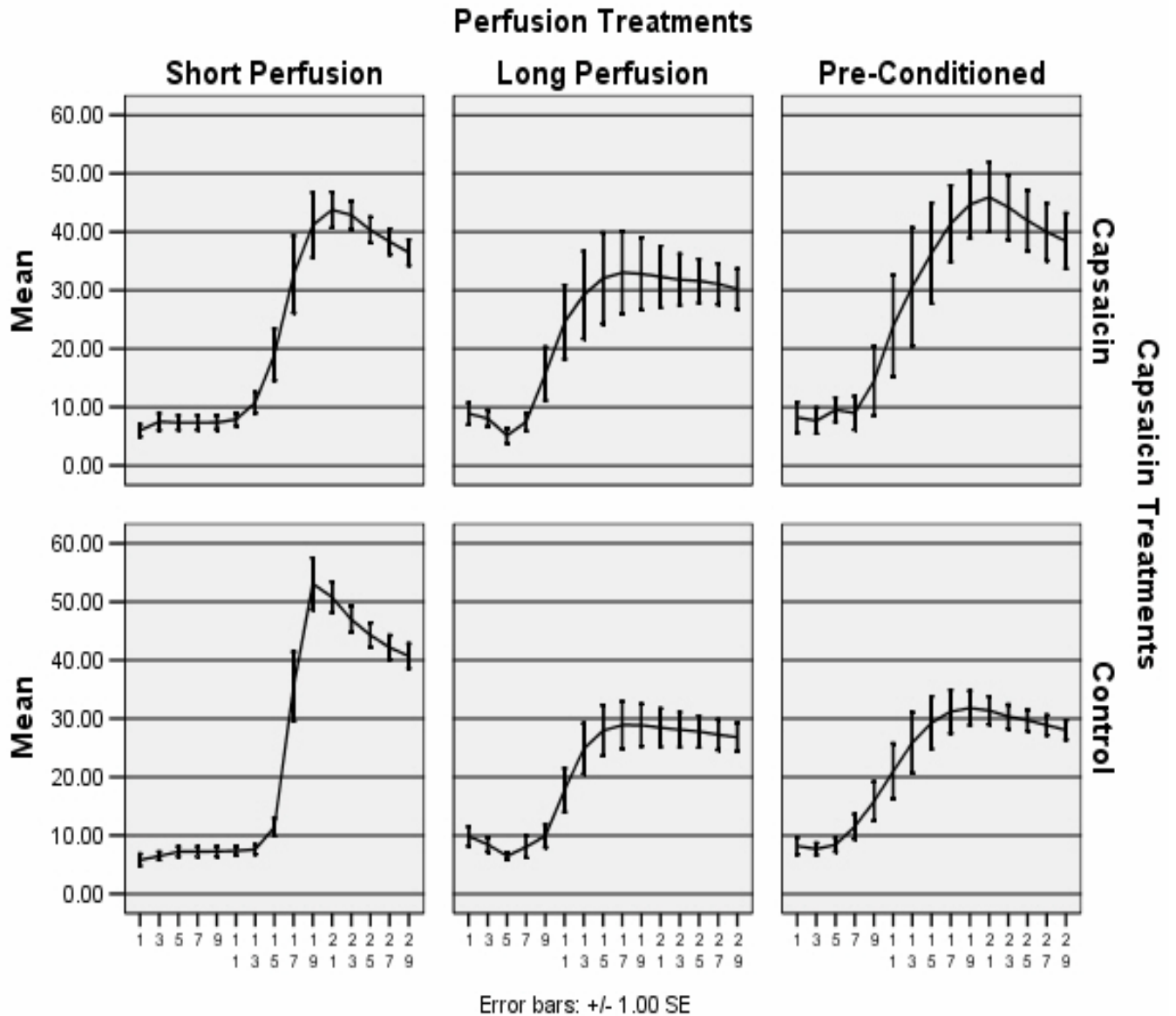


Figure 7

Time course of the sole measurable functional parameter (ischemic contracture) . Kinetic trends in the 6 experimental conditions.

X axis: consecutive measurements at 2 min intervals.

Y axis: means of the percent values and their S.E. (vertical bars).



### **ACKNOWLEDGEMENTS**

This work was financed by grants to R.D. and conducted using her Institute facilities, reagents, and housing; L.R. benefited from a “short-term” mobility visit financed by Centro Nazionale Ricerche (CNR), Italy, and by University of Ancona. Aligning data and getting ready the Tables of the 1<sup>st</sup> attachment and Figures was performed with Carlo Violet, Gerardo Galeazzi and Ivano Paglione’ expert technical assistance.

## REFERENCES

- [1] Argaud L, Gateau-Roesch O, Muntean D, Chalabreysse L, Loufouat J, Robert D, Ovize M. Specific inhibition of the mitochondrial permeability transition prevents lethal reperfusion injury. *J Mol Cell Cardiol* 2005; 38: 367-374.
- [2] Chen H, Liu LL, Ye LL, McGuckin C, Tamowski S, Scowen P, Tian H, Murray K, Hatton WJ, Duan D. Targeted inactivation of cystic fibrosis transmembrane conductance regulator chloride channel gene prevents ischemic preconditioning in isolated mouse heart. *Circulation* 2004; 110: 700-704.
- [3] Gottlieb RA. Debatable contribution of mitochondrial swelling to cell swelling in ischemia. *J Mol Cell Cardiol* 2003; 35: 735-737.
- [4] Diaz RJ, Batthish M, Backx PH, Wilson GJ. Chloride channel inhibition does block the protection of ischemic preconditioning in myocardium. *J Mol Cell Cardiol* 2001; 33: 1887-1889.
- [5] Heusch G, Liu GS, Rose J, Cohen MV, Downey JM. Swelling-activated chloride channels as effectors of ischemic preconditioning?. *J Mol Cell Cardiol* 2001; 33: 1891-1892.
- [6] Heusch G, Liu GS, Rose J, Cohen MV, Downey JM. No confirmation for a causal role of volume-regulated chloride channels in scheme preconditioning in rabbits. *J Mol Cell Cardiol* 2000; 32: 2279-2285.
- [7] Maier LS, Bers DM, Pieske B. Differences in Ca<sup>2+</sup>-handling and sarcoplasmic reticulum Ca<sup>2+</sup>-content in isolated rat and rabbit myocardium. *J Mol Cell Cardiol* 2000; 32: 2249-2258.
- [8] Fenton RA, Dickson EW, Mweyer TE, Dobson Jr JG. Aging reduces the cardioprotective effect of ischemic preconditioning in the rat heart. *J Mol Cell Cardiol* 2001; 32: 1371-1375.
- [9] Hilal-Dandan R, Kanter JR, Brunton LL. Characterization of G-protein signalling in ventricular myocytes from the adult mouse heart: differences from the rat. *J Mol Cell Cardiol* 2000; 32: 1211-1221.
- [10] Marber MS. Ischemic preconditioning in isolated cells. *Circ Res* 2000; 86: 926-931.
- [11] Lawrence C, Rodrigo GC. A Na<sup>+</sup>-activated K<sup>+</sup> current (I<sub>K,Na</sub>) is present in guinea pig but not rat ventricular myocytes. *Pflügers Archiv European J Physiol* 1999; 437: 831-838.
- [12] Brew EC, Mitchell MB, Rehring TF, Gamboni-Robertson F, McIntyre RC, Harken AH, Banerjee A. Role of bradykinin in cardiac



functional protection after global ischemia-reperfusion in rat heart. *Am J Physiol* 1995; 269: H1370-H1378.

[13] Kurz T, Tölg R, Richardt G. Bradykinin B2-receptor-mediated stimulation of exocytotic noradrenaline release from cardiac sympathetic neurons. *J Mol Cell Cardiol* 1997; 29: 2561-2569.

[14] Bugge E, Ytrehus K. Bradykinin protects against infarction but does not mediate ischemic preconditioning in the isolated rat heart. *J Mol Cell Cardiol* 1996; 28: 2333-2341.

[15] Bouchard J-F, Chouinard J, Lamontagne D. Role of kinins in the endothelium protective effect of ischemic preconditioning. *Br J Pharmacol* 1998; 123: 413-420.

[16] Weselcouch EO, Baird AJ, Sleph PG, Dzwonczyk S, Murray HN, Grover GJ. Endogenous catecholamines are not necessary for ischemic preconditioning in the isolated perfused rat heart. *Cardiovasc Res* 1995; 29: 126-132.

[17] Wang Y, Ashraf M. Activation of  $\alpha_1$ -adrenergic receptor during  $Ca^{2+}$  preconditioning elicits strong protection against  $Ca^{2+}$  overload injury via protein kinase C signaling pathway. *J Mol Cell Cardiol* 1998; 30: 2423-2435.

[18] Hearse DJ, Sutherland FJ. Catecholamines and preconditioning: studies of contraction and function in isolated rat hearts. *Am J Physiol* 1999; 277: H136-H143.

[19] Kolocassides KG, Galinanes M, Hearse DJ. Preconditioning accelerates contracture and ATP depletion in blood-perfused rat hearts. *Am J Physiol* 1995; 269: H1415-H1420.

[20] Kolocassides KG, Seymour A-M L, Galinanes M, Hearse DJ. Paradoxical effect of ischemic preconditioning on ischemic contracture ? NMR studies of energy metabolism and intracellular pH in the rat heart. *J Mol Cell Cardiol* 1996; 28: 1045-1057.

[21] Kolocassides KG, Galinanes M, Hearse DJ. Ischemic preconditioning, cardioplegia or both? Differing approaches to myocardial and vascular protection. *J Mol Cell Cardiol* 1996; 28: 623-634.

[22] Hearse DJ, Sutherland FJ. Ischemic preconditioning and exacerbation of contracture: does this occur with other preconditioning stimuli ?. XVIII European Congress International Society for Heart Research, Bologna, 18-21 June 1996.

[23] Hearse DJ, Ferrari R, Sutherland FJ. Cardioprotection: intermittent ventricular fibrillation and rapid pacing can induce

preconditioning in the blood-perfused rat heart. *J Mol Cell Cardiol* 1999; 31: 1961-1973.

[24] Thornton JD, Liu GS, Downey JM. Pretreatment with pertussis toxin blocks the protective effects of preconditioning: Evidence for a G-protein mechanism. *J Mol Cell Cardiol* 1993; 25: 311-320.

[25] Bohm M, Gierschik P, Schwinger RHG, Uhlmann R, Erdmann E. Coupling of M-cholinoceptors and A1 adenosine receptors in human myocardium. *Am J Physiol* 1994; 266: H1951-H1958.

[26] Tahir H, Mustafa SJ. Regulation of G proteins by adenosine receptor agonist in coronary artery. *Am J Physiol* 1994; 266: H1273-H1279.

[27] Cave AC, Collis CS, Downey JM, Hearse DJ. Improved functional recovery by ischemic preconditioning is not mediated by adenosine in the globally ischemic isolated rat heart. *Cardiov Res* 1993; 27: 663-668.

[28] Richardt G, Blessing R, Schömig A. Cardiac noradrenaline release accelerates adenosine formation in the ischemic rat heart: Role of neuronal noradrenaline carrier and adrenergic receptors. *J Mol Cell Cardiol* 1994; 26: 1321-1328.

[29] Headrick JP. Ischemic preconditioning: Bioenergetic and metabolic changes and the role of endogenous adenosine. *J Mol Cell Cardiol* 1996; 28: 1227-1240.

[30] Tani M, Sukanuma Y, Takayama M, Hasegawa H, Shinmura K, Ebihara Y, Tamaki K. Low concentrations of adenosine receptor blocker decrease protection by hypoxic preconditioning in ischemic rat hearts. *J Mol Cell Cardiol* 1998; 30: 617-620.

[31] Perlini S, Khoury EP, Norton GR, Chung ES, Fenton RA, Dobson JG, Meyer TE. Adenosine mediates sustained adrenergic desensitization in the rat heart via activation of protein kinase C. *Circ Res* 1998; 83: 761-771.

[32] Guo Y, Bolli R, Bao W, Wu WJ, Black Jr RG, Murphree SS, Salvatore CA, Jacobson MA, Auchampach JA. Targeted deletion of the A<sub>3</sub> adenosine receptor confers resistance to myocardial ischemic injury and does not prevent early preconditioning. *J Mol Cell Cardiol* 2001; 33: 825-830.

[33] Ungerer M, Chlistalla A, Richardt G. Upregulation of cardiac uptake1 carrier in ischemic and nonischemic rat heart. *Circ Res* 1996; 78: 1037-1043.

- [34] Li Y, Kloner RA. Does protein kinase C play a role in ischemic preconditioning in rat hearts ?. *Am J Physiol* 1995; 268: H426-H431.
- [35] Lundmark JL, Ramasamy R, Vulliet PR, Schaefer S. Chelerythrine increases Na-K-ATPase activity and limits ischemic injury in isolated rat hearts. *Am J Physiol* 1999; 277: H999-H1006.
- [36] Wang Y, Hirai K, Ashraf M. Activation of mitochondrial ATP-sensitive K<sup>+</sup> channel for cardiac protection against ischemic injury is dependent on protein kinase C activity. *Circ Res* 1999; 85: 731-741.
- [37] Asimakis GK, Inners-McBride K, Medellin G, Conti VR. Ischemic preconditioning attenuates acidosis and postischemic dysfunction in isolated rat heart. *Am J Physiol* 1992; 263: H887-H894.
- [38] Chen W, Wetsel W, Steenbergen C, Murphy E. Effect of ischemic preconditioning and PKC activation on acidification during ischemia in rat heart. *J Mol Cell Cardiol* 1996; 28: 871-880.
- [39] Cross HR, Murphy E, Bolli R, Ping P, Steenbergen C. Expression of activated PKC Epsilon (PKC $\epsilon$ ) protects the ischemic heart, without attenuating ischemic H<sup>+</sup> production. *J Mol Cell Cardiol* 2002; 34:361-367.
- [40] Chen L, Hahn H, Wu G, Chen C-H, Liron T, Schechtman D, Cavallaro G, Banci L, Guo Y, Bolli R, Dorn II GW, Mochly-Rosen D. Opposing cardioprotective actions and parallel hypertrophic effects of  $\delta$ PKC and  $\epsilon$ PKC. *Proc Natl Acad Sci* 2001; 98 : 11114-11119.
- [41] Das DK. Protein Kinase C isozymes signaling in the heart. *J Mol Cell Cardiol* 2003; 35: 887-889.
- [42] McCarthy J, McLeod CJ, Minners J, Faadiel Essop M, Ping P, Sack MN. PKC $\epsilon$  activation augments cardiac mitochondrial respiratory post-anoxic reserve. A putative mechanism in PKC $\epsilon$  cardioprotection. *J Mol Cell Cardiol* 2005; 38: 697-700.
- [43] Kang MK, Walker JW. Protein kinase C  $\delta$  and  $\epsilon$  mediate positive inotropy in adult ventricular myocytes. *J Mol Cell Cardiol* 2005; 38: 753-764.
- [44] Jin ZQ, Goetzi EJ, Karliner JS. Sphingosine kinase activation mediates ischemic preconditioning in murine heart. *Circulation* 2004; 110: 1980-1989.
- [45] Sanada S, Asanuma H, Tsukamoto O, Minamino T, Node K, Tashima S, Fukushima TT, Ogai A, Shinozaki Y, Fujita M, Hirata A, Okuda H, Shimokawas H, Tonmoike H, Hori M, Kitazake M. Protein kinase A as another mediator of ischemic preconditioning independent of protein kinase C. *Circulation* 2004; 119: 51-57.

[46] Hahn HS, Yussman MG, Toyokawa T, Marreez Y, Barrett TJ, Hilty KC, Osinska H, Robbins J, Dorn II GW. Ischemic protection and myofibrillar cardiomyopathy. Dose-dependent effects of *in vivo*  $\delta$ PKC inhibition. *Circ Res* 2002; 94: 741-748.

[47] Liu GS, Cohen MV, Mochly-Rosen D, Downey JM. Protein kinase C- $\epsilon$  is responsible for the protection of preconditioning in rabbit cardiomyocytes. *J Mol Cell Cardiol* 1999; 31: 1937-1948.

[48] Gan XT, Chakrabarti S, Karmazyn M. Modulation of  $\text{Na}^+/\text{H}^+$  exchange isoform-1 mRNA expression in isolated rat hearts. *Am J Physiol* 1999; 277: H993-H998.

[49] Bugge E, Ytrehus K. Inhibition of sodium-hydrogen exchange reduces infarct size in the isolated rat heart - a protective additive to ischemic preconditioning. *Cardiov Res* 1995; 29: 269-274.

[50] Xiao XH, Allen DG. Role of  $\text{Na}^+/\text{H}^+$  exchanger during ischemia and preconditioning in the isolated rat heart. *Circ Res* 1999; 85: 723-730.

[51] Karmazyn M, Gan XT, Humphreys RA, Yoshida H, Kusumoto K. The myocardial  $\text{Na}^+/\text{H}^+$  exchange. Structure, regulation, and its role in heart disease. *Circ Res* 1999; 85: 777-786.

[52] Gabel SA, Cross HR, London RE, Steenbergen C, Murphy E. Decreased intracellular pH is not due to increased  $\text{H}^+$  extrusion in preconditioned rat hearts. *Am J Physiol* 1997; 273: H2257-H2262.

[53] Nayler WG, Perry SE, Elz JS, Daly MJ. Calcium, sodium, and the calcium paradox. *Circ Res* 1984; 55: 227-237.4

[54] Grinwald PM, Brosnahan C. Sodium imbalance as a cause of calcium overload in post-hypoxic reoxygenation injury. *J Mol Cell Cardiol* 1987; 19: 487-495.

[55] Tani M, Neely R. Role of intracellular  $\text{Na}^+$  in  $\text{Ca}^{2+}$  overload and depressed recovery of ventricular function of reperfused ischemic rat hearts. *Circ Res* 1989; 65: 1045-1056.

[56] Imahashi K, Kusuoka H, Hashimoto K, Yoshioka J, Yamaguchi H, Nishimura T. Intracellular sodium accumulation during ischemia as the substrate for reperfusion injury. *Circ Res* 1999; 84: 1401-1406.

[57] Murphy E, Cross H, Steenbergen C. Sodium regulation during ischemia versus reperfusion and its role injury. *Circ Res* 1999; 84: 1469-1470.

[58] Humphreys RA, Haist JV, Chakrabarti S, Feng Q, Malcolm J, Arnold O, Karmazyn M. Orally administered NHE1 inhibitor cariporide reduces acute responses to coronary occlusion and reperfusion. *Am J Physiol* 1999; 276: H749-H757.

[59] Hartmann M, Decking UKM. Blocking  $\text{Na}^+\text{-H}^+$  exchange by cariporide reduces  $\text{Na}^+$ -overload in ischemia and is cardioprotective. *J Mol Cell Cardiol* 1999; 31: 1985-1995.

[60] Ladilov Y, Haffner S, Balsler-Schafer C, Maxeiner H, Piper HM. Cardioprotective effects of KB-R7943: a novel inhibitor of the reverse mode of  $\text{Na}^+/\text{Ca}^{2+}$  exchanger. *Am J Physiol* 1999; 276: H1868-H1876.

[61] Sawyer D, Suter TM, Apstein CS. The sting of salt on an old, but open, wound-is  $\text{Na}^+$  the cause of mitochondrial and myocardial injury during ischemia/reperfusion?. *J Mol Cell Cardiol* 2002; 34: 699-702.

[62] Iwai T, Tanonaka K, Inoue R, Kasahara S, Kamo N, Takeo S. Mitochondrial damage during ischemia determines post-ischemic contractile dysfunction in perfused rat heart. *J Mol Cell Cardiol* 2002; 34: 725-738.

[63] Hudman D, Rainbow RD, Lawrence CL, Standen NB. The origin of calcium overload in rat cardiac myocytes following metabolic inhibition with 2,4-dinitrophenol. *J Mol Cell Cardiol* 2002; 34: 859-871.

[64] Rodrigo GC, Lawrence CL, Standen NB. Dinitrophenol pre-treatment of rat ventricular myocytes protects against damage by metabolic inhibition and reperfusion. *J Mol Cell Cardiol* 2002; 34: 555-569.

[65] Van Wagoner DR, Bond M. Reperfusion arrhythmias: new insights into the role of the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger. *J Mol Cell Cardiol* 2001; 33: 2071-2074.

[66] Ylitalo KV, Ala-Rami A, Llimatta EV, Peuhkurinen KJ, Hassinen IE. Intracellular free calcium and mitochondrial membrane potential in ischemia/reperfusion and preconditioning. *J Mol Cell Cardiol* 2000; 32: 1223-1238.

[67] Shintani Y, Node K, Asauna H, Sanada S, Takashima S, Asano Y, Liao Y, Fujita M, Hirata A, Shinozaki Y, Fukushima T, Nagamachi Y, Okuda H, Kim J, Tomoike H, Hori M, Kitakaze M. Opening of  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels is involved in ischemic preconditioning in canine hearts. *J Mol Cell Cardiol* 2004; 37: 1213-1218

[68] Chen W, Gabel S, Steenbergen C, Murphy E. A redox-based mechanism for cardioprotection induced by ischemic preconditioning in perfused rat heart. *Circ Res* 1995; 77: 424-429.

[69] Arad M, de Jong JW, de Jong R, Huizer T, Rabinowitz B. Preconditioning in globally ischemic isolated rat hearts: effect of function and metabolic indices of myocardial damage. *J Mol Cell Cardiol* 1996; 28: 2479-2490.

[70] Wolfe CL, Sievers RE, Visseren FLJ, Donnelly TJ. Loss of myocardial protection after preconditioning correlates with the time course of glycogen recovery within the preconditioned segment. *Circulation* 1993; 87: 881-892.

[71] Sargent CA, Dzwonczyk S, Sleph P, Wilde M, Grover GJ. Pyruvate increases threshold for preconditioning in globally ischemic rat hearts. *Am J Physiol* 1994; 267: H1403-H1409.

[72] Albuquerque CP, Gerstenblith G, Weiss RG. Myocardial buffering capacity in ischemia preconditioning rat hearts. *J Mol Cell Cardiol* 1995; 27: 777-781.

[73] Bradamante S, Piccinini F, Delu C, Janssen M, de Jong JW. NMR evaluation of changes in myocardial high energy metabolism produced by repeated short periods of ischemia. *Biochim Biophys Acta* 1995; 1243: 1-8.

[74] Cross HR, Clarke K, Opie LH, Radda GK. Is lactate-induced myocardial ischemic injury mediated by decreased pH or increased intracellular lactate?. *J Mol Cell Cardiol* 1995; 27: 1369-1381.

[75] Finegan BA, Lopaschuk GD, Gandhi M, Clanachan AS. Ischemic preconditioning inhibits glycolysis and proton production in isolated working rat hearts. *Am J Physiol* 1995; 269: H1767-H1775.

[76] Schaefer S, Carr LJ, Prussel E, Ramasamy R. Effects of glycogen depletion on ischemic injury in isolated rat hearts: insights into preconditioning. *Am J Physiol* 1995; 268: H935-H944.

[77] Schjøtt J, Bakøy OE, Jones RA, Southon T, Jynge P. Preconditioning by brief ischemic episodes in the isolated rat heart assessed by <sup>31</sup>P NMR spectroscopy: dissociation between metabolic and functional recovery? *Scand J Clin Lab Invest* 1995; 55: 67-78.

[78] Heide RSV, Delyani JA, Jennings RB, Reimer KA, Steenbergen C. Reducing lactate accumulation does not attenuate lethal ischemic injury in isolated perfused rat hearts. *Am J Physiol* 1996; 270: H38-H44.

- [79] De Jonge R, Bradamante S, Speleman L, de Jong JW. Carbohydrates and purines in underperfused hearts, protected by ischemic preconditioning. *J Mol Cell Cardiol* 1998; 30: 699-708.
- [80] Tosaki A, Maulik N, Elliott GT, Blasig IE, Engelman RM, Das DK. Preconditioning of rat heart with monophosphoryl lipid A: A role for nitric oxide. *J Pharmacol Exp Ther* 1998; 285: 1274-1279.
- [81] Weselcouch EO, Baird AJ, Sleph P, Grover GJ. Inhibition of nitric oxide synthesis does not affect ischemic preconditioning in isolated perfused rat hearts. *Am J Physiol* 1995; 268: H242-H249.
- [82] Benkusky NA, Lewis SJ, Kooy NW. Attenuation of vascular relaxation after development of tachyphylaxis to peroxynitrite. *Am J Physiol* 1998; 275: H501-H508.
- [83] Nossuli TO, Hayward R, Jensen D, Scalia R, Lefer AM. Mechanisms of cardioprotection by peroxynitrite in myocardial ischemia and reperfusion injury. *Am J Physiol* 1998; 275: H509-H519.
- [84] Wang Y, Guo Y, Zhang SX, Wu W-J, Wang J, Bao W, Bolli R. Ischemic preconditioning upregulates inducible nitric oxide synthase in cardiac myocytes. *J Mol Cell Cardiol* 2002; 34: 5-15.
- [85] Ping P, Zhang J, Pierce WM, Bolli R. Functional proteomic analysis of protein kinase C- $\epsilon$  signaling complexes in the normal heart and during cardioprotection. *Circ Res* 2001; 88: 59-62.
- [86] Li M, Nishimura H, Iwakura A, Wecker A, Eaton E, Asahara T, Losordo DW. Endothelial progenitor cells are rapidly recruited to myocardium and mediate protective effect of ischemic preconditioning via "imported" nitric oxide synthase activity. *Circulation* 2005; 111: 1114-1120.
- [87] Richard V, Tron C, Thuillez C. Ischemic preconditioning is not mediated by oxygen derived free radicals in rats. *Cardiov Res* 1993; 26: 1321-1328.
- [88] Das DK, Engelman RM, Dobbs WA, Rousou JA, Breyer RH. The role of oxygen-derived free radicals in pathogenesis of reperfusion injury. *Ann New York Acad Sci* 1986; 463: 274-277.
- [89] Das DK, Engelman RM, Maulik N. Oxygen free radical signaling in ischemic preconditioning. *Ann N Y Acad Sci* 1999; 874: 49-65.
- [90] Mekhfi H, Veksler V, Mateo P, Maupoil V, Rochette L, Ventura-Clapier R. Creatine kinase is the main target of reactive oxygen species in cardiac myofibrils. *Circ Res* 1996; 78: 1016-1027.

[91] Arstall MA, Bailey C, Gross WL, Bak M, Balligand J-L, Kelly RA. Reversible S-nitrosation of creatine kinase by nitric oxide in adult rat ventricular myocytes. *J Mol Cell Cardiol* 1998; 30: 979-988.

[92] Neubauer S, Remkes H, Spindler M, Horn M, Wiesmann F, Prestle J, Walzer B, Ertl G, Hasenfuss G, Walliman T. Downregulation of the Na<sup>+</sup>-creatine cotransporter in failing human myocardium and in experimental heart failure. *Circulation* 1999; 100: 1847-1850.

[93] Ponticos M, Lu QL, Morgan JE, Hardie DG, Partidge TA, Carling D. Dual regulation of the AMP-activated protein kinase provides a novel mechanism for the control of creatine kinase in skeletal muscle. *EMBO J* 1998; 17: 1688-1699.

[94] Green DW, Murray HN, Sleph PG, Wang F-L, Baird AJ, Rogers WL, Grover GJ. Preconditioning in rat hearts is independent of mitochondrial F<sub>1</sub>F<sub>0</sub> ATPase inhibition. *Am J Physiol* 1998; 274: H90-H97.

[95] Vuorinen KK, Ylitalo K, Peuhkurinen K, Raatikainen P, Ala-Rami A, Hassinen IE. Mechanisms of ischemic preconditioning in rat myocardium. Roles of adenosine, cellular energy state and mitochondrial F<sub>1</sub>F<sub>0</sub>-ATPase. *Circulation* 1995; 91: 2810-2818.

[96] Vander Heide RS, Hill ML, Reimer KA, Jennings RB. Effect of reversible ischemia on the activity of the mitochondrial ATPase: relationship to ischemic preconditioning. *J Mol Cell Cardiol* 1996; 28: 103-112.

[97] Zucchi R, Yu G, Galbani P, Mariani M, Ronca G, Ronca-Testoni S. Sulfhydryl redox state affects susceptibility to ischemia and sarcoplasmic reticulum Ca<sup>2+</sup> release in rat heart. Implications for ischemic preconditioning. *Circ Res* 1998; 83: 908-915.

[98] Connaughton M, Kelly FJ, Haddock PS, Hearse DJ, Shattock MJ. Ventricular arrhythmias induced by ischaemia-reperfusion are unaffected by myocardial glutathione depletion. *J Mol Cell Cardiol* 1996; 28: 679-688.

[99] Ruuge EK, Ledenev AN, Lakomkin VL, Konstantinov AA, Ksenzenko MY. Free radical metabolites in myocardium during ischemia and reperfusion. *Am J Physiol* 1991; 261: 81-86.

[100] Yabe K-I, Nasa Y, Sato M, Iijima R, Takeo S. Preconditioning preserves mitochondrial function and glycolytic flux during an early period of reperfusion in perfused rat hearts. *Cardiov Res* 1997; 33: 677-685.



[101] Osada M, Netticadan T, Tamura K, Dhalla NS. Modification of ischemia-reperfusion-induced changes in cardiac sarcoplasmic reticulum by preconditioning. *Am J Physiol* 1998; 274: H2025-H2034.

[102] Brandes R, Maier LS, Bers DM. Regulation of mitochondrial [NADH] by cytosolic  $[Ca^{2+}]$  and work in trabeculae from hypertrophic and normal rat hearts. *Circ Res* 1998; 82: 1189-1198.

[103] Zucchi R, Ronca-Testoni S, Yu G, Galbani P, Ronca G, Mariani M. Postischemic changes in cardiac sarcoplasmic reticulum  $Ca^{2+}$  channels. *Circ Res* 1995; 76: 1049-1056.

[104] Rauch U, Schulze K, Witzendichler B, Schultheiss HP. Alteration of the cytosolic-mitochondrial distribution of high-energy phosphates during global myocardial ischemia may contribute to early contractile failure. *Circ Res* 1994; 75: 760-769.

[105] Garnier A, Rossi A, Lavanchy N. Importance of the early alterations of energy metabolism in the induction and the disappearance of ischemic preconditioning in the isolated rat heart. *J Mol Cell Cardiol* 1996; 28: 1671-1682.

[106] Rossi A, Kay L, Saks V. Early ischemia-induced alterations of the outer mitochondrial membrane and the intermembrane space: A potential cause for altered energy transfer in cardiac muscle?. *Mol Cell Biochem* 1994; 184: 401-408.

[107] Novel-Chaté V, Mateo P, Saks VA, Hoerter JA, Rossi A. Chronic exposure of rats to hypoxic environment alters the mechanism of energy transfer in myocardium. *J Mol Cell Cardiol* 1998; 30: 1295-1303.

[108] Whittaker P, Kloner RA, Przyklenk K. Intramyocardial injections and protection against myocardial ischemia. An attempt to examine the cardioprotective actions of adenosine. *Circulation* 1996; 93: 2043-2051.

[109] Krenz M, Baines CP, Heusch G, Downey JM, Cohen MV. Acute alcohol-induced protection against infarction in rabbit hearts: differences from and similarities to ischemic preconditioning. *J Mol Cell Cardiol* 2001; 33: 2015-2022.

[110] Belhomme D, Peynet J, Louzy M, Launay JM, Kitakaze M, Menashe' P. Evidence for preconditioning by isoflurane in coronary artery bypass graft surgery. *Circulation* 1999; 100: 340-344.

[111] Kukreja R, Ockaili R, Salloum F, Yin C, Hawkins J, Das A, Xi L. Cardioprotection with phosphodiesterase-5 inhibition. A novel preconditioning strategy. *J Mol Cell Cardiol* 2004; 36: 165-173.

[112] Broadhead MW, Kharbanda RK, Peters MJ, MacAllister RJ.  $K_{ATP}$  channel activation induces ischemic preconditioning of the endothelium in humans in vivo. *Circulation* 2004; 110: 2077-2082.

[113] Gross GJ. Sildenafil and endothelial dysfunction in humans. *Circulation* 2005; 111: 721-723.

[114] Van Lambalgen AA, Van Kraats AA, Mulder MF, Teerlink T, Van Den Bos GC. High-energy phosphates in heart, liver, kidney, and skeletal muscle of endotoxemic rats. *Am J Physiol* 1994; 266: H1581-H1587.

[115] Chong K-Y, Lai C-C, Lille S, Chang C, Su C-Y. Stable overexpression of the constitutive form of heat shock protein 70 confers oxidative protection. *J Mol Cell Cardiol* 1998; 30: 599-608.

[116] Cornelussen RN, Garnier AV, van Der Vusse GJ, Reneman RS, Snoeckx LHEH. Biphasic effect of heat stress pretreatment on ischemic tolerance of isolated rat hearts. *J Mol Cell Cardiol* 1998; 30: 365-372.

[117] Joyeux M, Godin-Ribuot D, Ribuot C. Resistance to myocardial infarction induced by heat stress and the effect of ATP-sensitive potassium channel blockade in the rat isolated heart. *Br J Pharmacol* 1998; 123: 1085-1088.

[118] Su C-Y, Chong K-Y, Owen OE, Dillmann WH, Chang C, Lai C-C. Constitutive and inducible hsp70s are involved in oxidative resistance evoked by heat shock or ethanol. *J Mol Cell Cardiol* 1998; 30: 587-598.

[119] Hamada K, Yamazaki J, Nagao T. Shortening of action potential duration is not prerequisite for cardiac protection by ischemic preconditioning or a  $K_{ATP}$  channel opener. *J Mol Cell Cardiol* 1998; 30: 1369-1379.

[120] Docherty JC, Gunter HE, Kuzio B, Shoemaker L, Yang L, Deslauriers R. Effects of cromakalim and glibenclamide on myocardial high energy phosphates and intracellular pH during ischemia-reperfusion:  $^{31}P$  NMR studies. *J Mol Cell Cardiol* 1997; 29: 1665-1673.

[121] Gross JG. Recombinant cardiac ATP-sensitive potassium channels and cardioprotection. *Circulation* 1998; 98: 1479-1480.

[122] Jilkina O, Kuzio B, Grover GJ, Kupriyanov VV. Effects of  $K_{ATP}$  channel openers P-1075, pinacidil, and diazoxide, on energetics and contractile function in isolated rat hearts. *J Mol Cell Cardiol* 2002; 34: 427-440.

[123] Schultz JEJ, Kwok WM, Hsu AK, Gross GJ. Terikalant, an inward-rectifier potassium channel blocker, does not abolish the cardioprotection induced by ischemic preconditioning in the rat. *J Mol Cell Cardiol* 1998; 30: 1817-1825.

[124] Meldrum DR, Cain BS, Meng XZ, Cleveland JC, Shames BD, Donnahoo KK, Banerjee A, Harken AH. Calcium preconditioning, but not ischemic preconditioning, bypasses the adenosine triphosphate-dependent potassium ( $K_{ATP}$ ) channel. *J Surg Res* 1999; 85: 77-82.

[125] Inoue I, Nagase H, Kishi K, Higuti T. ATP-sensitive  $K^+$  channel in the mitochondrial inner membrane. *Nature* 1991; 352: 244-247.

[126] Garlid KD, Paucek P, Yarov-Yarovoy V, Murray HN, Darbenzio RB, D'Alonzo AJ, Lodge NJ, Smith MA, Grover GJ. Cardioprotective effect of diazoxide and its interaction with mitochondrial ATP-sensitive  $K^+$  channels. Possible mechanism of cardioprotection. *Circ Res* 1997; 81: 1072-1082.

[127] Liu Y, Sato T, O'Rourke B, Marban E. Mitochondrial ATP-dependent potassium channels. Novel effectors of cardioprotection?. *Circulation* 1998; 97: 2463-2469.

[128] Oldenburg O, Cohen MV, Downey JM. Mitochondrial  $K_{ATP}$  channels in preconditioning. *J Mol Cell Cardiol* 2003; 35: 569-575.

[129] Liu Y, Sato T, Seharaseyon J, Szewczyk A, O'Rourke B, Marban E. Mitochondrial ATP-dependent potassium channels. Viable candidate effectors of ischemic preconditioning. *Ann N Y Acad Sci* 1999; 874: 27-37.

[130] Schwarz P, Diem R, Dun NJ, Forstermann U. Endogenous and exogenous nitric oxide inhibits norepinephrine release from rat heart sympathetic nerves. *Circ Res* 1995; 77: 841-848.

[131] Tominaga M, Horie M, Sasayama S, Okada Y. Glibenclamide, an ATP-sensitive  $K^+$  channel blocker, inhibits cardiac cAMP-activated  $Cl^-$  conductance. *Circ Res* 1995; 77: 417-423.

[132] Musters RJP, van der Meulen ET, van der Laarse WJ, van Hardeveld C. Differential effects of norepinephrine on contractile recovery of rat trabeculae following metabolic inhibition. *J Mol Cell Cardiol* 1998; 30: 435-440.

[133] Linden J. Cloned adenosine A3 receptors: pharmacological properties, species differences and receptor functions. *Trends Pharmacol Sci* 1994; 15: 298-306.

[134] Gan XT, Cook MA, Moffat MP, Karmazyn M. Transient ischemia in the presence of an adenosine deaminase plus a

nucleotide transport inhibitor confers protection against contractile depression produced by hydrogen peroxide. Possible role of glycogen. *J Mol Cell Cardiol* 1996; 28:1165-1176.

[135] Hill RJ, Oleynek JJ, Magee W, Knight DR, Tracey WR. Relative importance of adenosine A1 and A3 receptors in mediating physiological or pharmacological protection from ischemic myocardial injury in the rabbit heart. *J Mol Cell Cardiol* 1998; 30: 579-585.

[136] Jacobson KA. Adenosine A2 receptors: novel ligands and paradoxical effects. *Trends Pharmacol Sci* 1998; 19: 184-191.

[137] Schultz JEJ, Rose E, Yao Z, Gross GJ. Evidence for involvement of opioid receptors in ischemic preconditioning in rat hearts. *Am J Physiol* 1995; 268: H2157- H2161.

[138] Schultz JEJ, Hsu AK, Gross GJ. Morphine mimics the cardioprotective effect of ischemic preconditioning via a glibenclamide-sensitive mechanism in rat heart. *Circ Res* 1996; 78: 1100-1104.

[139] Schultz JEJ, Hsu AK, Barbieri JT, Li PL, Gross GJ. Pertussis toxin abolishes the cardioprotective effect of ischemic preconditioning in intact rat heart. *Am J Physiol* 1998; 275: H495-H500.

[140] Yu XC, Li HY, Wang HX, Wong TM. U50,488H inhibits effects of norepinephrine in rat cardiomyocytes- cross-talk between  $\kappa$ -opioid and  $\beta$ -adrenergic receptors. *J Mol Cell Cardiol* 1998; 30: 405-413.

[141] Liang BT, Gross GJ. Direct preconditioning of cardiac myocytes via opioid receptors and KATP channels. *Circ Res* 1999; 84: 1396-1400.

[142] Wu S, Li HY, Wong TM. Cardioprotection of preconditioning by metabolic inhibition in the rat ventricular myocyte: involvement of  $\kappa$ -opioid receptor. *Circ Res* 1999; 84: 1388-1395.

[143] Gross GJ. Role of opioids in acute and delayed preconditioning. *J Mol Cell Cardiol* 2003; 35: 709-718.

[144] Schultz JEJ, Hsu AK, Gross GJ. Ischemic preconditioning in the intact rat heart is mediated by  $\delta$ 1- but not  $\mu$ - or  $\kappa$ -opioid receptors. *Circulation* 1998; 97: 1282-1289.

[145] Rice PJ, Armstrong SC, Ganote CE. Concentration-response relationships for adenosine agonists during preconditioning of rabbit cardiomyocytes. *J Mol Cell Cardiol* 1996; 28: 1355-1365.

[146] Hoek TLV, Shao Z, Li C, Schumacker PT, Becker LB. Mitochondrial electron transport can become a significant source of

oxidative injury in cardiomyocytes. *J Mol Cell Cardiol* 1997; 29: 2441-2450.

[147] Hoek TLV, Li C, Shao Z, Schumacker PT, Becker LB. Significant levels of oxidants are generated by isolated cardiomyocytes during ischemia prior to reperfusion. *J Mol Cell Cardiol* 1997; 29: 2571-2583.

[148] Hoek TLV, Becker LB, Shao Z, Li C, Schumacker PT. Reactive oxygen species released from mitochondria during brief hypoxia induce preconditioning in cardiomyocytes. *J Biol Chem* 1998; 273: 18092-18096.

[149] Mocanu MM, Bell RM, Yellon DM. P13 Kinase and not p42/p44 appears to be implicated in the protection conferred by ischemic preconditioning. *J Mol Cell Cardiol* 2002; 34: 661-668.

[150] O'Rourke B. Evidence for mitochondrial K<sup>+</sup> channels and their role in cardioprotection. *Circ Res* 2004; 94: 420-432.

[151] Editorial. A "radical idea" comes of age: oxidant mitochondrial signalling in health and disease. *J Mol Cell Cardiol* 2004; 37: 1113-1117.

[152] Murphy E. Primary and secondary signalling pathways in early preconditioning that converge on the mitochondria to produce cardioprotection. *Circ res* 2004; 94: 7-16.

[153] Tong H, Rockman HA, Koch WJ, Steenbergen C, Murphy E. G protein-coupled receptor in ternalization signalling is required for cardioprotection in ischemic preconditioning. *Circ Res* 2004; 94: 1133-1141.

[154] Uchiyama T, Engelman RM, Maulik N, Das DK. Role of Akt signaling in mitochondrial survival pathway triggered by hypoxic preconditioning. *Circulation* 2004; 109: 3042-3049.

[155] Takeishi Y, Huang Q, Wang T, Glassman M, Yoshizumi M, Baines CP, Lee J-D, Kawakatsu H, Che W, Lerner-Marmarosh N, Zhang C, Yan C, Ohta S, Walsh RA, Berk BC, Abe J-i. Src family kinase and adenosine differentially regulate multiple MAP kinases in ischemic myocardium: modulation of MAP kinases activation by ischemic preconditioning. *J Mol Cell Cardiol* 2004; 33: 1989-2005.

[156] Gross ER, Peart JN, Hsu AK, Grover GJ, Gross GJ. K<sub>ATP</sub> opener-induced delayed cardioprotection: involvement of sarcolemmal and mitochondrial K<sub>ATP</sub> channels, free radicals and MEK1/2. *J Mol Cell Cardiol* 2003; 35: 985-992.

[157] Baines CP, Zhang J, Wang G-W, Zheng Y-T, Xiu JX, Cardwell EM, Bolli R, Ping P. Mitochondrial PKC $\epsilon$  and MAPK form signaling modules in the murine heart. Enhanced mitochondrial PKC $\epsilon$ -MAPK interactions and differential MAPK activation in PKC $\epsilon$ -induced cardioprotection. *Circ Res* 2002; 90: 390-397.

[158] Marais E, Genade S, Hulssamen B, Strijdom JA, Moolman JA, Lochner A. Activation of p38MAPK induced by a multi-cycle ischaemic preconditioning protocol is associated with attenuated p38 MAPK activity during sustained ischaemia and reperfusion. *J Mol Cell Cardiol* 2001; 33: 769-778.

[159] Marais E, Genade S, Strijdom H, Moolman JA, Lochner A. p38 MAPK activation triggers pharmacologically-induced  $\beta$ -adrenergic preconditioning, but not ischaemic preconditioning. *J Mol Cell Cardiol* 2001; 33: 2157-2177.

[160] Fryer RM, Hsu AK, Gross GJ. Mitochondrial K<sub>ATP</sub> channel opening is important during index ischemia and following myocardial reperfusion in ischemic preconditioned rat hearts. *J Mol Cell Cardiol* 2001; 33: 831-834.

[161] Laclau MN, Boudina S, Thambo JB, Tariosse L, Gouverneur G, Bonoron-Adele S, Saks VA, Garlid KD, Dos Santos P. Cardioprotection by ischemic preconditioning preserves mitochondrial function and functional coupling between adenine nucleotide translocase and creatine kinase. *J Mol Cell Cardiol* 2001; 33: 947-956.

[162] Armstrong SC, Shivell LC, Ganote CE. Differential translocation or phosphorylation of alpha B crystalline cannot be detected in ischemically preconditioned rabbit cardiomyocytes. *J Mol Cell Cardiol* 2000; 32: 1301-1314.

[163] Suzuki MJ, Nagase H, Day RM, Das DK. GATA-4 regulation of myocardial survival in the preconditioned heart. *J Mol Cell Cardiol* 2004; 37: 1195-1203.

[164] Jancso N, Jancso-Gabor A. Desensitization of sensory nerve endings (in Hungarian). *Kiserl Orvostud* 1949; 2 (Suppl.): 15.

[165] Caterina MJ, Schumacher MA, Tominaga M, Rosen TA, Levine JD, Julius D. The capsaicin receptor: a heat-activated ion channel in the pain pathway. *Nature* 1997; 389: 816-824.

[166] Griffiths CD, Geraghty DP, Eldershaw TPD, Colquhoun EQ. Acute and chronic effects of capsaicin in perfused rat muscle: the role of tachykinins and calcitonin gene-related peptide. *J Pharmacol Exp Ther* 1998; 287: 697-704.

[167] Mulderry PK, Ghatei MA, Rodrigo J, Allen JM, Rosenfeld MG, Polak JM, Bloom SR. Calcitonin gene-related peptide in cardiovascular tissues of the rat. *Neuroscience* 1985; 14: 947-954.

[168] Rubino A, Ralevic V, Burnstock G. Sympathetic neurotransmission in isolated rat atria after sensory-motor denervation by neonatal treatment with capsaicin. *J Pharmacol Exp Ther* 1997; 282: 671-675.

[169] Holzer P. Capsaicin: Cellular targets, mechanisms of action, and selectivity for thin sensory neurons. *Pharmacol Review* 1991; 43: 143-201.

[170] Szallasi A, Blumberg PM. Vanilloid receptors: new insights enhance potential as a therapeutics target. *Pain* 1996; 68: 195-208.

[171] Szallasi A, Blumberg PM. Vanilloid (capsaicin) receptors and mechanisms. *Pharmacol Rev* 1999; 51:159-211.

[172] Holman JJ, Craig RK, Marshall I. Human  $\alpha$ - and  $\beta$ -CGRP and rat  $\alpha$ -CGRP are coronary vasodilators in the rat. *Peptides* 1986; 7: 231-235.

[173] Franco-Cereceda A, Lundberg JM. Actions of calcitonin gene-related peptide and tachykinins in relation to the contractile effects of capsaicin in the guinea-pig and rat heart in vitro. *Naunyn-Schmiedeberg's Arch Pharmacol* 1988; 337: 649-655.

[174] Franco-Cereceda A, Saria A, Lundberg JM. Differential release of calcitonin gene-related peptide and neuropeptide Y from the isolated heart by capsaicin, ischaemia, nicotine, bradykinin and ouabain. *Acta Physiol Scand* 1989; 135: 173-187.

[175] Node K, Huo Y, Ruan X, Yang B, Spiecker M, Ley K, Zeldin DC, Liao JK. Anti-inflammatory properties of cytochrome P450 epoxygenase-derived eicosanoids. *Science* 1999; 285: 1276-1279.

[176] Fissilthaler B, Popp R, Kiss L, Potente M, Harder DR, Fleming I, Busse R. Cytochrome P450 2C is an EDHF synthase in coronary arteries. *Nature* 1999; 401: 493-497.

[177] Zygmunt PM, Petersson J, Andersson DA, Chuang H, Sorgard M, Di Marzo V, Julius D, Hogestatt ED. Vanilloid receptors on sensory nerves mediate the vasodilator action of anandamide. *Nature* 1999; 400:452-457.

[178] Yamato T, Aomine M, Noto H, Ikeda M, Ohta C. Capsaicin does not inhibit the intracellular calcium handling process in rat ventricular papillary muscle. *Gen Pharmacol* 1996; 27: 105-108.

- [179] Yamato T, Aomine M, Ikeda M, Noto H, Ohta C. Inhibition of contractile tension by capsaicin in isolated rat papillary muscle. *Gen Pharmacol* 1996; 27: 129-132.
- [180] Castle NA. Differential inhibition of potassium currents in rat ventricular myocytes by capsaicin. *Cardiov Res* 1992; 26: 1137-1144.
- [181] D'Alonzo AJ, Grover GJ, Darbenzio RB, Hess TA, Sleph PG, Dzwonczyk S, Zhu JL, Sewter JC. In vitro effects of capsaicin: antiarrhythmic and antiischemic activity. *Eur J Pharmacol* 1995; 272: 269-278.
- [182] Yaoita H, Sato E, Kawaguchi M, Saito T, Maehara K, Maruyama Y. Nonadrenergic noncholinergic nerves regulate basal coronary flow via release of capsaicin-sensitive neuropeptides in the rat heart. *Circ Res* 1994; 75: 780-788.
- [183] Oroszi G, Szilvassy Z, Nemeth J, Tosaki A, Szolcsanyi J. Interplay between nitric oxide and CGRP by capsaicin in isolated guinea-pig heart. *Pharmacol Res* 1999; 40:123-128.
- [184] Li Y, Xiao Z-S, Peng C-F, Deng H-W. Calcitonin gene-related peptide-induced preconditioning protects against ischemia-reperfusion injury in isolated rat hearts. *Eur J Pharmacol* 1996; 311: 163-167.
- [185] Schultz HD, Ustinova EE. Cardiac vagal afferent stimulation by free radicals during ischaemia and reperfusion. *Clin Exp Pharmacol Physiol* 1996; 23, 700-708.
- [186] Schultz HD, Ustinova EE. Capsaicin receptors mediate free radical-induced activation of cardiac afferent endings. *Cardiov Res* 1998; 38: 348-355.
- [187] Rossini L, Bernardi M. Cannabinoidi, vanilloidi e razionale farmacologico. *Lettere dalla Facolta'* 2001; 4: 15-20.
- [188] Rossini L, Martin E, Zhong M. Nitration of inducible nitric oxide synthase tyrosine residues in Raw 264.7 macrophages. *Pharmacologyonline* 2005; 2: 1-23.
- [189] Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* 1986; 74: 1124-1136.
- [190] Sarvazyan N. An alternative preconditioning mechanism ?. *J Mol Cell Cardiol* 1998; 30: 2785-2786.



- [191] Rossini L. Riflessi condizionati da stimoli termici nella cavia. I. Effetti della narcosi, dell' ipotermia e di farmaci psicotropi. Arch Sci Biol 1962; 46: 356-369.
- [192] Terzuolo CA, Chance B, Handelman E, Rossini L, Schmelzer P. Measurements of reduced pyridine nucleotides in a single neuron. Biochim Biophys Acta 1966; 126: 361-372.
- [193] Rossini L, Cohen HP, Handelman E, Lin S, Terzuolo CA. Measurements of oxidoreduction processes and ATP levels in an isolated crustacean neuron. Ann N Y Acad Sci 1966; 137: 864-876.
- [194] Rossini L, Rossini P, Chance B. Continuous read-out of cytochrome b, flavin and pyridine nucleotide oxido-reduction processes in the perfused frog heart and contracting skeletal muscle. Pharmacol Res 1991; 23: 349-365.
- [195] Rossini L, Bernardi M, Concettoni C, De Florio L, Deslauriers R, Moretti V, Piantelli F, Pignini P, Re L, Rossini P, Tonnini C. Some approaches to the pharmacology of multisubstrate enzyme systems. Pharmacol Res 1994; 29: 313-335.
- [196] Lee JWK, Rossini L, Saunders JK, Deslauriers R. Seasonal variation in isolated perfused *Xenopus Laevis* heart as characterized by <sup>31</sup>P and <sup>13</sup>C NMR spectroscopy: a new digitalis effect. Proceedings: Int Soc Magn Res/European Soc Magn Res, Nice, Aug. 19-25, 1995.
- [197] Bernardi M, Deslauriers R, Docherty J, Galeazzi G, Rossini L, Rossini P. Spectral analysis of intercycle heart fluctuations in the diethyl-ether-anaesthetized or pithed rat treated with l-hyoscyamine. J Autonom Pharmacol 1997; 17: 27-34.
- [198] Bernardi M, Deslauriers R, Docherty J, Rossi C, Rossini L, Rossini P, Tonnini C. Spectral analysis of intercycle heart fluctuations in the diethyl-ether-anaesthetized or pithed rat treated with prazosin, dl-propranolol, endothelin-1,  $\alpha$ -r atriopeptin and ACE-inhibitors. J Autonom Pharmacol 1998; 18: 271-280.
- [199] Maulik N, Engelman RM, Rousou JA, Flack III JE, Deaton D, Das DK. Ischemic preconditioning reduces apoptosis by upregulating anti-death gene Bcl-2. Circulation 1999; 100: 369-375.
- [200] Okamura T, Miura T, Iwamoto H, Shirakawa K, Kawamura SW, Ikeda Y, Iwatate M, Matsuzaki M. Ischemic preconditioning attenuates apoptosis through protein kinase C in rat hearts. Am J Physiol 1999; 277: H1997-H2001.
- [201] Patel HH, Hsu A, Moore J, Gross GJ. BW373U86, a  $\delta$  opioid agonist, partially mediates delayed cardioprotection via a free radical

mechanism that is independent of opioid receptor stimulation. *J Mol Cell Cardiol* 2001; 33: 1455-1465.

[202] Patel HH, Fryer RM, Gross ER, Bunday RA, Hsu AK, Isbell M, Eusebi LOV, Jensen RV, Gullans SR, Insel PA, Nithipatikom K, Gross GJ. 12-Lipoxygenase in opioid-induced delayed cardioprotection. Gene array, mass spectrometric, and pharmacological analyses. *Circ Res* 2003; 92: 676-682.

[203] Patel HH, Hsu AK, Peart JN, Gross GJ. Sarcolemmal K ATP channel triggers opioid-induced delayed cardioprotection in the rat. *Circ Res* 2002; 91: 186-188.

[204] Wang Y, Guo Y, Zhang SX, Wu W-J, Wang I, Bao W, Bolli R. Ischemic preconditioning upregulates inducible nitric oxide synthase in cardiac myocytes. *JU Mol Cell Cardiol* 2002; 34: 5-15.

[205] Wang Y, Kodani E, Wang J, Zhang SX, Takano H, Tang X-L, Bolli R. Cardioprotection during the final stage of the late phase ischemic preconditioning is mediated by neuronal NO synthase in concert with cyclooxygenase-2. *Circ Res* 2004; 95: 84-91.

[206] Dickson EW, Blehar DJ, Carraway RE, Heard SO, Steinberg G, Przyklenk K. Naloxone blocks transferred preconditioning in isolated rabbit hearts. *J Mol Cell Cardiol* 2001; 33: 1751-1756.

[207] Patel HH, Moore J, Hsu AK, Gross GJ. Cardioprotection at a distance: mesenteric artery occlusion protects the myocardium via an opioid sensitive mechanism. *J Mol Cell Cardiol* 2002; 34: 1317-1323.

[208] Heusch G, Schulz R. Remote preconditioning. *J Mol Cell Cardiol* 2002; 34: 1279-1281.

[209] Argaud L, Gateau-Roesch O, Raisky O, Loufouat J, Robert D, Ovize M. Postconditioning inhibits mitochondrial permeability transition. *Circulation* 2005; 111: 194-197.

[210] Tsang A, Hausenloy DJ, Mocanu MM, Yellon DM. Post-conditioning: a form of "modified reperfusion" protects the myocardium by activating the phosphatidylinositol 3-kinase-Akt pathway. *Circ Res* 2004; 95: 230-232.

[211] Rossini L, Bernardi M, Galeazzi G, Moroni L, Pettinari F, Pignini P, Rossini P, Tonnini C, Vagionis G, Violet C. Domini del tempo e di frequenza in fenomeni biomedici. II. *Atti Acc. March. Scienze, Lettere Arti* 2005; 38: 211-256.

[212] Opie LH. Myocardial Energy Metabolism. *Adv Cardiol* 1974; 12: 70-83.

[213] Ferdinandy P, Panas D, Schulz R. Peroxynitrite contributes to spontaneous loss of cardiac efficiency in isolated working rat hearts. *Am J Physiol* 1999; 276: H1861-H1867.

[214] Strasser RH, Simonis G, Schön SP, Braun MU, Ihl-Vahl R, Weinbrenner C, Marquetant R, Kübler W. Two distinct mechanisms mediated a differential regulation of protein kinase C isozymes in acute and prolonged myocardial ischemia. *Circ Res* 1999; 85: 77-87.

[215] Ogura T, Kasamaki Y, McDonald TF. Force-relaxant actions of dimethyl sulfoxide on guinea-pig and rabbit papillary muscles. *J Mol Cell Cardiol* 1996; 28: 1777-1788.

[216] Chance B, Williamson JR, Jamieson D, Schoener B (1965) Properties and kinetics of reduced pyridine nucleotide fluorescence of the isolated and in vivo rat heart. *Biochemische Zeitschrift* 1965; 341: 357-377.

[217] Chance B, Mayer D, Rossini L. A time-sharing instrument for direct readout of oxidation-reduction states in intracellular compartments of cardiac tissue. *IEEE Trans On Bio-Med Eng* 1970; BME-17: 118-121.

[218] Chance B, Salkovitz IA, Kovach AGB. Kinetics of mitochondrial flavoprotein and pyridine nucleotide in perfused heart. *Am J Physiol* 1972; 213: 207-218.

[219] Benson RC, Meyer RA, Zaruba ME, McKhann GM. Cellular autofluorescence - is it due to flavins ?. *J Histochem Cytochem* 1979; 27: 44-48.

[220] Bergamini PG, Palmas G, Piantelli F, Sani M, Cingolani ML, Leone L, Re L, Roda G, Rossini L. A multi- $\lambda$  device for bioluminescence measurements in vivo. *Chem Biomed Environ Instrumentation* 1980; 10: 289-309.

[221] Sato T, Sasaki N, Seharaseyon J, O'Rourke B, Marban E. Selective pharmacological agents implicate mitochondrial but not sarcolemmal  $K_{ATP}$  channels in ischemic cardioprotection. *Circulation* 2000; 101: 2418-2423.

[222] Nuutinen EM. Subcellular origin of the surface fluorescence of reduced nicotinamide nucleotides in the isolated perfused rat heart. *Basic Res Cardiol* 1984; 79: 49-58.

[223] Williamson RJ, Jameson D. Metabolic effects of epinephrine in the perfused rat heart I. Comparison of intracellular redox states, tissue  $pO_2$ , and force of contraction. *Mol Pharmacol* 1996; 2: 191-205.

- [224] Eto K, Tsubamoto Y, Terauchi Y, Sugiyama T, Kishimoto T, Takahashi N, Yamauchi T, Kubota N, Murayama S, Aizawa T, Akanuma Y, Aizawa S, Kasai H, Yazaki Y, Kadowaki T. Role of NADH shuttle system in glucose-induced activation of mitochondrial metabolism and insulin secretion. *Science* 1999; 283: 981-985.
- [225] Barlow CH, Chance B. Ischemic areas in perfused rat hearts: measurements by NADH fluorescence photography. *Science* 1976; 193: 909-910.
- [226] Steenbergen C, Deleeuw G, Barlow C, Chance B, Williamson JR. Heterogeneity of the hypoxic state in perfused rat heart. *Circ Res* 1977; 41: 606-615.
- [227] Steenbergen C, Williamson JR. Heterogeneous coronary perfusion during myocardial hypoxia. *Advances in Myocardiology* 1980; 2: 271-283.
- [228] Di Sarra B, Piantelli F, Moretti V, Re L, Rossini L, Tonnini C. Physio-pharmaco-toxicological in vivo read-out: An interuniversity integrated analytical center. Issues, results and perspectives. *Quad March Med* 1989; 5: 183-185.
- [229] Frank KH, Kessler M, Appelbaum K, Dümmler W. The Erlagen micro-lightguide spectrophotometer EMPHO I. *Phys Med Biol* 1989; 34: 1883-1900.
- [230] Ince C, Coremans JMCC, Bruining HA. In vivo NADH fluorescence. *Adv Exp Med Biol* 1992; 317: 277-296.
- [231] Hulsmann WC, Ashruf JF, Bruining HA, Ince C. Imminent ischemia in normal and hypertrophic Langerdorff rat hearts; effects of fatty acids and superoxyde dismutase monitored by NADH surface fluorescence. *Biochim Biophysica Acta* 1993; 1181: 273-278.
- [232] Ince C, Ashruf JF, Avontuur JAM, Wieringa PA, Spaan JAE, Bruining HA. Heterogeneity of the hypoxic state in rat heart is determined at capillary level. *Am J Physiol* 1993; 264: H294-H301.
- [233] Ashruf JF, Ince C, Bruining HA. Regional ischemia in hypertrophic Langendorff-perfused rat hearts. *Am J Physiol* 1999; 277: H1532-H1539.
- [234] Brasch F, Neckel M, Volkmann R, Schmidt G, Hellige G, Vetterlein F. Mapping of capillary flow, cellular redox state, and resting membrane potential in hypoperfused rat myocardium. *Am J Physiol* 1999; 277: H2050-H2064.
- [235] Efimov IR, Nikolski VP, Salama G. Optical imaging of the heart. *Circ Res* 2004; 95: 21-33.