PRELIMINARY PHYTOCHEMICAL AND ANTI-MICROBIAL EVALUATION OF DIFFERENT EXTRACTS OF LEAVES OF ACACIA MONILIFORMIS GRISE

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Summary

Acacia moniliformis (family Mimosaceae) commonly known as "Auri", is a fast-growing, crooked tree, native of Australia and also found in Indonesia, Papua etc. The present study aimed to explore the presence of phytochemical substance and of to know the efficacy of antimicrobial activity of different solvent extracts of leaves of Acacia moniliformis by disc diffusion method. This study reveals that chloroform extract showed significant antimicrobial activity against microorganisms used in this study.

Keywords: Acacia moniliformis, Flavonoids, Disc diffusion method, Antimicrobial activity

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Introduction

Acacia moniliformis is a fast-growing, crooked tree which is used as a source of firewood and charcoal production. It is grown upto 30m height. Commonly known as "Ear leaf acacia" or ear pod wattle belonging to the family Mimosaceae Medicinally it was found to have antimutagenic, chemopreventive (1) ,antihelmintic (2)antioxidant property(3) and sperm immobilizing activity(4). This study explored phytochemicals present in various extracts of leaf parts and antimicrobial activity of different extracts, clarifies the preliminary information of the leaf parts of the species.

Materials and Methods

Plant collection and authentication

The leaves of *Acacia moniliformis* was collected from Vatticherrikkuru, A.P. India. The plant specimen was authenticated by the Botanist Dr. S.M.Khasim, Department of Botany and microbiology, Acharya Nagarjuna University, Guntur. A.P. and voucher specimen was deposited on the department for future reference.

Preparation of plant Extracts

Fresh plant material was collected in bulk, washed under running tap water to remove adhering material, dried under shade and pulverized in a mechanical grinder. The coarse powder was passed through sieve no.60#. The different extracts were carried out by using 1 kg of dried coarse leaf powder of *Acacia moniliformis* with the help of Soxhlet extractor. Different solvents used for the extractions are with petroleum ether (60-80°C), n-Hexane, chloroform, acetone, ethanol and water according to the polarity basis. They were then filtered through muslin cloth and were concentrated under reduced pressure to obtain residues.

Preliminary Phytochemical studies

Preliminary phytochemical screening of the leaves extracts of Acacia auriculiformis was performed as per standard procedure.(5,6)

Test microorganisms

Microorganisms like Klebsiella pneumoniae, Staphylococcus subtillis, Escherichia coli and Staphylococcus aureus were used for the screening of antimicrobial activity.

Evaluation of antimicrobial activity

The antimicrobial activities of different extracts of Acacia auriculiformis leaves were determined at different concentrations (10µg/ml, 15 µg/ml,20µg/ml in dimethyl sulfoxide) against microbes by agar diffusion technique using a paper disc. Microbial cultures were firmly swept over the agar (nutrient agar medium) plate using sterile cotton swab to make uniform culture lawns. The saturated disc of different concentration of test compounds was placed on agar plates. Allowed the plates to stand at room temperature for 30 minutes (pre-incubation). Inoculated plates were incubated at 37°C for 24 hr. The inhibition zones were recorded with the help of scale and the result is summarized in Table 2. Sterile dimethyl sulfoxide without plant extracts served as negative control DMSO (20µl/disc).(7,8)

Results and Discussion

The different extracts explored different phytoconstituents such as Glycoside, proteins, tannins and resins majorly. Flavonoids, Saponins , waxes, Carbohydrates and are present only trace amount. The alkaloids, mucilage, volatile oils, steroids are absent in all the above extracts. Results were tabulated in table no 1. The antimicrobial activity of the extracts at different concentration was determined. All the six extracts at different concentration (10µg/ml,15 µg/ml, 20µg/ml in dimethyl sulfoxide) exhibited antimicrobial activity against all microbial strains tested. Results were tabulated in table no 2. Chloroform extract exhibited comparably a high degree of activity than the other extracts (petroleum ether, n-hexane, ethyl acetate, ethanol and water). In the present antimicrobial screening, 20µg/ml of all the extracts were taken for the evaluation of the activity. Chloroform extract (20µg/ml) was found to possess the broadest antimicrobial activity and, n-hexane, ethyl acetate, petroleum ether was found to be moderately active, while ethanol and water extract did not show considerable activity. The ethanolic extract was most effective against K.Pnemonia and S.Aureus with diameter of zone of inhibition 1.1cm and 1.2cm at a concentration of 20µg/ml respectively. It was least effective against E.Coli (Gram negative bacteria). Amongst the Gram negative bacteria, the ethanolic extract showed less activity against all the four microorganism. Phytochemical studies on *Acacia moniliformis* leaves revealed the presence of carbohydrates, glycosides, tannins, Saponins, and phenolic compounds. The antimicrobial activity of *Acacia moniliformis* may be due to one/more group of above phytoconstituents.

Preliminary phytochemical screening of different extracts of leaves of Acacia moniliformis grise

Table: 1

TEST	PET.ETHER	N-HEXANE	CHLOROFORM	ACETONE	ETHANOL	WATER
Alkaloids						
Anthraquinone Glycosides	2			++		++
Cardiac Glycosides			++	++		++
Saponin Glycosides						++
Coumarian Glycosides			++		++	
Carbohydrates	·			++		
Flavanoides						++
Protein	++		++		++	
Amino acids	++		++	`	++	
Steroids						
Tannins					++	++
Resins					++	++
Volatile oil						
Waxes	++					
Mucilage						

⁺⁺ present, -- absent

Anti-microbial evaluation of different extracts of leaves of Acacia moniliformis grise

Table:2 Average zone of inhibition (cm)

	Pet ether		n-hexane		Chloroform		Ethyl acetate		Ethanol			Aqueous		Ι	OMSO			
Micro Organism																		
/disc 20	10μ 15με	g 20µg	10μg	15µg	20μg	10με	ց 15με	g 20μg	10μ	ց15μք	g 20μg	10μg	; 15με	g 20μg	10μg	15μg	g 20μg	20μ1
K.Pneumor	nia 0.7 0.9	9 1.1	0.7	1.0	1.3	0.9	1.1	1.4	0.7	1.0	1.3	0.7	0.9	1.1	-	-	-	-
B.Subtilis	0.9 1	.1 1.3	0.8	1.1	1.3	0.7	1.2	1.4	0.8	1.0	1.2	0.7	0.9	1.0	-	-	-	-
E.Coli	0.8 0	.9 1.2	0.7	0.9	1.2	0.8	0.9	1.0	0.8	0.9	1.2	-	-	-	0.6	0.7	0.9	-
S.Aureus	0.8 1	.0 1.3	0.9	1.1	1.4	0.9	1.0	1.2	0.7	0.9	1.2	0.7	0.8	1.2	-	-	-	-

Conclusion

The present study reveals preliminary information of leaves of *Acacia moniliformis*. From the results, it can be concluded that the *Acacia moniliformis* leaves extracts found to be used as narrow-spectrum antimicrobial agent after extensive investigation. Further work will emphasize the isolation and characterization of active principles responsible for antimicrobial activity of leaf extracts of *Acacia moniliformis*.

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References

- 1)_Kamaljit Kaur, Saroj Arora, Michael E. Hawthorne, Swayamjot Kaur, Subodh Kumar, and Rajendra G. Mehta_Drug and Chemical Toxicology, 2002, Vol. 25, No. 1: Pages 39-64
- 2) Sinha Babu SP, Sarkar D, Ghosh NK, Saha A, Sukul NC, Bhattacharya S "Enhancement of membrane damage by saponins isolated from Acacia auriculiformis". Jpn J Pharmacol. 1997 Dec;75(4):451-4

- 3) A. Pakrashi, H. Ray, B.C. Pal, S.B Mahato "Evaluation of antioxidant potential of ethyl acetate extract/fractions of *Acacia auriculiformis* A. Cunn" Food and Chemical Toxicology ScienceDirect Volume 45, Issue 7, July 2007, Pages 1216-1223
- 4) A. Pakrashi, H. Ray, B.C. Pal, S.B Mahato "Sperm immobilizing effect of triterpene saponins from acacia auriculiformis" ScienceDirect Volume 43, Issue 5, May 1991, Pages 475-483
- 5) Kokate CK, Purohit AP, Gokhale SB, Pharmacognosy, 39th Edition, Nirali Prakashan, Pune, 2005:607-611.
- 6) Harborne JB, Phytochemical methods, A Guide to Modern Techniques of Plant Analysis, Edition 2, Chapman & Hall, London, 2005, 85, 86, 126, 196.
- 7) Gulzar A.1*., Manjul P. S. 1, Anita S. 1, Upendra K. 1, Yatendra K. 2Preliminary phytochemical and antimicrobial screening of leaves extract of *Acacia catechuWilld Journal of Pharmacy Research* 2010;3(11):2583-2584.
- 8) Jaraid E, Edwin S, Pandian S, Damodaran S, Jain P, Patel J, Antimicrobial activity of leaves of *Lantana camera Linn* and *Calotropis procera R. Br*.Indian drugs 2008; 45(9): 736-738.