

## DOES *SYZYGIUM CUMINI* POSSESS SIGNIFICANT PHARMACOLOGICAL EFFECTS? AN OVERVIEW

Saleem, U.<sup>1</sup>; Ali, N.<sup>1</sup>; Ahmad, B.<sup>2\*</sup>

<sup>1</sup>Faculty of Pharmaceutical Sciences, GC University, Faisalabad - Pakistan.

<sup>2</sup>Riphah Institute of Pharmaceutical Sciences, Riphah International University, Lahore-Pakistan.

\*[ahmadbprof@gmail.com](mailto:ahmadbprof@gmail.com)

### Abstract

*Syzygium cumini* has extensive folklore uses and most of them, now, have scientific evidence also. This review expresses the medicinal significance of *Syzygium cumini* on the basis of its profound pharmacological activities.

**Key words:** *Syzygium cumini*, Folklore uses, Pharmacological activities

## Introduction

*Syzygium cumini* (*S. Cumini*) commonly called Jamun belongs to family "Myrtaceae" which comprises about 150 genera and 3600 species and found all over the Indian sub-continent [1-3]. Chemical constituents found in flowers are quercetin, kaemferol, iso-quercetin, myricetin, quercetin-3-D-glucoside, myricetin-3-L-arabinoside, oleanolic acid, dihydromyricetin, eugenol-triterpenoid A, acetyl oleanolic acid and eugenol-triterpenoid B [5]. Roots contain isorhamnetin 3-O-rutinoside and flavonoid glycosides [6]. Leaves are rich in quercetin, acylated flavonol glycosides [7], myricetin, myrcetin, myrcetin 3-O-4-acetyl-L-rhamnopyransides [8], esterase, triterpenoids [9], tannins and galloyl carboxylase [10]. Fruits are source of glucose, raffinose, fructose [11], mallic acid [12], citric acid, anthocyanins [13], gallic acid, malvidin-3-laminaribioside, delphinidin-3-gentioside, petunidin-3-gentioside [14], malvidin [15], cyaniding diglycoside and petundin. The fruits sourness is due to gallic acid and fruits color is due to anthocyanins [14]. Bark is rich in friedelin, betulinic acid, beta sitosterol, epi-fridelanol [18], myricetin, kaempferol, ellagic acid, gallic acid [19], tannins [20], flavonoids and bergenins [21].

## Folklore uses

Bark is used in the treatment of sore throat, asthma, bronchitis, dysentery, and ulcers. It is also known to have astringent and blood purifier. Fruit has stomachic and diuretic effect [25]. Ashes of leaves are used for strengthening teeth and gums [26]. Leaves juice along with goat's milk and honey is being used for treatment of dysentery with bloody discharge [25]. Seed extract is used to treat cough, cold, skin problems such as rashes, fever, gastric and intestinal ulcers [27-28].

In all over the world, the fruits are also used for variety of the ailments including cough and ringworm infection [29-30]. This plant has been included in several herbal preparations to exploit its therapeutic potential. Several herbal formulations have been prepared in combination with this plant [31].

## Pharmacological activities

### Anti-bacterial activity

Essential oil, aqueous, methanol, and methylene chloride extracts of leaves of *S. cumini* were evaluated for the anti-bacterial activity against *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Neisseria gonorrhoeae*, *Bacillus subtilis* and *Enterococcus faecalis* using disc

diffusion method to measure minimum inhibitory concentration.. The methanol extract was found to exhibit maximum antibacterial activity [32, 33].

### Anti-oxidant activity

FRAP and DPPH in-vitro assay were used to evaluate antioxidant activity of methanol and methylene chloride extract of leaves and essential oils of *S. cumini*. Methanol extract showed greater antioxidant potential than methylene chloride extract[33].

### Anti-allergic activity

Leaves aqueous extract was tested for anti-allergic activity by performing test on male swiss webster mice, wistar rats and BALB/c mice. Histamine injection, 100µg/paw, was injected in hind paw of mice that induced significant edema. Extract was administered orally that showed 52% inhibition in edema at 50 mg/kg. Serotonin injection, 100µg/paw, was administered parenterally in hind paw of mice for induction of edema. Extract given orally showed maximum reduction (51 %) in edema at 100 mg/kg. On other hand, edema induced by platelet aggregating factor, 1µg/paw, was not affected by oral extract of *S. cumini* at doses 25, 50 or 100 mg/kg [34].

### Anti-inflammatory activity

Ethanol extract of bark was evaluated for above mentioned activity. Anti-inflammatory activity was determined at acute (Carrageenan induced paw edema), sub-acute (formaldehyde and, kaolin-carrageenan induced paw edema) and chronic (cotton pellet granulation test) levels in rats. All the tests revealed significant anti-inflammatory power of the extract [35].

### Central nervous system activity

Rota rod test and actophotometer test were performed to evaluate CNS depressant activity of methanol and ethyl acetate extracts of *S. cumini* seed in mice. Extracts were administered orally at 200- and 400 mg/kg dose levels. Both tests showed decrease in activity tested which is indicative of CNS depressant effect of extract [36].

### Anti-nociceptive activity

Formalin and hotplate tests were performed to investigate anti-nociceptive activity of hydro-alcoholic extract of *S. cumini* leaves in rats. Extract was administered i.p. at 100- and 300 mg/kg concentrations. Both the tests revealed significant dose dependent anti-nociceptive effect [37].

### **In vivo anti-diabetic activity**

Streptozotocin induced diabetic rat model was used in this study. Methanol and ethyl acetate extracts (200- and 400 mg/kg) were given orally to diabetic rats that displayed significant antidiabetic activity. Mycaminose compound, isolated from *S. cumini*, showed antidiabetic effect at 50 mg/kg oral dosing. The underlying mechanism could be secretagogue effect on beta cells of pancreas [38].

### **Antigenotoxic effect**

Ethanol and aqueous extract of *S. cumini* seeds showed antigenotoxic effect against URE and DMBA genotoxins. Both extracts were administered orally for five days prior to exposure to genotoxins [39].

### **Positive inotropic effect**

Legendroff's heart perfusion method was used to study inotropic effect of ethanol extract of *S. cumini* seeds. At 4mg/mL concentration, heart beat increased 42.85% and at 8mg/mL concentration, there was 60% increase in the heart beat indicating positive inotropic effect [40].

### **Antispasmodic activity**

Contractions were induced in the isolated rat uterus smooth muscles with KCl. Ethanol extract of *S. cumini* seeds exhibited 55% inhibition in muscle contractions that is indicative of seeds antispasmodic activity [40].

### **In-vitro antidiabetic activity**

Starch is hydrolyzed into maltose and oligosaccharides by pancreatic  $\alpha$  amylase. The inhibition of  $\alpha$  amylase results in less starch digestion which reduces the post-prandial blood glucose levels in diabetic patients. Aqueous extract of *S. cumini* seeds showed 98% inhibition of  $\alpha$  amylase activity. The compounds, tetrahydroxy flavanone and betulinic acid, were identified to be responsible for this activity [41].

### **Conclusion**

*S. cumini* possesses significant antioxidant potential that seems to be contributing towards its pharmacological effects, which make basis for folklore uses of the plant to treat a number of pathological conditions. Further studies (pharmacological/toxicological) and isolation of active principles could prove an exotic addition to the existing pool of lead compound.

### **References**

1. Chase, M.W., Reveal, J.L., A phylogenetic classification of the land plants to accompany APG III. *Bot J Linn Soc*

- 2009;161(2):122-127.
2. Gamble, J.S., Flora of the Presidency of Madras 1923.
3. Hooker, J.D., The Flora of British India. Vol. 5. L. Reeve 1890.
4. Craveiro, A.A., Andrade, C.H.S., Matos, F.J.A., et al. Essential oil of *Eugenia jambolana*. *J Nat Prod* 1983;46(4):591-592.
5. Nair, A.R., Subramanian, S., Chemical examination of flowers of *Eugenia-jambolana*. *J Sci Ind Res* 1962;21(9):457.
6. Vaishnav, M.M., Tripathy, A.K., Gupta, K.R., Flavonoid glycosides from roots of *Eugenia jambolana*. *Fitoterapia* 1992;63:259-260.
7. Mahmoud, I.I., Marzouk, M.S., Moharram, F.A., et al. Acylated flavonol glycosides from *Eugenia jambolana* leaves. *Phytochemistry* 2001;58(8):1239-1244.
8. Timbola, A.K., Szpoganicz, B., Branco, A., et al. A new flavonol from leaves of *Eugenia jambolana*. *Fitoterapia* 2002;73(2):174-176.
9. Gupta, G.S., Sharma, D.P., Triterpenoid and other constituents of *Eugenia jambolana* leaves. *Phytochemistry* 1974;13(9):2013-2014.
10. Bhatia, I.S., Sharma, S.K., Bajaj, K.L., Esterase & galloyl carboxylase from *Eugenia jambolana* (Lam.) leaves. *Indian journal of experimental biology* 1974.
11. Srivastava, H.C., Paper chromatography of fruit juices. *J Sci Ind Res B* 1953;12:363-365.
12. Lewis, Y.S., Dwarakanath, C.T., Johar, D.S., Acids and sugars in *Eugenia jambolana*. *J Sci Ind Res C* 1956;15:280-281.
13. Jain, M.C., Seshadri, T.R., Anthocyanins of *Eugenia jambolana* fruits. *Indian journal of chemistry* 1975.
14. Venkateswarlu, G., On the nature of the colouring matter of the jambul fruit (*Eugenia-jambolana*). *J Indian Chem Soc* 1952;29(6):434-437.
15. Sharma, J.N., Sheshadri, T.R., Survey of anthocyanins from Indian sources Part II. *J Sci Ind Res* 1955;14:211-214.
16. Noomrio, M.H., Dahot, M.U., Nutritive value of *Eugenia jambosa* fruit. *J Islam Acad Sci* 1996;9(1):9-12.
17. Veigas, J.M., Narayan, M.S., Laxman, P.M., et al. Chemical nature, stability and bioefficacies of anthocyanins from fruit peel of *Syzygium cumini* Skeels. *Food Chemistry* 2007;105(2):619-627.
18. Sengupta, P., Das, P.B., Terpenoids and related compounds part IV triterpenoids the stem-bark of *Eugenia jambolana* Lam. *Indian Chem Soc* 1965;42(4):255-258.
19. Bhargava, K.K., Dayal, R., Seshadri, T.R., Chemical components of *Eugenia jambolana* stem bark. *Current science* 1974.
20. Bhatia, I.S., Bajaj, K.L., Chemical constituents of the seeds and bark of *Syzygium cumini*. *Plant Med* 1975.
21. Kopanski, L., Schnelle, G., Isolation of Bergenin from Barks of *Syzygium cumini*. *Planta medica* 1988;54(6):572.
22. Kumar, A., Naqvi, A.A., Kahol, A.P., et al. Composition of leaf oil of *Syzygium cumini* L, from north India. *Indian Perfum* 2004;48:439-441.
23. Vijayanand, P., Jagan Mohan Rao, L., Narasimham, P., Volatile flavour components of jamun fruit (*Syzygium cumini* L). *Flavour Frag J* 2001;16(1):47-49.
24. Gupta, D.R., Agrawal, S.K., Chemical examination of the unsaponifiable matter of the seed fat of *Syzygium cumini*. *Sci Cult* 1970;36(5).
25. Nadkarni, K.M., Medica, I.M., revised by AK Nadkarni. *The Indian Materia Medica* 1976;1:969.
26. Kirtikar, K.R., Basu, B.D., *Indian Materia Medica*. Dehra Dun, India 1987;3:333-335.
27. Duraipandiyar, V., Ayyanar, M., Ignacimuthu, S., Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from Tamil Nadu, India. *BMC complementary and alternative medicine* 2006;6(1):1.

28. Satish, S., Mohan, D.C., Ranhavendra, M.P., et al. Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp. *Int J Agric Tech* 2007;3(1):109-119.
29. Reynertson, K.A., Basile, M.J., Kennelly, E.J., Antioxidant potential of seven myrtaceous fruits. *Ethnobotany Res App* 2005;3:025-036.
30. Jain, S.K., Dictionary of Indian folk medicine and ethnobotany. Deep publications 1991.
31. Sagrawat, H., Mann, A.S., Kharya, M.D., Pharmacological potential of *Eugenia jambolana*: A review. *Pharmacognosy Magazine* 2006;2(6):96.
32. Kannan, P., Dhasarathan, P., Efficiency of in-vitro antibacterial activity of *Syzygium cumini* phenolic extract from leaves. *Asian Journal of Pharmaceutical and Clinical Research* 2015;8(6).
33. Mohamed, A.A., Ali, S.I., El-Baz, F.K., Antioxidant and antibacterial activities of crude extracts and essential oils of *Syzygium cumini* leaves. *Plos one* 2013;8(4):e60269.
34. Brito, F.A., Lima, L.A., Ramos, M.F.S., et al. Pharmacological study of anti-allergic activity of *Syzygium cumini* (L.) Skeels. *Brazilian J Med Biol Res* 2007;40(1):105-115.
35. Muruganandan, S., Sirinivasan, K., Chandra, S., et al. Anti-inflammatory activity of *Syzygium cumini* bark. *Fitoterapia* 2001;72(4):369-375.
36. Kumar, A., Padmanabhan, N., Krishnan, M.R.V., Central nervous system activity of *Syzygium cumini* seed. *Pakistan J Nut* 2007;6(6):698-700.
37. Avila-Peña, D., Peña, N., Quintero, L., et al. Antinociceptive activity of *Syzygium jambos* leaves extract on rats. *J Ethnopharmacology* 2007;112(2):380-385.
38. Kumar, A., Ilavarasan, R., Jayachandran, T., et al. Anti-diabetic activity of *Syzygium cumini* and its isolated compound against streptozotocin-induced diabetic rats. *J Med Plants Res* 2008;2(9):246-249.
39. Arun, R., Prakash, M.V.D., Abraham, S.K., et al. Role of *Syzygium cumini* seed extract in the chemoprevention of in vivo genomic damage and oxidative stress. *J Ethnopharmacology* 2011;134(2):329-333.
40. Archana, N., Ramasamy, M., Raj, C.D., Pharmacological screening of ethanolic extract of *syzygium cumini* seed on isolated smooth muscle strip and heart. *Int J Pharm Pharmac Sei* 2012;4:108-110.
41. Karthic, K., Kirthiram, K.S., Sadasivam, S., et al. Identification of  $\alpha$ -amylase inhibitors from *Syzygium cumini* Linn seeds. *Indian J Exp Biol* 2008;46(9):677-680.
42. Anandharajan, R., Jaiganesh, S., Shankemayanan, N.P., et al. In vitro glucose uptake activity of *Aegles marmelos* and *Syzygium cumini* by activation of Glut-4, PI3 kinase and PPAR $\gamma$  in L6 myotubes. *Phytomedicine* 2006;13(6):434-441.