Pharmacologyonline 3: 1-5 (2009)

EVALUATION OF ANTIFUNGAL ACTIVITY OF METHANOLIC EXTRACT OF LEAVES AND STEMS OF SOLANUM SISYMBRIIFOLIUM LAM.

Jagruti Vaghela^{1*}, Mehul Rana¹, Vaibhavi Savalia³, N.R. Sheth²

1* & 1 Department of Pharmacognosy, S.J. Thakkar Pharmacy College, Rajkot, Gujarat, India.

2 Professor and Head, Department of Pharmaceutical Sciences, Saurashtra University, Rajkot, Gujarat, India.

3 Department of Pharmacognosy, R.K. College of Pharmacy, Rajkot, Gujarat, India.

Summary

Invasive fungal infections are significant causes of morbidity and moratality, particularly in immunocompromised patients. *In-vitro* antifungal activity of methanolic extract of leaves (MELS) and stems (MESS) of *Solanum sisymbrifolium* Lam. was assessed against *Aspergillus niger*, *A.Flavus*, *A.xylinium* and *Candida albicans* by agar well diffusion method using Fluconazole (100 μ g/ml) as a positive control. The stem extract was shown large zone of inhibition against all three species of *Aspergillus* and also active against *Candida albicans* compare to stem extract. The MESS was found active against all three species of *Aspergillus* and also active against *Candida albicans*. MESS has significant antifungal activity compare to MELS.

Keywords: Solanum sisymbrifolium, Antifungal activity, Aspergillus niger, Aspergillus flavus, Aspergillus xylinium and Candida albicans.

*Corresponding Author Jagruti P Vaghela, Department of Pharmacognosy, S.J. Thakkar Pharmacy College, Rajkot-360005 Gujarat, India. M.No: +91-9904230519 E-mail: Jagruti 910@yahoo.com

Introduction

Higher incidence of fungal infection caused by various species of fungi has been reported, hence, research on bioactive substances that may lead to the discovery of new compounds is required. There has been a dramatic increase in the use of antifungal agents for the treatment of both systemic and localized fungal infection but the expanded use of antifungal agents has accelerated the development of resistance to antifungal drugs followed by frequent therapeutic failures and increasing mortality rate^[1]. The most important and dreadful species of fungi includes *Aspergillus niger*, *A.flavus*, *A.xylinium* and *Candida albicans*. Despite advances in antifungal therapies, many problems related to drug resistance and toxicity remains to be solved, for example, the toxicity of Amphotercin and development of clinically resistance strains of various fungal species on continuous use of Azoles.

This situation highlights the need for advent of safe, novel and effective antifungal agents. The main objective of the present study is to explore the antifungal potential of *Solanum sisymbrifolium* Lam. It is commonly known as Red Buffalo bur belongs to Solanaceace family. It is a native of South America. In India it was found in Central, East, North and South^[2-6]. The Leaves pinnately lobed with yellow spines, alternate; Flowers pale blue to white, 11/4 in. in diameter; The Berry globose, 2.5 cm in diameter, yellowish-red, included in the accrescent calyx which has a prickly tube, with the apex open; Stem branched, woody at base, villose-pubescent with long viscid hairs and armed all over with bright yellow prickles.

In traditional medicine it is used in the treatment of respiratory diseases, as anti-inflammatory drug. The drug is of high interest as it has been mentioned as possible treatment of female infertility and for promoting conception in females^[7,8]. Roots are used in the treatment of hypertensive diseases in Paraguay^[12], as diuretic, analgesic, contraceptive, antisyphilitic and hepatoprotective in Argentine^[9]. Aerial parts are used in Argentine to treat diarrhea, infections of respiratory and urinary tracts^[10] and Flowers are used as analgesic in India^[11]. Leaves are as febrifuge in Peru and as diuretic in Brazil^[12,13]. The plant is also used as emenagogue and for fertility regulation^[14,15]. The hypotensive effect of the root extract and its components shown in both normo- and hypotensive rats^[16,17]. Alkaloid solasodine from leaves^[18], alkaloids from roots^[19-21], Lignans and Steroids from fruits^[21,24] are already reported. The present study aims at evaluation of leaf and stem extract for its antifungal activity.

Materials and Methods

Collection and identification of Plant material.

Collection of stem and leaf part of *Solanum sisymbriifolium* were done from the wild sources near by Saurashtra university campus during August/September, 2007 and identification and authentication were done by Dr. Reddy, Department of Bioscience, S. P. University, V. V. Nagar, Gujarat. The voucher specimen no. Herbarium/2007-08/03 of the collected sample was deposited in the Department of Pharmaceutical Sciences, Saurashtra University, Rajkot, Gujarat for future reference.

Newsletter

Vaghela *et al*.

Prepartion of methanolic extract

About 500 gm of dried powder of both leaf and stem were obtained from fresh leaves and stem weighing about 3 and 4 kg, respectively. They were powder and extracted with 80 % methanol using the soxhlet apparatus (soxhlet F) for about 48 hours. After extraction, the methanolic extracts filtered through Whatmann filter paper No.1. The filtrate were dried in the vacuum distillation and then in dessicator.

Antifungal activity

Cup-plate method was used for screening the antifungal activity of methanolic extract of leaf (MELS) and stem (MESS) of *Solanum sisymbrifolium*^[25,26]. A commercial sample of fluconazol^[27] was used as a standard and Potato-Dextrose Agar Media was used as culture medium.

The petri dishes filled to depth of 4.5 mm with potato dextrose agar medium and were placed on a level surface so as to ensure that the level of the medium was of uniform thickness. The petri dishes were sterilized at 160-170 $^{\circ}$ C for 1 hour before use.

Small sterile borer of uniform size having an internal diameter of 6-8 mm and made up of stainless stell was placed at 10cm height. Six holes were made in the medium with the sterile borer. Two holes for MELS, MESS each, one hole for positive control (Fluconazol) and one for solvent control dimethyle sulfoxide (DMSO). Solutions of the standard and the extract being examined were prepared in five different concentrations (10 mg/ml, 20 mg/ml, 30 mg/ml, 40 mg/ml and 50 mg/ml) using sterile DMSO as a solvent. For experiment 50 ul of the solution of fluconazol, DMSO, MELS and MESS were filled with the help of micro pipette. The zones of the inhibition were measured in diameter (CM) produced around the hole after incubation at 37 ° C for 24 hours. The experiment was done in triplicates. The results were expressed in diameter of inhibition zones \pm S.E.M.

Result and Discussion

The results of present study are shown in Table 1. The zone of inhibition of MESS with five different concentrations were found to be comparable with that of standard drug in case of *A.niger*, *A.flavus*, *A.xylinium* and *C.albicans*, reflecting the potency of extract against these pathogens, whereas diameter of zone of inhibition of MELS with five different concentrations were found to be far less then standard drug. In case of *A.niger*, when the concentration of the 40 and 50 mg/ml of MESS were used, the diameter of zone of inhibition, was more compare to other concentrations used, but in the cases of *A.xylinium*, *A.flavus* and *C.albicans*, it was observed that increase in concentration inevitably increases the diameter of zone of inhibition.

The use of methanol as a solvent for exctraction and subsequent good antifungal activity exhibited by the MESS extract.

Name of the	Concentration	Zone of	Zone of	Zone of
organism	(mg/ml)	inhibition (cm)	inhibition (cm)	inhibition (cm)
or guinsin	(with MELS	with MESS	with Fluconazol
Aspergillus	10	1.13	1.87	2.01
niger	20	1.21	1.83	2.0
_	30	1.26	1.78	2.2
	40	1.39	1.89	2.0
	50	1.433 ± 0.88	1.9 ± 0.50	2.0 ± 0.33
Aspergillus	10	1.10	1.5	1.63
flavus	20	1.19	1.76	2.01
	30	1.25	1.75	2.2
	40	1.40	1.81	2.1
	50	1.71	1.92	2.1
Aspergillus	10	1.02	1.48	1.63
xylinium	20	1.31	1.74	1.93
	30	1.39	1.80	2.01
	40	1.42	1.84	2.2
	50	1.55	1.89	2.0
Candida	10	1.00	1.75	2.2
albicans	20	1.21	1.71	2.01
	30	1.30	1.83	2.01
	40	1.36	1.90	2.0
	50	1.43	1.94 ± 0.50	2.1

Table 1: The concentration of zone of inhibition values at different concentrations of 80% of methanolic extact of leaf and stem of *Solanum sisymbrifolium* Lam. with Fluconazol as standard drug

MESS: Methanolic extract of Stem of *Solanum sisymerifolium* MELS: Methanolic ectract of leaves of *Solanum sisymerifolium*

Conclusion

Almost all the antifungal agents, in-use shown toxic side $effects^{[28]}$ and relatively expensive. In search of efficacious, less toxic and economical antifungal agent this study was performed and it was found that 80% metahnolic extract of stem of *S.siosymbrifolium* Lam. (MESS) posseses good antifungal activity and MELS posseses less antifungal activity when compared with Fluconazol. The future prospective includes isolation and characterization of active constituents responsible for anti-inflammatory activity of the plant.

Acknowledgments

The authors are thankful to the Head of Department of Pharmaceutical Sciences, Saurasthra University, Rajkot for his extended help.

Newsletter

Vaghela *et al.*

References

- 1. Jorgensen JH, Turnidge JD and Washington JA, antimicrobial susceptible test: Dilution and Disk Diffusion methods, *In*: Manual of Clinical Microbiology, Washington DC, ASM press, 1999:1526-1530.
- Bhattacharya P.K. and Krishnendu Sarkar, (1998): Flora of West Champaran District, Bihar / / P.K. Bhattacharyya and Krishnendu Sarkar. Botanical Survey of India, Culcutta, 1998; 5: 581.
- 3. Islam M. Flora of Weeds of North East India, 1st Edition. 1996: 231.
- 4. Gaqmble J.S. Flora of Presidency of Madras. 1921; 2: 121.
- 5. Fyson P.F. The flora of the Nilgiri and Pulney Hill-tops. 1975; 3:111.
- 6. Fyson P.F. Flora of South Indian Hill Stations.1977; 1: 201.
- 7. Dr. Chunekar KC, Dr. Pandey G. Bhavprakash Nighantu, Varanasi.1999.
- 8. Singh AP, Gupta A. Bhavapraksha Nighantu, Chaukhambha Orientalia, New Delhi. 2007.
- 9. González Torres DM. Catalogo de Plantas Medicinales Usadas en el Paraguay, El País, Asunción. 1984; 312.
- 10. Filipoy AJ, A new steroidal saponin from *Solanum sisymbriifolium*, J of Ethnopharmacology. 1994; 44(3): 181.
- 11. Perez C, Anesini C, Inhibition of pseudomonas aeruginosa by Argentinean medicinal plants. Fitoterapia. 1994; 65: 169.
- 12. Siddiqi TO, Ahmad J, Khan SU, Jayed K, Khan MSY. Philipp J Sci. 1990;119:41.
- 13. Duke JA, Vasquez R, Amazonian Ethnobotanical Dictionary. CRC press, Boca raton, Ann arbor, London, 1994; 181.
- 14. Simoes CMO, Falkenberg M, Auler Mentz L, Schenkel EP, Amoros M, Antiviral activity of south brozilian medicinal plant extracts, Phytomedicine 1999; 6: 205.
- 15. Hnatsczyn O, Arenas P, Moreno AR, Rondina RDV, Coussio JD, preliminary phytochemical study of Paraguayan medical plants, plant regulating fertility from medicinal folkfore, Rev Soc Cient (Asunción) 1974; 14: 23.
- 16. Martínez-Crovetto R, Fertility regulating plants used in popular medicine in northeasten argentina, Parodiana 1981; 1: 97.
- 17. Ibarrola DA, Ibarrola MH, Vera C, Montalbetti Y, Ferro EA, Hypotensive effect of crude root extract of *solanum sisymberiifolium* (solanaceae) in normo and hypertensive rats, J of Ethnopharmacol 1996; 54: 7.
- 18. Ibarrola DA, Hellión-Ibarrola MC, Montalbetti Y, Heinichen O, Alvarenga N, Figueredo A *et al.*, Isolation of hypotensive compounds from *solanum sisymbriifolium*. *Lam.*, J Ethnopharmacol 2000; 70: 301.
- 19. Chand R, Kumar S, Sharma AK, Srivastava L, Indian Drugs 1995; 32:362.
- 20. Evans WC, Somanabandhu A, Nitrogen containing non steroidal secondary metabolites of *Solanum, Cyphomandra*, Lycianthes & Margaranthus, Phytochemistry 1980; 19: 2351.
- 21. Maldoni BE, Alkaloids en dos species de Lycium, An Asoc Quim Argent 1984; 72: 265.
- 22. Mazundar BC, Sci Cult 1984; 50: 122.
- 23. Chakravarty AK, Mukhopadhyay S, Saha S, Pakrashi SC, A neolignan and sterols in fruits of *solanum sisymriifolium*, Phytochemistry 1996; 41: 935.
- 24. Soxhlet F, Die Gewichtsanalytische Bestimmung Des Milchfettes, Polytechnisches J (Dinger's) 1879; 232: 461.
- 25. Barry AL, Lea, The antimicrobial suspectibility test, *In*: Principle and Practices by and Fibiger Publication, Philadelphia 1976: 907-909.
- 26. Pelczar MJ, Chan ECS, Krieg NR, "Microbiology" Int. Edn. McGraw Hill, New York, 1993: 578.
- 27. Purai Pandiyan V, Ignacimuthu S, Antibacterial and antifungal activity of *Cassia fistula* L, an ethanomedicinal plant, Journal of Ethanopharmacology 2007; 112: 590 -594.
- 28. Albengers HE, Loui L, Tilement JP, Systemic antifungal agents: drug Interaction of Clinical Significance, Drug Safety 1998; 18(2): 83-97.