

Identification of Mushroom Lectins and Its Medicinal Properties

G. DHAMODHARAN, S. MIRUNALINI*.

Department of Biochemistry and Biotechnology, Faculty of Science, Annamalai University,
Tamil Nadu, India; mirunasankar@gmail.com

Summary

This review summarizes the existing information about mushroom lectins. Lectins are abundant in nature like plants, animals, fungi, bacteria and viruses. They are multivalent proteins or glycoproteins of non-immune origin that bind specifically and reversibly to carbohydrates, resulting in agglutination of cells or precipitation of glycoconjugates. The properties of lectins have been known for some time and extensively studied. Their ability to cause disease and to function as markers which delineate specific pathological processes has been reported, but until recently, the role of lectins in the treatment of disease has only been suggested by a variety of sources.

Keywords: lectin, cancer, mushroom, fungi and *Agaricus bisporus*.

***Address for Correspondence:**

Mirunalini Shankaran,
Department of Biochemistry and Biotechnology,
Annamalai University,
Annamalai Nagar-608002,
Tamil Nadu,
India.
Telephone number: 09442424438,
E-mail: mirunasankar@gmail.com

Introduction

The word lectin is from the Latin *legere* which means "to bind" or to "pick and choose." Lectins were first isolated in 1888 by Stillmark at Estonia University. Lectins are found in most plants but are particularly high in legumes and grains (1). Seafood such as shellfish, eel, halibut and flounder also has high lectin contents. The amount of lectin concentration generally accounts for 1% to 3% of the protein content of the specific food, and in the case of plants, the amount is dependent upon the degree of plant maturation (2).

Lectins are non-immunologic protein-polysaccharide molecules having a strong binding affinity for the complex carbohydrates which are abundant on cell surfaces (3). Lectins contain one or more sites specific to carbohydrate binding called Carbohydrate Recognition Domains (CRD) (4,5). Lectins may interact with carbohydrates through hydrogen bonds, metal coordination, Van der Waals, and hydrophobic interactions (6). Basically, hydroxyl groups on sugar molecules can serve as both a donor and an acceptor to cooperate in hydrogen bonds (7).

The binding nature of Lectins is similar to antibodies, forming an irreversible covalent bond. An example of a high molecular weight polysaccharide which conveys a protective effect is arabinogalactan, found in a variety of foods and herbal medicines (8). This class of molecules has been shown to occupy the binding sites of various microorganisms, preventing them from attaching to cellular surfaces and making it easier for the immune system to eliminate them. Lectins are multivalent proteins or glycoproteins of non-immune origin that bind specifically and reversibly to carbohydrates, resulting in agglutination of cells or precipitation of glycoconjugates (9).

Lectins play a crucial role in diverse biological processes, particularly in host defense mechanisms, inflammation, and metastasis (10). Owing to their binding specificities, lectins are employed in a number of biochemical and clinical research areas (11). Lectins are known to cause a number of biological effects including lymphocyte proliferation (blastogenesis) and the induction of cytokine production as well as having the ability to inhibit specific antibody stimulated T cell activity (12).

In recent years, mushroom lectins have become of more interest, mainly due to the discovery of some of these lectins exhibiting potent biological activities. The lectins from

Agaricus bisporus, *Ganderma lucidum*, *Volvariella volvacea*, *Boletus satanas* Lenz, *Flammulina velutipes*, *Lentinus edodes*, and *Agrocybe cylindracea* exhibit potent mitogenic activities (13). In addition, some lectins including mushroom lectins express potential activities such as immunoenhancing, vasorelaxing, hypotensive, and antimicrobial activities (14). *Agaricus bisporus* lectin shows its antiproliferation activity against human colon cancer cell lines HT29 and breast cancer cell lines (MCF-7) (15). *Volvariella volvacea* lectin possesses antitumor activity to sarcoma S-180 cells (16). These clearly indicate that mushroom lectins might be employed as drugs or therapeutic reagents for pharmaceuticals. Mushrooms are valuable source of lectins for drug discovery.

Sources of lectin

Lectins are widely distributed in nature, and occur in diverse organisms ranging from fungi, plants, animals, bacteria, and viruses (17).

Fungi

Several fungi can express high levels of saline-soluble and low molecular weight lectins. The parasitic fungus, *Arthrobotrys oligospora*, contains a multi specific lectin that can bind to fetuin and mucins (18). Recently, fungal lectins especially from either mushrooms or filamentous fungi have been the focus of research. The high content of lectins in mushrooms has been detected in diverse species of genera *Lactarius*, *Russula*, *Boletus*, *Phallus*, and *Hygrophorus*. *Hemolysins* are found in families *Hygrophoraceae* and *Strophariaceae*; in genera *Amanita*, *Mycena*, *Agrocybe*, *Oudemansiella*, *Hebeloma*, and *Gymnopilus*; and in many ascomycetes (19). Lectins were localized on caps, stipes, and mycelia of mushrooms, and variations in lectin contents occurred upon their carpophore ages and the time and place of harvest (20). Lectins from *Pleurotus cornucopiae* and *Tricholoma mongolicum* have been reported to be isolated from mycelia (21). Surprisingly, the lectin from *Pleurotus cornucopiae* mycelium could be detected only in dikaryotic, not in monokaryotic mycelium, and it disappeared during the formation of fruit bodies (22).

Lectin from Mushroom

Over 2,500 different mushrooms grow in the wild around the world. Mushrooms have been a part of the human diet for thousands of years. They also have been used frequently in homeopathic medicine. Mushroom consumption has been markedly increasing throughout the world and involves a variety of species. Various edible mushrooms are consumed for enjoyment as well as their health benefits such as containing relatively few calories and relatively high amounts of vegetable proteins. Their fruiting bodies, on a dry weight basis, contain about 39.9% carbohydrate, 17.5% protein and 2.9% fats, with the rest consisting of minerals (23).

Mushroom lectins, with an emphasis on those from the following species which have been most extensively characterized including various *Agaricus species*, *Ganoderma lucidum*, *Amanita pantherina*, *Boletus satanas*, *Coprinus cinereus*, *Flammulina velutipes*, *Grifola frondosa*, *Hericium erinaceum*, *Ischnoderma resinatum*, *Lactarius deterrimus*, *Laetiporus sulphureus*, *Tricholoma mongolicum* and *Volvariella volvacea* (24). It is noted that the mushroom lectins exhibit a diversity of chemical characteristics. Some of them are monomeric, whereas others are dimeric, trimeric or tetrameric. Studies on immunomodulatory and antitumour, cytotoxic activities have been carried out on lectins from *Agaricus bisporus*, *Boletus satanas*, *Flammulina velutipes*, *Ganoderma lucidum*, *Grifola frondosa*, *Tricholoma mongolicum* and *Volvariella volvacea* (25).

Molecular structure of mushroom lectin

Lectin is a heterogeneous group of oligomeric protein that varies widely in size, structure, molecular organization, as well as in the constitution of their binding sites (26). Depending on the carbohydrate binding specificity of lectins, affinity chromatography can be effectively used for their purification (27).

Cell agglutination

Each lectin molecule, which generally contains two or more carbohydrate binding sites, can interact with cells by combining to sugars on their surface, thus, cross linking the cells and resulting in the phenomena of cell agglutination and their subsequent precipitation (34). So the red blood cell agglutination or hemagglutination of lectins is the major attribute of these proteins, and used routinely for their detection and characterization. Lectins also form cross-links between polysaccharides or glycoprotein molecules in solution, and induce their precipitation reaction (35).

Antiviral activity

Wood et al. (1999) proposed that monocot mannose-binding lectins, e.g. *Liliaceae*, *Amyryllidaceae*, and *Orchidaceae* lectins, exhibit anti-retroviral activity. That is possible for applications in crop protection field (36). *Pleurotus ostreatus* lectin (POL) has also been tested for its ability to inhibit HIV-1 reverse transcriptase. No antiviral activity has been displayed but instead POL exerted a powerful antitumour activity (37).

Agaricus Blazei stimulates lymphocyte T-cell and Helper T-cell production. The polysaccharide contained in *Agaricus Blazei* stimulates production of interferon and interleukin that indirectly function to destroy and prevent the proliferation of cancer cells. Also, *Agaricus Blazei* turned out to be a very powerful antiviral agent preventing viruses from entering tissues.

Antibacterial activity

Agaricus bisporus and *Pleurotus sajor caju* have been assayed *in vitro* for their anti-microbial activities using aqueous and organic solvent extracts. It has been shown that *Escherichia coli* 390, *Escherichia coli* 739, *Enterobacter aerogenes*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* were most sensitive to aqueous, ethanol, methanol and xylene extracts of these mushrooms (Tambekar et al. 2006).

Antifungal activity

Extracts from fermented broth and mycelium of 15 strains of *Lentinus edodes* have been shown to be active against gram-positive and gram negative bacteria, yeasts and mycelial fungi,

including dermatophytes and phytopathogens. The strains differed by the set of the organisms susceptible to the action of the extracts. Strains of *L. edodes* combining marked antibacterial properties and high yields of water soluble polysaccharides were screened. (39, 40)

Phagocytes

Mammalian phagocytes express a wide variety of surface lectins that mediate detection of self and foreign carbohydrates, and these receptors cooperate in detection of microbes. Zymosan is a stimulatory cell wall extract of *Saccharomyces cerevisiae* primarily containing β -glucans as well as other components such as mannans, mannoproteins, and chitin (41, 42).

Mitogenic stimulation of lymphocytes

Fresh fruiting bodies of the wild ascomycete mushroom (*Xylaria hypoxylon*) a lectin with N-terminal sequence resemblance to a part of *Aspergillus oryzae* genome. The lectin exhibited highly potent antiproliferative activity toward tumor cell lines, and exerted a potent anti-mitogenic action on mouse splenocytes. The hemagglutinating activity of the lectin was inhibited by inulin and xylose. The distinctive features of this lectin comprise unique sugar specificity, and highly potent hemagglutinating, antiproliferative and anti-mitogenic activities. *X. hypoxylon* lectin differs in molecular mass, N-terminal sequence and sugar specificity from previously reported ascomycete mushroom lectins.

The hemagglutinating activity of pinto bean lectin was stable within the pH range of 3–12 and the temperature range of 0–70 °C. By using the (3H-methyl)-thymidine incorporation assay, it was shown that the lectin had the ability to evoke a mitogenic response from murine splenocytes but it did not inhibit proliferation of L1210 leukemia cells. The pinto bean lectin inhibited HIV-1 reverse transcriptase.

A novel lectin having specificity towards a complex glycoprotein asialofetuin was purified from tubers of *Arisaema flavum* (Schott.) In Oucetrlony's double immunodiffusion, the antisera raised against *A. flavum* lectin showed distinct lines of identity with those of other *araceous* lectins. AFL showed potent mitogenic activity towards BALB/c splenocytes and human lymphocytes in comparison to Con A, a well-known plant mitogen. AFL also showed significant in vitro antiproliferative activity towards J774 and P388D1 murine cancer cell lines. (43,44).

Antiproliferative activity and cytotoxicity

Lectins, in particular mushroom lectins, have recently been shown to be of great interest since they have been reported as potential anticancer reagents that can seek out and stop multiplication of cancer cells (45). For example, *Agaricus bisporus* lectin shows antiproliferation activities against human colon cancer cell lines HT29 and breast cancer cell lines MCF-7. The Gal β 1-3GalNAc-binding lectin (ABL) elicits a pronounced dose-dependent decline of ³Hthymidine incorporation to these cancer cells with maximal effects of 87% and 50% for HT29 and MCF-7 respectively at 25 μ g/ml in serum free medium (46).

Chenguang ZHAO et al reported the anticancer activity of a novel lectin from the edible mushroom *A. aegerita*. The lectin is active against several kinds of tumour cell lines and significantly inhibits the growth of S-180 cells *in vivo*. It is well known that the antitumour mechanism of lectins may come from their immunomodulatory activity. Determining the detailed mechanism of action of AAL and exploring its toxicity and pharmacokinetics may lead to the development of a novel antitumour drug.

Biocontrolling agents

Since some monocot mannose-binding lectins display the potent host defense activity to bacteria that attack plants and even to insects, it is possible for the application of these plant lectins in crop protection against insects and nematodes (47). Additionally, monocot lectins from bulbs often exhibit anti-retroviral activity that might be investigated as human immunodeficiency virus (HIV) drugs (48).

Cancer diagnosis

Lectins have become a well-established means for understanding varied aspects of cancer and metastasis. Evidence is now emerging that lectins are dynamic contributors to tumor cell recognition (surface markers), cell adhesion and localization, signal transduction across membranes, mitogenic stimulation, and augmentation of host immune defense, cytotoxicity, and apoptosis.

In 1963, Aubb, Burger and others discovered that a plant lectin, wheat germ agglutinin, has selectively agglutinating property to murine tumour cells. It has been revealed that neoplastic

cells are differing from normal cells at the glycoconjugates on the cell surface (49, 50). The edible mushroom lectin from *Agaricus bisporus* (ABL) has antiproliferative effects on a range of cell types. This investigation was undertaken to test whether it might have inhibitory activity on Tenon's capsule fibroblasts in *in vitro* models of wound healing and therefore have a use in the modification of scar formation after glaucoma surgery.

Pharmaceutical products

Lectins with well-defined carbohydrate specificities are now available commercially either as free or immobilized proteins on Sepharose for the purification and isolation by affinity chromatography of glycoproteins, glycopeptides, and oligosaccharides, for example; concanavalin A, lentil lectin, pea lectin, *Phaseolus vulgaris*, *Griffonia simplicifolis*, *Ricinus communis*, and *Maackia amurensis* lectins (51).

***Agaricus bisporus* Lectin (ABL)**

Among 50 mushroom lectins, ABL is well documented because *A. bisporus* is the most popular edible mushroom in western cuisines. The lectin from the common mushroom *A. bisporus*, the most popular edible species in western countries, has potent antiproliferative effects on human epithelial cancer cells, without any apparent cytotoxicity (52). This property confers to it an important therapeutic potential as an antineoplastic agent. ABL has antiproliferative effects on a wide range of cell types. ABL caused a dose-dependent inhibition of proliferation and lattice contraction without significant toxicity (53). ABL might be especially useful where subtle modification of healing is needed, as in eye surgery for glaucoma. ABL has reversed the proliferation of colorectal and breast cancer cells in humans (54) The Galb1–3GalNAca (TF antigen)-binding lectin from the common edible mushroom (*Agaricus bisporus*) has a potent anti-proliferative effect without any apparent cytotoxicity.

Discussion

Lectins have been localized on the caps, stipes and mycelia of mushrooms, and variations in lectin content occur depending on the carpophores age and the time and place of harvest. In mushrooms, lectins probably play an important role in dormancy, growth and morphogenesis, morphological changes consequent on parasitic infection and molecular recognition during the

early stages of mycorrhization (55). Mushroom lectins and application in taxonomical, embryological and bacteriological studies, study of the modifications in membrane glycoconjugates and cancer formation, cell sorting, sorting of mutant and tumour cells and isolation of membrane and serum glycoconjugates. Guillot and Kanska (1977) cited a number of mushroom lectins as examples to illustrate their statement that affinity chromatography, ion exchange chromatography and gel filtration were used in the isolation of lectins. They concluded that mushroom lectins display differences in the number of subunits, molecular weight, carbohydrate content, amino acid composition, isoelectric point, carbohydrate specificity and specificity toward human erythrocytes (56).

The multifarious, potentially exploitable activities which some mushroom lectins including *Agaricus bisporus*, *Ganoderma lucidum*, *Flammulina velutipes* and *Volvariella volvacea* express such as mitogenic, immunoenhancing, antiproliferative, antitumour, vasorelaxing and hypotensive activities may be of interest to immunologists, oncologists and cardiologists (57). *A. bisporus* lectin (ABL) inhibits cell proliferation in a wide range of cells without cytotoxicity, suggesting that this lectin might affect a tumor cellular process fundamental to cell division (58). Most of the lectins discussed in this review originate from edible mushrooms. The not-too-infrequent inclusion of mushrooms as a dietary component, and the finding that some lectins retain their biological activity after passage through the gastrointestinal tract, should make research on mushroom lectins appealing to a lot of investigators. Many aspects of mushroom lectin biology merit examination. To mention just one or two, more information pertaining to the physiological role of lectins in mushrooms is necessary. Whether it is related to defence against assault from pathogenic organisms as in the case of plants awaits corroboration (59).

The details of the mechanism of action of mushroom lectins remain to be ascertained. Comparative research has in the past yielded valuable products. This may at least partially explain why lectin research has encompassed organisms across the animal and plant kingdoms. The continuous research on mushroom lectins may uncover information which could be manipulated for the production of pharmaceuticals and nutraceuticals will be processed shortly.

References

1. Nachbar M, Oppenheim J. Lectins in the United States Diet: A Survey of Lectins in Commonly Consumed Foods and a Review of the Literature. *The Amer JI of Clin Nutrition*. 1980; 33: 2338-2345.
2. Tatsumi M, Arai Y, Itoh T. Purification and characterization of a lectin from the shellfish, *Saxidomus purpuratus*. *J Biochem*. 1982, 91(4):1139-1146.
3. Sharon N, Lis H. Lectins proteins with a sweet tooth: functions in cell recognition. *Essays Biochem*. 1995, 30: 59-75.
4. Rini JM. Lectin structure. *Annual Review of Biophysics and Biomolecular Structure*. 1995, 24: 551-577.
5. Drickamer K. Two Distinct Classes of Carbohydrate-recognition domains in Animal Lectins. *The J Of Bio Chem*. 1988,263(20): 9557-9560.
6. Weis WI, Drickamer K. Structure basis of lectin-carbohydrate recognition. *Annual Review of Biochemistry*. 1996, 65: 441-443.
7. Zhang GQ, Sun J, Wang H X, and Ng T B. A novel lectin with antiproliferative activity from the medicinal mushroom *Pholiota adiposa*,” *Acta biochimica Polonica*, 2009, 56(3): 415–421,
8. Elgavish S Shaanan B. Lectin-carbohydrate interactions: Different folds, common recognition principles. *Trends in Biochemical Sciences*. 1997, 22: 462-467.
9. Mo H, Winter HC, Goldstein IJ. Purification and characterization of Neu5Ac α 2-6Gal β 1-4Glc/GlcNAc-specific lectin from the fruiting body of the polypore mushroom *Polyporus squamosus*. *J of BioChem*, 2000, 275 (14): 10623-10629.
10. Freed D. Lectins in Food: Their Importance in Health and Disease. *J of Nutr Med*, 1991: 2, 45-64.
11. Cash HL, C.V. Whitham , C.L. Behrendt , and L.V. Hooper . 2006. Symbiotic bacteria direct expression of an intestinal bactericidal lectin. *Science 2006*, 313 : 1126 – 1130 .
12. Imberty A, Gautier C, Lescar J, Perez S, Wyns L, Loris R. An unusual carbohydrate binding site revealed by the structures of two *Maackia amurensis* lectins complexed with sialic acid-containing oligosaccharides. *J of Bio Chem*. 2000, 275(23): 17541-17548.
13. Singh RS, Bhari R, Kaur HP. Mushroom lectins: current status and future perspectives. *Crit Rev Biotechnol*. 2010, 30(2):99-126.

14. Watrang E. Lectins inhibit the Aujeszky's disease virus-induced interferon-alpha production of porcine peripheral blood mononuclear cells. *J Interferon Cytokine Res.* 1995, 15(4): 301-8.
15. Parslew R, Jones KT, Rhodes JM, Sharpe GR. The antiproliferative effect of lectin from the edible mushroom (*Agaricus bisporus*) on human keratinocytes: preliminary studies on its use in psoriasis. *Br J Dermatol.* 1999,140(1):56-60.
16. Chenguang ZHAO1, Hui SUN1, Xin TONG, Yipeng Q. An antitumour lectin from the edible mushroom *Agrocybe aegerita*. *Biochem J.* 2003, 374: 321–327.
17. Faezeh Yazdani Moghaddam, Jamshid Darvish, Nasser Mahdavi Shahri, Abdulmir S, Siti Khalija Daud. Lectin Histochemistry Assay in Colon Tissues for Inter-species Characterization. *American J of Biochemistry and Biotechnology.* 2009, 5 (1): 7-13.
18. Irazoqui FJ, Vides MA, Nores GA . Structural requirements of carbohydrates to bind *Agaricus bisporus* lectin. *Glycobiology.* 1999,9(1):59-64.
19. Mancheño JM, Tateno H, Sher D, Goldstein IJ. *Laetiporus sulphureus* lectin and aerolysin protein family. *Adv Exp Med Biol.* 2010, 677:67-80.
20. Conrad F, Rudiger H. The lectin from *Pleurotus ostreatus*: Purification, characterization and interaction with a phosphatase. *Phytochemistry.* 1994, 36: 277±283
21. Rosen S, Bergstrom J, Karlsson KA, Tunlid A. A multispecific saline-soluble lectin from the parasitic fungus *Arthrobotrys oligospora*. Similarities in the binding specificities compared with a lectin from the mushroom *Agaricus bisporus*. *Europ J of Biochemistry.* 1996, 238: 830-837.
22. Guillot J, Kanska G. Lectins in higher fungi. *Biochemical Systematics and Ecology,* 1997, 25: 203-230.
23. Yang N, Li DF, Feng L, Xiang Y, Liu W, Sun H, Wang DC. Structural basis for the tumor cell apoptosis-inducing activity of an antitumor lectin from the edible mushroom *Agrocybe aegerita*. *J Mol Biol.* 2009, 387(3):694-705.
24. Brechtel, R., Watzig, H., and Rudiger, H. The lectin from the mushroom *Pleurotus ostreatus*: a phosphatase-activating protein that is closely associated with an alpha-galactosidase activity. *Plant Science.* 2001, 160(5): 1025-1033.
25. Ng TB, Ooi VEC. Lectin activity in fruiting bodies of the edible mushroom *Tricholoma mongolicum*. *Biochemistry and Molecular Biology International.* 1998, 44(1): 135-141.

26. Oguri S, Ando A, Nagata Y. A novel development stage-specific lectin of the basidiomycete *Pleurotus cornucopiae*. *J of Bacteriology*. 1996 178 (19): 5692-5698.
27. Olausson J, Tibell L, Jonsson BH, Pahlsson P. Detection of a high affinity binding site in recombinant *Aleuria aurantia* lectin. *Glycoconj J*. 2008, 25(8):753-762.
28. Bessler W, Goldstein IJ. Equilibrium dialysis studies on two lima bean lectins. *Arch Biochem Biophys*. 1974, 165(1):444-445.
29. Tsivileva OM, Nikitina VE, Loshchinina EA. Isolation and characterization of *Lentinus edodes* (Berk.) singer extracellular lectins. *Biochemistry (Mosc)*. 2008, 73(10):1154-61.
30. Lis H, Sharon N. Lectins: carbohydrate-specific proteins that mediate cellular recognition. *Chemical Reviews*. 1998, 98(2): 637-674.
31. Suzuki T, Amano Y, Fujita M, Kobayashi Y, Dohra H, Hirai H, Murata T, Usui T, Morita T, Kawagishi H. Purification, characterization, and cDNA cloning of a lectin from the mushroom *Pleurocybella porrigens*. *Biosci Biotechnol Biochem*. 2009, 23,73(3):702-709.
32. Crenshaw, R. W., Harper, S. N., Moyer, M. & Privalle, L. S. (1995). Isolation and characterization of a cDNA clone encoding a lectin gene from *Agaricus bisporus*. *Plant Physiology* 107, 1465±1466.
33. Yoshioka K, Sato Y, Murakami T, Tanaka M, Niwa O. One-step detection of galectins on hybrid monolayer surface with protruding lactoside. *Anal Chem*. 2010, 15, 82(4):1175-1178.
34. Li YR, Liu QH, Wang HX, Ng TB. A novel lectin with potent antitumor, mitogenic and HIV-1 reverse transcriptase inhibitory activities from the edible mushroom *Pleurotus citrinopileatus*. *Biochim Biophys Acta*. 2008, 1780(1):51-57.
35. Peumans WJ, Van Damme EJM. Lectins as plant defense proteins. *Plant Physiology*. 1995, 109:347-352.
36. Broekaert, W.F., and Peumans, W.J. Lectin release from seeds of *Datura stramonium* and interference of the *Datura stramonium* lectin with bacterial motility. In Bog-Hansen, T.C., and Van Driessche, E. 1986,5:57-65.
37. Wood, S.D., et al. (1999). Structure of the native (unligated) mannose-specific bulb lectin from *Scilla campanulata* (bluebell) at 1.7 Å resolution. *Acta Crystallographica Section D-Biological Crystallography*. D55: 1264-1272.
38. Sharon N, Lis H. Lectins: Department of Biophysics. The Weizmann Institute of Science Rehovot, Israel. 1989.

39. Pusztai A. Plant Lectins. Cambridge: The Rowett Research Institute, Aberdeen Cambridge University Press. 1991.
40. Wood SD. Structure of the native (unligated) mannose-specific bulb lectin from *Scilla campanulata* (bluebell) at 1.7 Å resolutions. *Acta Crystallographica*. 1991, 55: 1264-1272.
41. Wang H, Gao J, Ng TB. A new lectin with highly potent antihepatoma and antisarcoma activities from the oyster mushroom *Pleurotus ostreatus*. *Biochemical and Biophysical Research Communication*. 2000, 275: 810-816.
42. Doyle, R.J., and Slifkin, M. *Lectin-Microorganism Interactions*. New York: Marcel Dekker.1994.
43. Wang HX, Ng TB, Ooi VE, Liu WK, Chang ST. Actions of lectins from the mushroom *Tricholoma mongolicum* on macrophages, splenocytes and life-span in sarcoma-bearing mice. *Anticancer* 1997, 17(1A):419-24.
44. Tang NY, Yang JS, Lin JP, Hsia TC, Fan MJ, Lin JJ, Weng SW, Ma YS, Lu HF, Shen JJ, Lin JG, Chung JG. Effects of *Agaricus blazei* Murill extract on immune responses in normal BALB/c mice. *In Vivo*. 2009 ;23(5):761-766.
45. Sarangi I, Ghosh D, Bhutia SK, Mallick SK, Maiti TK. Anti-tumor and immunomodulating effects of *Pleurotus ostreatus* mycelia-derived proteoglycans. *Int Immunopharmacol*. 2006, 6(8):1287-97.
46. Kent D, Sheridan CM, Tomkinson HA, White SJ, Hiscott P, Yu L, Grierson I. Edible mushroom (*Agaricus bisporus*) lectin inhibits human retinal pigment epithelial cell proliferation in vitro. *Wound Repair Regen*. 2003, 11(4):285-91.
47. Yu LG, Fernig DG, White MR, Spiller DG, Appleton P, Evans RC, Grierson I, Smith JA, Davies H, Gerasimenko OV, Petersen OH, Milton JD, Rhodes JM. Edible mushroom (*Agaricus bisporus*) lectin, which reversibly inhibits epithelial cell proliferation, blocks nuclear localization sequence-dependent nuclear protein import. *J Biol Chem*. 1999, 274(8):4890-4899.
48. Zimecki M, Artym J, Cisowski W, Mazol I, Włodarczyk M, Glensk. Immunomodulatory and anti-inflammatory activity of selected osthole derivatives. *Z Naturforsch C*. 2009, (5-6):361-368.
49. Kenyon G. Mushroom: The new medicine. *BBC News*. 1995.

50. Wang H, Ng TB, Ooi VEC. Lectins from mushrooms. *Mycological Research*. 1998, 102(8): 897-906.
51. Wright LM, Van Damme EJM, Barre A, Allen AK, Leuven FV, Reynolds CD, Rouge P, Peumans WJ. Isolation, characterization, molecular cloning and molecular modeling of two lectins of different specificities from bluebell (*Scilla campanulata*) bulbs. *Biochemical Journal*. 1999, 340: 299-308.
52. Wu AM, Wu JH, Herp A, Liu JH. Effect of polyvalencies of glycotopes on the binding of a lectin from the edible mushroom, *Agaricus bisporus*. *Biochem J*. 2003, 15(371):311-20.
53. Batterbury M, Tebbs CA, Rhodes JM, Grierson I. *Agaricus bisporus* (edible mushroom lectin) inhibits ocular fibroblast proliferation and collagen lattice contraction. *Exp Eye Res*. 2002, 74(3):361-70.
54. Lu-Gang Yu‡, David G. Fernig§, Michael R. H. White§, David G. Edible Mushroom (*Agaricus bisporus*) Lectin, Which Reversibly Inhibits Epithelial Cell Proliferation, Blocks Nuclear Localization Sequence-dependent Nuclear Protein Import*. *J bio che*. 1999, 274: 4890-4899.
55. Han CH, Liu QH, Ng TB, Wang HX. A novel homodimeric lactose-binding lectin from the edible split gill medicinal mushroom *Schizophyllum commune*. *Biochem Biophys Res Commun*. 2005, 14: 252-7.
56. Yu L, Fering D, Smith J, Milton J, Rhods J. Reversible Inhibition of Proliferation of Epithelial Cell Lines by *Agaricus bisporus* (edible mushroom) Lectin. *Cancer Res*. 1983, 38: 289-350.
57. Shi YL, James AE, Benzie IF, Buswell JA. Mushroom-derived preparations in the prevention of H₂O₂-induced oxidative damage to cellular DNA. *Teratog Carcinog Mutagen*. 2002; 22(2):103-11.
58. Chrispeels MJ, Raikhel NV. Lectins, lectin genes, and their role in plant defense. *The Plant Cell*. 1991, 3: 1-9.
59. Cash, H.L., C.V. Whitham , C.L. Behrendt , and L.V. Hooper . Symbiotic bacteria direct expression of an intestinal bactericidal lectin. *Science*. 2006, 313: 1126 – 1130.