ANTIOXIDANT AND ANTICHOLINERGIC ACTIVITY OF RUBIA CORDIFOLIA

Rupali Patil^{1*}, Rajendra Gadakh¹, Hanmant Gound¹, Sanjay Kasture²

- 1. *Department of Pharmacology, MGV's Pharmacy College, Panchavati, Nashik- 422 003, Maharashtra, India.
- 2. Pinnacle Biomedical Research Institute, Bhopal, MP, India

Summary

Rubia cordifolia Linn (Rubiaceae) is an important medicinal plant commonly used in the traditional and Ayurvedic system of medicine for treatment of different ailments. In this study, the *in vitro* antioxidant status of methanolic extract of roots and rhizomes of *R. cordifolia* (M-RC) was determined. The effect of M-RC on tacrine-induced tremulous jaw movements in rats and sodium nitrite-induced hypoxia in mice was studied. IC₅₀ value for lipid peroxidation of a linoleic acid emulsion was found to be 120 µg/ml. IC₅₀ value for free radical and hydroxyl radical scavenging activity were found to be 130 ppm and 135 ppm, respectively. In M-RC, $1.8679 \pm 0.29 µg/g$ gallic acid phenol equivalents were detected. M-RC significantly inhibited tacrine-induced vacuous chewing movements (VCM), tongue protrusions (TP) and orofacial bursts (OB). M-RC also significantly potentiates sodium nitrite-induced hypoxia and decreased the latency for death after sodium nitrite administration. The study concludes that *R. cordifolia* has an anticholinergic activity which may be attributed to antioxidant activity and presence of phenolic compounds.

Keywords: Phenol content, hypoxia, tacrine, vacuous chewing movements, *Rubia cordifolia*

Introduction

Free radicals are generated in the body due to the metabolism and disease.^[1] In order to protect themselves against free radicals, organisms are having endogenous (catalase, superoxide dismutase, glutathione peroxidase/reductase) and exogenous (vitamin C and E) defenses. But these defense systems are not sufficient in critical situations like oxidative stress, UV exposure etc., when the production of free radicals significantly increases.^[2]

An excess generation of free radicals and a deficient cellular antioxidant defense system may lead to a state of oxidative stress. Oxidative stress and products of lipid peroxidation are involved in the physiology of various neurological disorders such as stroke, Parkinson's disease, Alzheimer's disease,^[3] carcinogenesis.^[4] Idiopathic and neuroleptic-induced parkinsonism are often treated with muscarinic receptor antagonists.^[5,6,7]

The anticholinesterase inhibitor tacrine is used therapeutically to improve memory function in patients with early and late onset Alzheimer's disease. But it can lead to the production of parkinsonian side-effects such as bradykinesia, rigidity, tremor.^[8,9] Ott and Lannon^[8] demonstrated that tacrine-induced parkinsonism could be ameliorated by L-3, 4-dihydroxyphenylalanine (L-DOPA). One of the motor effects produced by cholinomimetics is tremulous jaw movements (also known as 'VCM' or 'purposeless' chewing). These are characterised as rapid, vertical deflections of the lower jaw that resemble chewing but are not directed at any stimulus. They share some characteristics with human parkinsonian symptoms.^[10,11] Tacrine-induced tremulous jaw movements can be suppressed by antiparkinsonian agents. Apomorphine, benztropine block tacrine-induced jaw movements.^[12]

Rubia Cordifolia Linn (Rubiaceae) also known as 'manjistha' is an important medicinal plant. It possesses anti-tumor,^[13] antistress and antihyperglycemic,^[14] antimicrobial,^[15] hepatoprotective,^[16] radio protective,^[17] anticancer^[18] activities. This plant has also been listed officially as herbal medicine in the Chinese Pharmacopeia for the treatment of arthritis, dysmenorrhea, hematorrhea and hemostasis, which are free radical related diseases. In the present study, we have assessed the *in vitro* antioxidant activity and anticholinergic activity. Total phenolic content was determined using Folin-Ciocalteu reagent.

Materials and methods

Animals

Male Wistar rats $(150 \pm 20 \text{ g})$ and Swiss albino mice $(22 \pm 2g)$ were used for the study. Animals were housed in a colony cages and maintained at $25^\circ \pm 2^\circ$ C, 12:12 h light/dark cycle and $50 \pm 5\%$ RH with free access to food and water *ad libitum*. Animals were acclimatized to laboratory conditions before test. All the experiments were carried out during the light period (08.00-16.00 h). The Institutional Animal Ethical Committee of MGV's Pharmacy College, Nashik approved the protocol of the study.

Chemicals

Folin-ceiocalteu reagent (Merck, India), tacrine (Sigma, MO), Vitamin E (Merck, India) were used for the study. All other chemicals and reagents were of analytical grade.

Plant material and Extraction

Roots and rhizomes of *Rubia cordifolia* Linn. were obtained from Ayurved Seva Sangh, Nashik, India and authenticated by Dr. P. G. Diwakar, Joint director, Botanical Survey of India, Pune (Voucher specimen number: RAP 1) The collected material was extracted with acetone. The marc was dried and extracted with methanol by Soxhlet extractor. The extract was filtered and dried. The yield of methanolic extract (M-RC) was found to be 5.2 % w/w.

Phytochemical studies

M-RC was subjected to identification of phytoconstituents as suggested by Evans.^[19]

Determination of total antioxidant activity

Total antioxidant activity of M-RC was determined spectrophotometrically using the thiocyanate method.^[20] The absorbance was measured at 500 nm. All data reported are the average of triplicate analyses. Percent inhibition of lipid peroxide generation was calculated using following formula.

% inhibition= [(absorbance of control- absorbance of test)/ absorbance of control] x 100

Determination of Free Radical Scavenging Activity

DPPH scavenging activity was measured by the spectrophotometer.^[21] To an ethanolic solution of DPPH (200 µM), 0.05 ml of the test compounds dissolved in ethanol were added at different concentrations. An equal amount of ethanol was added to the control. After 20 min the decrease in absorbance of the test mixtures (due to quenching of DPPH free radicals) was read at 517 nm and the percentage inhibition calculated. Each trial has been done in triplicate. IC_{50} value was calculated by plotting graph %Scavenging verses Concentration.

% scavenging = [(absorbance of control- absorbance of test)/(absorbance of control] x 100

Hydroxyl radical scavenging activity

This method involves *in-vitro* generation of hydroxyl radicals using Fe³⁺/ascorbate/ EDTA/ H₂O₂ system using Fenton reaction. Scavenging of this hydroxyl radical in presence of antioxidant is measured. The hydroxyl radicals formed by the oxidation is made to react with DMSO (dimethyl sulphoxide) to yield formaldehyde. The intensity of yellow color formed by reaction of formaldehyde with NASH reagent (2M ammonium acetate with 0.05M acetic acid and 0.02M acetyl acetone in distilled water) is measured at 412nm against reagent blank.^[22] The activity is expressed as % hydroxyl radical scavenging.

IC₅₀ value was calculated by plotting graph %Scavenging verses Concentration.

% Hydroxyl radical scavenging activity= [(absorbance of control- absorbance of test)/ absorbance of control] x 100

Amount of total phenolics

Total phenolics present in the M-RC was determined spectrophotometrically with the Folin-Ciocalteu reagent, according to the method suggested by Slinkard and Singleton.^[23] The absorbance was measured at 760 nm. All tests were performed in triplicate. The concentration of total phenolics in MERC was determined as µg gallic acid equivalents, using the following equation obtained from a standard gallic acid graph:

Absorbance = 0.159 x gallic acid (µg) + 0.0290

Effect on Tacrine-induced orofacial dyskinesia

Rats were divided in 6 groups, each containing five animals. The rats received orally, vehicle and M-RC (100, 200 and 300 mg/kg) 1 h prior to tacrine (2.5 mg/kg, i.p.). Vitamin E (10 mg/kg, p.o.) was used as a reference. Immediately after injection of tacrine, rats were placed in a Plexiglas observation box $(22x22x22 \text{ cm}^3)$ for a 10 min habituation period. An observer blind to treatment recorded the number of vacuous chewing movements (VCM), tongue protrusions (TP), and number of orofacial bursts (OB) as described by Cousins.^[12] All rats were observed for 1 h period.

Effect on sodium nitrite-induced hypoxia

Sodium nitrite (250 mg/kg, s.c.) was administered to induce hypoxia.^[24,25] M-RC (100, 200 and 300 mg/kg, p.o.) was administered 1 h before treatment with sodium nitrite. Duration to induce hypoxia was recorded.

Statistical analysis

Data are expressed as mean \pm SEM. Statistical analysis was done by using one way analysis of variance (ANOVA) followed by Dunnett's test. Values with P < 0.05 were considered statistically significant.

Results

Phytochemical studies

M-RC revealed presence of glycosides, phenolics, flavonoids, and saponins.

Total antioxidant activity

IC₅₀ value for M-RC was found to be 120 μ g/ml.

Free radical and hydroxyl radical Scavenging activity

The IC₅₀ values of M-RC by DPPH free radical scavenging and hydroxyl radical scavenging method was found to be 135 and 130 ppm respectively. The results are summarized in table1.

Table 1: % Scavenging activity

Concentration (ppm)	% Scavenging activity by	
	DPPH method	Hydroxyl radical
		scavenging method
10	12.88 ± 0.01	22.96 ± 0.03
25	22.88 ± 0.02	31.11 ± 0.05
50	29.26 ± 0.05	39.74 ± 0.02
100	44.75 ± 0.09	49.02 ± 0.06
250	69.02 ± 0.04	53.32 ± 0.08
IC ₅₀	135	130

Each value in the table is obtained by calculating the average of three experiments and expressed as mean \pm SEM.

Amount of total phenolic content

M-RC contained $1.8679 \pm 0.29 \ \mu g/g$ Gallic acid equivalents of phenols. The phenolic compounds may contribute directly to the antioxidant action.^[26]

Tacrine-induced vacuous chewing movements

Tacrine shows significant increase in VCM, TP, OB. M-RC dose-dependently reduced tacrineinduced VCM, TP, OB. Vitamin E was used as a reference standard. It also significantly reduced tacrine-induced VCM, TP, OB (Table 2).

Table 2: Effect of *R. cordifolia* on tacrine-induced orofacial dyskinesia

Group	Number of		
	VCM	TP	OB
Control	119 ± 2.72	14.25 ± 0.85	55 ± 2.90
Tacrine (2.5)	$1141 \pm 12.93*$	$46 \pm 0.91*$	$259 \pm 2.58*$
M-RC 100	385 ± 10.41 **	21.5±1.32**	$137 \pm 5.30 **$
M-RC 200	243.5 ± 3.4 **	14.75 ± 1.11 **	118 ± 6.31 **
M-RC 300	223 ± 3.22**	$14 \pm 1.09^{**}$	$103 \pm 2.39 **$
Vitamin E 10	261.2 ± 9.44 **	$19.75 \pm 0.8 **$	$102.2 \pm 6.23 **$

n=5. Data are mean \pm SEM. **P* < 0.05 compared to control. ***P* < 0.05 compared to tacrine. VCM: vacuous chewing movement, TP: tongue protrusion, OB: orofacial burst, M-RC: methanolic extract of *R. cordifolia*

Sodium nitrite-induced hypoxia

M-RC significantly decreased the latency for death after sodium nitrite administration (Table 3).

Group	Time of respiratory arrest (min)
Control	22.55 ± 2.02
M-RC 100	$17.82 \pm 0.63*$
M-RC 200	$14.79 \pm 0.51*$
M-RC 300	$14.25 \pm 1.05*$

Table 3: Effect of R. cordifolia on Sodium nitrite-induced hypoxia

n=5. Data are mean \pm SEM. *P < 0.05 compared to control.

Discussion

Most of the mammals have an inherent mechanism to prevent and neutralize the free radical induced damage. In biochemical system, superoxide radical and H_2O_2 react together to form a singlet oxygen and hydroxyl radical, which can attack and destroy almost all known biochemicals.^[27] Hydroxyl radicals are the major ROS, causing lipid oxidation and enormous biological damage.^[28] It is apparent from the present study that the M-RC possesses antioxidant activity. M-RC showed antioxidant activity by inhibiting hydroxyl radical and DPPH scavenging. The roots of *R. Cordifolia* could be ranked as anthraquinone rich due to the strong violet colour indicated by the Borntreger's test. It was already reported that naturally occurring phenolic compounds have free radical scavenging properties, due to their hydroxyl groups.^[29] The majority of isolated molecules from the roots of *R. Cordifolia* elucidate as anthraquinones.^[30] It has been estimated that family Rubiaceae is rich in anthraquinone compounds.^[31,32] M-RC showed presence of phenolic compounds. The antioxidant potential may be attributed to the presence of polyphenolic compounds and anthraquinone glycosides.

Tacrine, the acetylcholinesterase inhibitor, used in treatment of early and late onset of Alzheimer's disease. Animal studies have shown that cholinomimetic drugs produce a wide variety of motor effects.^[33] Administration of tacrine is associated with a wide variety of peripheral and central side effects.^[35] Some of the central side effects of tacrine are extrapyramidal motor dysfunctions; these include several Parkinsonian symptoms such as cogwheel rigidity, tremor, and bradykinesia.^[8,35] There is a substantial literature showing that Parkinsonism can be exacerbated by cholinergic stimulation and alleviated by muscarinic antagonism.^[5,6,36] Previous studies with animals also have shown that cholinomimetic drugs have pronounced motor effects. The induction of parkinsonian symptoms by cholinergic involvement in idiopathic and neuroleptic-induced parkinsonism.^[5,6] Cholinomimetic drugs induce perioral movements in rats.^[12,37,38] In rats, drugs that stimulate muscarinic cholinergic receptors produce a number of different orofacial movements, the most common of which is known as vacuous jaw movements. ^[38,39] M-RC significantly decreased the latency for death after treatment with sodium nitrite. It may suggest cholinergic inhibition as observed with inhibition of tacrine-induced VCM, TP, and OB. It may be concluded that M-RC has significant antioxidant activity. The study highlights the importance of *R. Cordifolia* as an alternative system of medicine for parkinsonism.

Conclusion

The study concludes that *R. cordifolia* has an anticholinergic activity which may be attributed to antioxidant activity and presence of phenolic compounds.

Acknowledgements

The authors are grateful to the management and Principal, MGV's Pharmacy College, Nashik for providing research facilities.

References

- 1. Yeum KJ, Aldini G, Chung Hy. The activities of antioxidant nutrients in human plasma depend on the localization of attacking radical species. The J Nutrition 2003;133,2688-2691
- 2. Mondon P, Leclercq L, Lintner K. Evaluation of free-radical scavenger effects of Helianthus annus extracts using new ex vivo tripping methods. Cosmetics, Aerosols, and Toiletries in Australia. 1999;12:87-98.
- 3. Beal MF. Oxidative damage in neurodegenerative diseases. Neuroscientist 1958;3: 327-333.
- 4. Galati G, O'Brien PJ. Potential toxicity of flavonoids and other dietary phenolics: significance for their chemopreventive and anticancer properties. Free rad Biol Med 2004;37:287-303.
- 5. Duvoisin RC.Cholinergic-anticholinergic antagonism in parkinsonism. Arch Neurol 1967;17:124-136.
- 6. McEvoy JP. The clinical use of anticholinergic drugs as treatment for extrapyramidal side effects of neuroleptic drugs. J Clin Psychopharmacol 1983;3: 288-301.
- 7. Tarsy D. Neuroleptic-induced extrapyramidal reactions: classification, description and diagnosis. Clin Neuropharmacol 1983;6:509-529.
- Ott BR, Lannon MC. Exacerbation of Parkinsonism by tacrine. Clin Neuropharmacol 1992;15: 322-325
- 9. Keltner NL. Tacrine: a pharmacological approach to Alzheimer's disease. J Psychosoc Nurs Ment Health Serv 1994;32:37.
- 10. Salamone JD, Johnson CJ, McCullough LD, Steinpreis RE. Lateral striatal cholinergic mechanisms involved in oral motor activities in the rat. Psychopharmacol 1990;102:529-534.
- 11. Jicha G, Salamone JD. Vacuous Jaw movements and feeding deficits in rats with ventrolateral striatal dopamine depletions: Possible model of Parkinsonian symptoms. J Neurosci 1991;11:3822-3829.
- 12. Cousins MS, Carriero DL, Salamone JD. Tremulous jaw movements induced by the acetylcholinesterase inhibitor tacrine: Effects of antiparkinsonian drugs. Eur J Pharmacol 1997;322:137-145.
- 13. Adwankar MK, Chitnis MP. In vivo anti-cancer activity of RC-18:a plant isolate from *Rubia Cordifolia* Linn. against a spectrum of experimental tumor models. Chemotherapy 1982;28:291-293.
- 14. Patil RA, Jagdale SC, Kasture SB. Antihyperglycemic, antistress and nootropic activity of roots of *Rubia cordifolia* Linn. Indian J Exp Biol 2006;44:987-992.
- 15. Singh R, Jain A, Panwar S. Antimicrobial activity of some natural dyes. Dyes and Pigments 2005;66:99-102.
- 16. Rao MGM, Rao CV, Pushpangadan P. Hepatoprotective effects of rubiadin, a major constituent of *Rubia Cordifolia* Linn. J Ethnopharmacol 2006;103:484-490.
- 17. Tripathi YB, Singh AV. Role of *Rubia Cordifolia* Linn. in radiation protection. Indian J Exp Biol 2007;457:620-625.

- 18. Son JK, Jung SJ, Jung JH. Anticancer constituents from the roots of *Rubia cordifolia* L. Chem Pharm Bull 2008;562:213-216.
- 19. Evans WC. Trease and Evans' Pharmacognosy, Elsevier, New Delhi, India. 2005: 223, 230, 247, 289, 339.
- 20. Mistuda H, Yuasumoto K, Iwami K. Antioxidation action of indole compounds during the autoxidation of linoleic acid. Nihon Eiyo Shokuryo Gakkai-Shi 1996;19:210-214
- Keto K, Terao S, Shimamoto N, Hirata M. Studies on scavengers of active oxygen species. J Med Chem 1988:31: 793-798.
- 22. Rajeshwar Y, Kumar GP, Gupta M, Maunder UK. Studies on in-vitro antioxidant activities of methanol extract of *Mucuna pruriens* (Fabaceae) seeds. Euro Bull Drug Res 2005;13(1):31-39.
- 23. Slinkard K, Singleton VL. Total phenol analyses: Automation and comparison with manual methods. Am J Enol Viticulture 1977;28:49-55
- 24. Vogel HG, Vogel WH. Drug discovery and evaluation, Springer, 2nd edition, New York, NY. 2002:643.
- 25. Jain NN, Ohal CC, Shroff SK, Bhutada RH, Somani RS, Kasture VS, Kasture SB. *Clitoria ternatea* and the CNS. Pharmacol Biochem Behav 2003;75:529-536.
- 26. Rice-Evans CA, Miller NJ, Bolwell PG, Bramley PM, Pridham JB. The relative antioxidant activity of plant derived polyphenolic flavonoids. Free Radic Res 1995;22:375-383.
- 27. Chakraborti S, Naik AA, Reddy GR. Phenylhydrazine mediated degradation of bovine serum albumin and membrane proteins of human erythrocytes. Biochem Biophys Acta 1990;1028:89-94.
- 28. Aurand LW, Boonme NH, Gidding GG. Superoxide and singlet oxygen in milk lipid peroxidation. J Dairy Sci 1977;60:363-369.
- 29. Diplock AT. Will the `good fairies' please prove to us that vitamin E lessens human degenerative disease? Free Radic Res 1997;27:511-532.
- 30. Singh R, Geetanjali. Isolation and synthesis of anthraquinones and related compounds of *Rubia Cordifolia*. J Serbian Chem Soc 2004;70:937-942.
- 31. Verpoorte R, Wijnsma R, Mulder-Krieger TH. Primary and secondary metabolism of plant cell cultures. Berlin, Heidberg:Springer-Verlag. 1985:196-208.
- 32. Wijnsma R, Verpoorte R, Mulder KT. Anthraquinones in callus cultures of *Cinchona ledgeriana*. Phytochem 1984;23:2307-2311.
- 33. Cools AR, Hendriks G, Korten J. The acetylcholine-dopamine balance in the basal ganglia of Rhesus monkeys and its role in dynamic, dystonic, dyskinetic epileptoid activity. J Nerv Trans 1975;36:91–105.
- 34. Salamone JD, Baskin PB. Vacuous jaw movements induced by reserpine and low-dose apomorphine: possible model of parkinsonian tremor. Pharmacol Biochem Behav 1996;53:179.
- 35. Montville NJ. Physicians Desk Reference. Medical Economics, 1995.
- 36. Noring U, Povlesen UJ, Casey DE, Gerlach J. Effect of a cholinomimetic drug (RS 86) in tardive dyskinesia and drug related parkinsonism. Psychopharmacol (Berlin) 1984;84:569-571.
- 37. Chesler EJ, Salamone JD. Effects of acute and repeated clozapine injections on cholinomimeticinduced vacuous jaw movements. Pharmacol Biochem Behav 1996;54:619-624.
- 38. Salamone JD, Lahes MD, Channell SL, Iversen SD. Behavioral and pharmacological characterization of the mouth movements induced by muscarinic agonists in the rat. Psychopharmacol 1986;88:467-471.
- Baskin P, Gianutsos G, Salamone JD. Repeated scopolamine injections sensitize rats to pilocarpineinduced vacuous jaw movements and enhance striatal muscarinic receptor binding. Pharmacol Biochem Behav 1994;49:437.