

**CONTROL OF BIODETERIORATION USING A FRACTION
ISOLATED FROM LEAVES OF RICINUS COMMUNIS LINN.**

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Summary

In archives, especially in those that are located in tropical climate, biodeterioration causes increasing damages every year to funds, which leads occasionally to the irreversible loss of documents heritage that is treasured in it. The different controlling methods are often chemicals that have limitations due to their costs and to the negative impact they can cause on the environment and on the substrate that receives it. In virtue of this situation, search of alternatives as the rational use of natural products that come from plants has been done. That is why the purpose of this work was to evaluate the effect of an isolated fraction from *Ricinus communis* Linn on isolated bacteria from indoor environments of archives as well as in the control of rodents. The evaluation fraction (FRC2A) was obtained using ethyl acetate extraction from an aqueous extract of leaves of *Ricinus communis*. The raticide effect was determined by the Median Lethal Dose (LD₅₀) oral administration in 70 hybrid mice B₆D₂F₁ while the bactericidal effects were evaluated by growth inhibition method by diffusion of the product on agar holes, using strains of *Enterobacter agglomerans*, *Bacillus polymixa* and *Streptomyces* sp. for this essay. The calculated LD₅₀ was 388.08 mg/Kg and the concentrations that showed the highest antimicrobial activity were 50 and 100 mg/mL. The obtained effects could be related with the presence of ricinine and different flavonoids in the evaluated fraction.

Key words: *Ricinus communis* Linn, Antimicrobial activity, Biodeterioration, Natural products, Plants extracts

Introduction

Medicinal plants are an important therapeutic aid for various ailments. Scientific experiments on the antimicrobial properties of plant components were first documented in the late 19th century (1). Their antimicrobial activity has been known for millennia since in China, India, then later in Egypt and Greece different parts of medicinal plants have been used to cure specific ailments. Also, raticide and insecticides properties have been detected in plant so many years ago (2).

Usually, in the biodeterioration control has been used chemical products (biocides) but they are toxic and frequently their degradation is difficult, being persistent in the natural environment and causing contamination of areas distant from the site of treatment. However, the use of plant in the biodeterioration control is recent (3).

The use of natural compounds, such as plant extracts that will be biodegraded more easily and be more environmentally acceptable is advantageous. Many of these compounds are phenols, terpenoids, essential oils, alkaloids, polypeptides, flavonoids and other substances (4).

The aim of this paper was to study the effect of one fraction of *Ricinus communis* on the toxicity in mice and antimicrobial activity against different bacteria isolated from indoor air of repositories of documents in National Archive of Cuba.

Materials and Methods

Isolation of the FRC2A from leaves *Ricinus communis* Linn

The leaves of *Ricinus communis* Linn were collected in Havana City (Cuba). After it were dried and milled, and the powder obtained was extracted with hot water. The extract was evaporated in vacuum until a ¼ of volume. It was extracted with EtOAc. After evaporated of the EtOAc, the residue was dissolved in hot ethanol and left to cool. The crystals thus obtained were recrystallized three times, dried at 40 °C. The ethanol solution was evaporated in vacuum and obtained FRC2A. The identification of crystals was determinate by IR spectroscopic method. Also, it was realized a chromatographic in TLC system (Kieselgel 60 F254): EtOAc: MetOH: H₂O (100:17:13); detection: ALCL₃ 5 % in MetOH, UV 366 nm to FRC2A. In it were used rutine and recinine as patron.

Determination of LD₅₀

The acute toxicity of fraction isolated (FRC2A) from the leaves of *Ricinus communis* Linn was determined in male mice hybrids B₆D₂F₁ (18 – 20 g). The animals were fasted 8 hours prior to the experiment. Animals were administered with single dose of fraction and observed for its mortality 48 hours study period toxicity (short-term). The LD₅₀ was calculated using Litchfield and Wilcoxon Methods.

Determination of the antimicrobial activity

Enterobacter agglomerans (Gram negative bacteria), *Bacillus polimixa* and *Streptomyces* sp. (both Gram positive bacteria) isolated from the indoor air of repositories of National Archive of Cuba were used. These strains were maintained in Nutrient Agar slant for 24 h. The suspensions of the bacterial strains correspond to 1 x 10⁶ CFU/ml of the Mc Farland scale.

Antimicrobial activity of the fraction FRC2A was determined by the diffusion method with holes (5). Petri dishes with 15 ml of sterile Nutrient Agar were seeded with the appropriate bacterial suspension. Then in the holes (5 mm) 10 µL of the extract of different concentration were added. One additional hole - with sterile water- was included in the test as controls. After incubation for 24 h at 28 °C, the plates were observed for zones of growth inhibition, and the diameter of these zones was measured in millimeters. Additionally, and for comparative purposes, thymol (2%) was included in the test. Each test was performed in triplicate.

Results

The identification of ricinine in the crystals was deduced from spectroscopic data in comparison with literature report. The IR spectrum showed an intense band at 2222 cm⁻¹ attributed to the nitrile group and the band relative to the carbonyl group was at 1659 cm⁻¹. In the TLC system were showed more that 7 elements predominate between they ricinine and rutine.

The LD₅₀ was estimated at 388.08 mg/Kg. for oral administration of the fraction (linear regression from a log-logit plot; r = 0.95). No effect was observed after vehicle administration. Before death the animals became exophtalmic, presented muscular tremors, and sometimes walked dragging the hid paws. In all cases of sudden death, after the animals stopped breathing their hearts continued beating for several minutes. The effects were significant at doses of 500 mg/Kg. The animals died in the first 2 hours after receive the drug (Table 1).

Table 1. Estimation of the LD₅₀ of FRC2A

GROUP	DOSE mg/Kg	UNITS	TOTAL	VALUE OBSERVED	VALUE WAITED
1	250	0	10	0.12	0.41
2	300	1	10	10	6.11
3	350	2	10	20	26.76
4	400	4	10	40	57.21
5	450	9	10	90	81.29
6	500	10	10	97.88	93.59

It was possible to obtain a high antimicrobial activity when the concentration of the fraction was the highest (100 mg/mL). The activity was decaying when the concentration of the extract diminished (Table 2).

Table 2. Average inhibition halo diameter (mm) at different concentrations of fraction FRC2A.

Concentration	<i>Enterobacter agglomerans</i>	<i>Bacillus polimixa</i>	<i>Streptomyces</i> sp.
100 mg/mL	10	5	8
50 mg/mL	10	3	6
25 mg/mL	5	0	1
12.5 mg/mL	0	0	0

Discussion

The phytochemical analysis of the fraction FRC2A showed the presence the flavonoids rutine and the alkaloid ricinine. The data obtained is consistent with literature reports. Farah (6) suggested that ricinine might inhibit the mitochondrial respiratory chain and Pérez (7) reported that flavonoids may responsible for the antimicrobial activity.

The antimicrobial activity of *Ricinus communis* had been determined previously (8-10). Some of these authors have reported that this plant has antifungal activity. However, an insignificant activity was determined by Upasani et al. (9).

In this work a moderate activity was observed. However, the activity for *Enterobacter agglomerans* was higher than other bacteria. Possibly because *E. agglomerans* is a Gram negative bacteria and it seems to be the most sensitive of all. Likewise, *Bacillus polimixa* was the most resistant in all the bacteria. This result was not surprising, in general, this bacterial genera is the most resistant than Gram positive bacteria (11) (Lennette, 1985. In the case of *Bacillus polimixa* it is worth mentioning that it is a spore-forming species.

References

1. Zaika LL. Spices and herbs: their antimicrobial activity and its determination. *J Food Safety* 1975; 9: 97-118.
2. Duke, S.O. Natural pesticides from plants. In: J. Janick and J.E. Simon (eds.), *Advances in new crops*. Timber Press, Portland, OR. 1990: 511-517.
3. Rakotonirainy M, Raison MA, Flieder F, Borchersen K. Evaluation of the Fungistatic and Fungicidal Activity of Six Essential Oils and their Related Compounds. In: *25 Years School of Conservation. The Jubilee Symposium*. Copenhagen Konservatorskolen, Det Kongelige Danske Kunstakademi. 1998: 121-130.
4. Duke, J. Dr. Duke's Phytochemical and Ethnobotanical Databases. Website. URL: <http://www.ars-grin.gov/duke/>, 2006. [Fecha de consulta 29/8/06].
5. de Paula J, Martins A. Acción antibacteriana de extractos hidroalcohólicos de *Rubus urticaefolius*. *Rev. Cubana Plant Med.* 2000; 5: 26-29.
6. Ferraz A. Pharmacological evaluation of ricinine, a central nervous system stimulant isolated from *Ricinus communis*. *Pharmacology Biochemistry and Behavior*. 1999; 63: 367-375.
7. Pérez C. Potencial medicamento contra bacterias y hongos resistentes. Entre las sierras y la farmacia: nuevo antimicrobiano de origen vegetal. 2004. Website. URL: http://www.fcen.uba.ar/prensa/noticias/2004/noticias_03may_2004.html
8. Martín M, Cortez E, Burgos C, López S, Corrales C. Efecto de extractos alcohólicos de plantas silvestres sobre la inhibición de crecimiento de *Aspergillus flavus*, *Aspergillus niger*, *Penicillium chrysogenum*, *Penicillium expansum*, *Fusarium moniliforme* y *Fusarium poae*. *Rev Iberoam Micol* 2002; 19: 84-88.
9. Upasani SM, Kotkar HM, Mendki PS, Maheshwari VL. Partial characterization and insecticidal properties of *Ricinus communis* L foliage flavonoids. *Pest Manag Sci* 2003; 59: 1349- 1354.
10. Alcalá de Marcano D., Vargas N., Pire A. Efecto de extractos vegetales y fungicidas sintéticos sobre el crecimiento micelial in Vitro de *Sclerotium rolfsii* y *Thielaviopsis basicota*. *Rev Fac Agron (LUZ)* 2005; 22: 315-323.
11. Lennette, E.H. *Manual of Clinical Microbiology*. 4th Edition Washington DC. American Association for Microbiology 1985: 978-987.