

**BIOACTIVE COMPOUNDS FROM THE FRUITS
OF *ZANTHOXYLUM LEPRIEURII*[†]**

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Summary

Four acridone alkaloids, tegerrardin **1**, 1-hydroxy-3,4-dimethoxy-*N*-methylacridone **2**, arborinine **3** and xanthoxoline **4** and a coumarin, scoparone **5** have been isolated from the fruits of *Zanthoxylum leprieurii*, used as spice in many Cameroonian traditional dishes. Compounds **3** and **4** displayed a good *in vitro* antiplasmodial activity against 3D7 strains of *Plasmodium falciparum* with IC₅₀ values of 4.5 ± 1.0 and 4.6 ± 0.6 µg/ml, respectively. Compounds **1** and **3** are good chelating agents with 90% and 61% Radical Scavenging Activity (RSA), respectively. Compound **2** showed good activity in the superoxide anion scavenging assay with 90.3% RSA. The structures of the compounds were elucidated on the basis of their spectroscopic and physical data.

Key words: *Zanthoxylum leprieurii*, acridone alkaloids, xanthoxoline, antiplasmodial, antioxidant.

Introduction

In years past, there has been a growing interest in the study of the bioactivity of phytochemical constituents in food (1). Several studies indicate that a plant-based diet protects against the development of chronic diseases such as atherosclerosis, diabetes, cancer, cirrhosis and ageing-related diseases (2). Dietary plants contain hundreds of antioxidants many of which are believed to act in a synergistic way, protecting the body against oxidative stress (3).

In a program aimed at identifying bioactive substances in diet, the fruits of *Zanthoxylum leprieurii* Guill. & Perr. (Rutaceae) (syn. *Fagara leprieurii*) which are used as spice in many traditional Cameroonian dishes (4), have been studied. *Z. leprieurii* is an under storey forest tree, 15-80 ft. high with stems covered with large broad-based thorns having sharp points (5).

Previous studies on the fruits of this plant included the antibacterial and antifungal analyses of the volatile constituents as well as the phytochemical studies of a sample collected from Nigeria (4,6,7). We report herein the antiplasmodial and antioxidant activities of compounds isolated from the ethyl acetate extract of these fruits.

Methods

Plant materials

The fruits of *Z. leprieurii* Guill. & Perr were purchased from a local market in Yaoundé (Cameroon) and identified by Mr. Nana, a Botanist at the National Herbarium of Yaoundé, Cameroon, where a voucher specimen (N^o 10669 / SRFCAM) was deposited.

Extraction and isolation of constituents

The dried fruits (1 kg) of *Z. leprieurii* were chopped in a blender and extracted with ethyl acetate (4 L) for 48 hrs. at room temperature. The filtrate was concentrated to give an orange-brown oil. Twenty grams (20 g) of the concentrated extract were applied to a silica gel Column Chromatography (CC) and eluted with increasing amounts of EtOAc in petroleum ether (pet. ether). Sixty fractions of 250 ml each were collected and pooled according to their TLC profile giving four major fractions (A-D). Fraction B obtained from the main column with pet. ether-EtOAc (80:20–75:35) was further chromatographed over a silica gel CC and eluted with pet. ether-

EtOAc of increasing polarity to give 1-hydroxy-3,4-dimethoxy-*N*-methylacridone **2** (orange crystals, 30 mg) and tegerrardin A **1** (yellow pellets, 100 mg). Fraction C obtained from the main column with pet. ether-EtOAc (70:30) was applied to a silica gel CC, eluted with pet. ether-EtOAc (80:20) to furnish arborinine **3** (yellow pellets, 400 mg) and scoparone **5** (colorless needles, 2 g). On the other hand, fraction D, obtained from the main column with pet. ether-EtOAc (50:50), was passed through a silica gel CC and eluted with pet. ether-EtOAc (50:50) to give xanthoxoline **4** (yellow pellets, 200 mg). The structures of compounds **1-5** were elucidated by analysing their ¹H and ¹³C NMR and mass spectra data and by comparison with those already reported (8,9,10,11).

Antioxidant activity

Iron chelating scavenging assay

The Fe⁺²-chelating ability of samples was determined following the modified method of Decker and Welch (12). The Fe⁺² ions were monitored by measuring the formation of ferrous ion-ferrozine complex. The pure compound (0.5 mM) was mixed with FeCl₂ (0.0625 mM) and ferrozine (0.5 mM). The resulting mixture was then shaken and left at room temperature for 10 min. The absorbance of the resulting mixture was measured at 562 nm. A lower absorbance of reaction mixture indicated a higher Fe⁺²-chelating ability. Radical scavenging activity of pure compounds was calculated by using the following formula:

$$\% \text{ RSA} = 100 - \{(\text{OD test compound} / \text{OD control}) \times 100\}.$$

1,1-Diphenyl-2-picrylhydrazyl (DPPH) radical scavenging assay

The assay was adapted from Shaheen and others (13) methodology. Test samples were allowed to react with DPPH, a stable free radical for half an hour at 37 °C. The concentration of DPPH was kept as 300 μM. Ten microlitres of test samples (0.5 mM) were dissolved in 90 μL DMSO, while DPPH solution was prepared in ethanol. After incubation, decrease in absorption was measured at 515 nm using multiplate reader (Spectra MAX-340, Molecular devices, CA, USA). Percent Radical Scavenging Activity (% RSA) of samples was determined in comparison with a DMSO treated control group. Radical scavenging activity was calculated using the following formula:

$$\% \text{ RSA} = 100 - \{(\text{OD test compound} / \text{OD control}) \times 100\}.$$

Superoxide radical scavenging assay

The assay protocol was adapted from Pieker and Fridovich (14) methodology. Ten microlitres of sample (0.5 mM) was mixed with 40 μ L of β -nicotinamide adenine dinucleotide (NADH-280 μ M), 40 μ L of nitro blue tetrazolium (NBT-80 μ M), 20 μ L of phenazine methoxysulphate (PMS-8 μ M), 10 μ L of sample (1 mM) and 90 μ L of phosphate buffer (0.1 M, pH 7.4). The reagents were prepared in buffer, while the sample was prepared in DMSO. The reaction was performed in 96-well microtitre plate at room temperature and absorbance was measured at 560 nm. The formation of superoxide was monitored by measuring the formation of water soluble blue Formazan dye. A lower absorbance of reaction mixture indicated a higher scavenging activity of the sample. Percent Radical Scavenging Activity (% RSA) of the sample was determined in comparison with propyl gallate as control using the following formula: % RSA = $100 - \{(\text{OD test compound} / \text{OD control}) \times 100\}$.

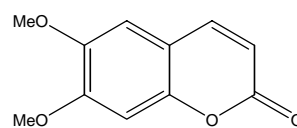
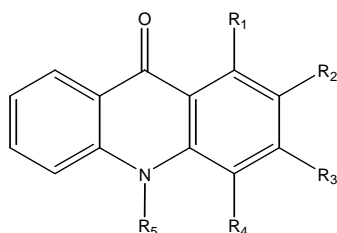
Antiplasmodial activity

In-vitro antiplasmodial activity was performed on the *Plasmodium falciparum* 3D7 strains using the parasite lactate dehydrogenase assay (pfLDH) as described by Makler and others (15). The 96-well plate format was employed for the assay. Plant constituents and standard drugs (chloroquine diphosphate) were serially diluted (two fold) with a complete culture medium in 96-well plates. A solution of infected RBC with 2% parasitemia and 1% hematocrit was added to make a total volume of 200 μ l in each well. Plates were incubated in a candle jar with 5% CO₂, 5% O₂ and 90% N₂ at 37 °C for 72 hours. To carry out the Malstat reaction (an immunocapture diagnostic for malaria), 20 μ l in 100 μ l of Malstat solution from each well of plate was added to respective well of Malstat plate. Plates were placed in an agitating water bath at 37 °C for 30 minutes. Twenty five microlitres (25 μ l) solution (1:1 solution of NBT (2 mg/ml) and PES (0.1 mg/ml) was added in each well and the plates were kept in the dark for a complete reaction and color development (a positive reaction was seen as a dark blue or reaction product. Plates were read at 650 nm and the OD was recorded. IC₅₀ value was calculated with the help of an EZfit computer program.

Results and Discussion

Four acridone alkaloids, namely tegerrardin A **1** (10), 1-hydroxy-3,4-dimethoxy-*N*-methylacridone **2** (8), arborinine **3** and xanthoxoline **4** (6,9) and a coumarin, scoparone **5** (11) have been isolated from the fruits of *Z. leprieurii*. Compound **1** has been reported only from *Teclea gerrardi* (10), whereas only one source (8) reports the isolation of compound **2**. Compounds **3**, **4** and **5** have been previously reported from the fruits of *Z. leprieurii* and various Rutaceae species, whereas compounds **1** and **2** are reported herein for the first time from that plant.

All the isolated acridone alkaloids **1-4**, scoparone **5** and crude extract of the fruits of *Z. leprieurii* were tested against the 3D7 strains of *Plasmodium falciparum*. After 72 hrs. of incubation, compounds **3** and **4** were found to exhibit a good *in vitro* antiplasmodial activity with IC₅₀ values of 4.5 ± 1.0 and 4.6 ± 0.6 µg/ml, respectively whereas compounds **1**, **2** and **5** and the crude EtOAc extract were found to be inactive against the parasites (Table 1). The lack of activity in constituents and the crude extract may be due to the antagonistic effect of the different constituents present in the crude extract. The antiplasmodial effects of compounds **3** and **4** were comparable to those of some natural acridone alkaloids reported earlier (10,16,17,18,19).



5. Scoparone

- 1.** R₁ = OCH₃, R₂ = R₄ = H, R₃ = OH, R₅ = CH₃
Tegerrardin A
- 2.** R₁ = OH, R₂ = H, R₃ = R₄ = OCH₃, R₅ = CH₃
1-Hydroxy-3,4-dimethoxy-*N*-methylacridone
- 3.** R₁ = OH, R₂ = R₃ = OCH₃, R₄ = H, R₅ = CH₃
Arborinine
- 4.** R₁ = OH, R₂ = R₃ = OCH₃, R₄ = H, R₅ = H
Xanthoxoline

Table 1: Antiplasmodial activity of compounds **1-5** and crude of extract of *Z. leprieurii* against *Plasmodium falciparum* (3D7 strain), after 72 hrs. of incubation at 37 °C

Compound	IC ₅₀ (µg/ml)
Tegerrardin A (1)	17.3 ± 3.0
1-Hydroxy-3,4-dimethoxy-N-methylacridone (2)	-
Arborinine (3)	4.5 ± 1.0
Xanthoxoline (4)	4.6 ± 0.6
Scoparone (5)	> 25
Crude	> 25
Chloroquine diphosphate (standard)	0.0250 ± 0.01

Compounds **1-5** were screened for their antioxidant activities (Table 2) employing the superoxide, iron chelating, and DDPH scavenging assays. Compound **2** showed the highest activity (RSA 75%) in the superoxide anion scavenging assay, comparable to that of n-propyl gallate used as standard (90%). Compounds **1**, and **3 - 5** exhibited low activity. On metal-chelating assay, arborinine **3** showed the highest iron-chelating ability, followed by tegerrardin A **1** (90 and 61% RSA respectively) in comparison with the standard EDTA (98% RSA). Results suggest therefore that arborinine **3** and tegerrardin A **1** can be considered as strong and moderate chelating agents, respectively. In addition, compound **3** exhibited both iron binding and antiplasmodial activity which may be strongly interrelated. Iron is known to play a critical role in host-parasite interaction (20,21), and is essential for life, because it is required for oxygen transport, respiration, and for activity of many enzymes. Hence it is important for the growth of bacteria, fungi, protozoan parasites and mammalian cells. Iron deprivation can therefore be detrimental to *Plasmodium falciparum* and may be a useful approach for the treatment of malaria. Ironically, iron supplementation in anaemic children and women areas where malaria is endemic may encourage malaria infection (21). The mechanism of action of iron chelators has however not been clearly elucidated. Several hypotheses have been suggested: iron-binding, inhibition of iron-dependent enzymes (ribonucleases, phosphoenolpyruvate carboxykinase, superoxide dismutase, etc) and the formation of free

radicals. According to El Nehir and Karakya (22), iron is capable of generating free radicals from peroxides by Fenton reactions, and the minimisation of the Fe^{2+} ions in the Fenton reaction affords protection against oxidative damage. The present result indicates a positive correlation between the *in vitro* antiplasmodial activities of the iron chelating agents and their iron binding potentials (23). More studies need to be carried out on arborinine **3** for its possible use as an efficient and non-toxic iron chelating agent. The present findings suggest the potential of arborinine **3**, 1-hydroxy-3,4-dimethoxy-*N*-methylacridone **2** and xanthoxoline **4** in malarial treatment. The synergy of these compounds should be considered because when compounds **3** and **4** exhibit antiplasmodial effect, compound **2** and **3** due to their antioxidant activities are expected to boost the patient immune system (24). To the best of our knowledge, this is the first report of the iron chelating activity of acridone alkaloids.

All tested compounds were found to be inactive in the DPPH radical scavenging assay.

Table 2: Antioxidant activities of compounds **1-5**.

Compound	Conc. (μM)	Superoxide anion scavenging assay % RSA	Metal chelating assay % RSA	DPPH radical scavenging assay % RSA
1	0.5	25	61	5.5
2	0.5	75	-	
3	0.5	33	90	2
4	0.5	18	3	10
5	0.5	24	24	0.5
n-Propyl gallate (standard)	0.5	91	-	- 90
EDTA (standard)	1 mM	-	98	
Ascorbic acid	0.5			92

The antioxidant activities of acridone alkaloids appear to have not been extensively studied. However, Wansi and others (25) reported the moderate free radical scavenging of some prenylated acridone alkaloids isolated from *Oriciopsis glaberrima* (Rutaceae).

In conclusion, the fruits of *Z. leprieurii* are found to possess antioxidant and antiplasmodial potentials. 1-Hydroxy-3,4-dimethoxy-*N*-methylacridone, arborinine, and xanthoxoline isolated in the course of this work may be a potential antimalarial therapy due to their possible synergistic effects. The consumption of these fruits may therefore be beneficial as source of antioxidants and prevention against malaria in endemic areas like Cameroon.

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References

1. Soobrattee MA, Neergheen VS, Luximon-Ramma A, Aruomab OI, Bahorun T. Phenolics as potential antioxidant therapeutic agents: Mechanism and actions. *Mut Res* 2005; 579:200–213.
2. Halliwell, B, Gutteridge JMC. The definition and measurement of antioxidant in biological systems. *Free Rad Biol Med* 1999; 18:125-126.
3. Stangeland T. Antioxidants in Ugandan fruits and vegetables. In: Book of extended abstracts of the 12th Symposium of the Natural Product Research Network for Eastern and Central Africa. Kampala: Makerere University Press. 2007:27.
4. Lamaty G, Menut C, Bessiere JM, Aknin M. Aromatic plants of tropical Central Africa. II. A comparative study of the volatile constituents of *Zanthoxylum leprieurii* (Guill. et Perr.) Engl. and *Zanthoxylum tessmannii* Engl. leaves and fruit pericarps from the Cameroon. *Flav Fragr J* 1989; 4:203-205.

5. Hutchison J, Dalziel JM. Flora of West Tropical Africa, 2nd edition, volume 1, part 2. London, UK: Crown Agents for Overseas governments and Administration. 1958.
6. Reisch J, Adesina SK, Bergenthal D. Constituents of *Zanthoxylum leprieurii* fruit pericarps. Part 103: Natural product chemistry. Pharmazie 1985; 40:811-812.
7. Tatsajeu LN, Ngang JEJ, Etoa FX. Antibacterial and antifungal activity of *Xylopiya aethiopica*, *Monodora myristica*, *Zanthoxylum xanthoxyloides* and *Zanthoxylum leprieurii* from Cameroon. Fitoterapia 2003; 64:469-472.
8. Baudouin G, Tillequin F, Koch M, Dau M-E.T H, Guilhem J, Pusset J, Chauviere, G. Plantes de Nouvelle-Caledonie, XCII, alcaloïdes de *Sarcomelicope leiocarpa*. J Nat Prod 1985; 48:260-265.
9. Spatafora C, Tringali C. Bioactive metabolites from the bark of *Fagara macrophylla*. Phytochem anal 1997; 8:139-142.
10. Waffo AFK, Coombes PH, Crouch NR, Mulholland DA, El Amin SMM, Smith PJ. Acridone and furoquinoline alkaloids from *Teclea gerrardii*. (Rutaceae: Toddalioideae) of southern Africa. Phytochemistry 2007; 68:663-667.
11. Ma CH, Ke W, Sun ZL, Peng JY, Li ZX. Large-scale isolation and purification of scoparone from *Herba artemisiae scopariae* by high-speed counter-current chromatography. Chromatographia 2006; 64:83-87.
12. Decker EA, Welch B. Role of ferritin as a lipid oxidation catalyst in muscle food. J Agric Food Chem 1990; 38:674-677.
13. Shaheen F, Ahmad M, Tareq M, Khan H, Jalil S, Ejaz A, Sultankhodjaev MN, Arfan M, Choudhary MI, Atta-ur-Rahman. Alkaloids of *Aconitum* leaves and their anti-inflammatory, antioxidant and tyrosinase inhibition activities. Phytochemistry 2005; 66:935-940.
14. Pieker SD, Fridovich I. Iron chelating scavenging assay on the mechanism of production of superoxide by reaction mixtures containing NADH. Archiv Biochem Biophys 1984; 228:155-158.
15. Makler MT, Ries JM, Williams JA, Bancroft JE, Piper RC, Gibbins BL, Hinrichs DJ. Parasite lactate dehydrogenase as an assay for *Plasmodium falciparum* drug sensitivity. Amer J Trop Med Hyg 1993; 48:739-741.
16. Weniger B, Um B-H, Valentin A, Estrada A, Lobstein A, Anton R, Le Maille M, Sauvains M. Bioactive acridones from *Swinglea glutinosa*. J Nat Prod 2001 ; 64:1221-1223.

17. Queener SF, Fujioka H, Nishiyama Y, Furukawa H, Bartlett M, Smith JW. *In vitro* activities of acridone alkaloids against *Pneumocystis carinii*. Antimicrob Ag Chemother 1991; 35:377-379.
18. Fujioka H, Kato N, Fujita M, Fujimura K, Nishiyama Y. Activities of new acridone alkaloid derivatives against *Plasmodium yoelii in vitro*. Drug Res 1990; 40:1026-1029.
19. Michael JP. Quinoline, quinazoline and acridone alkaloids. Nat Prod Rep 2007; 24:223-246.
20. Egan TJ, Combrinck JM, Egan J, Hearne GR, Marques HM, Ntenti S, Sewells BT, Smith PJ, Taylor D, Van Schalkwyk DA, Walden J C. Fate of haem iron in the malaria parasite *Plasmodium falciparum*. Biochem J. 2002; 365:343-347.
21. Pradines B, Ramiandrasaa F, Basco LK, Bricard L, Kunesch G, Bras JL. *In vitro* activities of novel catecholate siderophores against *Plasmodium falciparum*. Antimicrob Ag Chemother 1996; 40:2094-2098
22. El Nehir S, Karakaya S. Radical scavenging and iron-chelating activities of some greens used as traditional dishes in Mediterranean diet. Inter J Food Sci Nutr 2004; 55: 67-74.
23. Pradines B, Rolian JM, Ramiandrasoa F, Fusai T, Mosnier J, Rogier C, Daries W, Baret E, Kunesch G, Le Bras J, Parzy D. Iron chelators as antimalaria agents: *in vitro* activity of dicatecholates against *Plasmodium falciparum*. J Antimicrob Chemother 2002; 50:177-187.
24. Wade C. The role of antioxidant in boosting the immune system. In: Eat away illness. New York: Parker Publishing Company. 1992:293-298.
25. Wansi JD, Wandji J, Meva'a LM, Waffo AFK, Ranjit R, Khan SN, Asma A, Iqbal CM, Lallemand MC, Tillequin F, Tane ZF. α -Glucosidase inhibitory and antioxidant acridone alkaloids from the stem bark of *Oriciopsis glaberrima* Engl. (Rutaceae). Chem Pharm Bull 2006; 54:292-296.