

**THE ANTIOXIDANT AND H⁺K⁺ ATPASE INHIBITORY EFFECT OF
ANDROGRAPHIS PANICULATA AND ANDROGRAPHOLIDE –IN VITRO AND
IN VIVO STUDIES**

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Summary

Andrographis paniculata popularly known as kalmegh of ayurveda and the andrographolide present in the leaves have been widely included in various drug formulations used for liver and gastric ailments. The aim of the present study was to evaluate the *in vitro* antioxidant and the antiulcer activities of the extracts of aerial parts of *Andrographis paniculata* prepared by using solvents such as petroleum ether, ethyl acetate, ethanol, hydroalcohol and of purified andrographolide. The antiulcer effect was assessed by the capacity to inhibit the H⁺K⁺ ATPase activity of sheep gastric mucosal cells *in vitro*. The antiulcer activity was also evaluated *in vivo* with different experimental ulcer models in rats. The antioxidant activity was determined by using DPPH, superoxide, hydroxyl and nitric oxide generating models and by reducing power. The HPLC-UV chromatogram analysis revealed the presence of important constituents such as Quercetin and Formononetin. Among all the fractions studied, hydroalcoholic fraction exhibited significant free radical scavenging activity in all the models. The activities were found to be dose dependent and this study indicates that both hydroalcoholic extract and andrographolide exhibit free radical scavenging activity *in vitro*. The test compounds were found to suppress sheep mucosal H⁺K⁺ ATPase activity *in vitro* and the ulcer score *in vivo*. So further study is needed to confirm the gastroprotective property of *Andrographis paniculata*.

Key words: *Andrographis paniculata*; andrographolide; free radicals; scavenging activity; H⁺K⁺ATPase; antiulcer property.

Introduction

Andrographis paniculata also known as king of bitters is the member of plant family Acanthaceae and has been used for centuries in Asia to treat gastrointestinal discomfort, liver disorders and a variety of other chronic and infectious diseases. It is included in twenty six ayurvedic formulations shown in ancient Indian pharmacopoeia. The abortifacient, acrid, analgesic, anti-inflammatory^{1, 2}, antibacterial, antiviral³, antipyretic^{4, 5}, hypoglycemic, cardioprotective, hepatoprotective and immune enhancement properties of *Andrographis paniculata* have shown that the plant has a broad range of pharmacological effects.

Andrographolide, a chief constituent of the leaves is a bitter water soluble lactone exhibiting protective effect on CCl₄ induced hepatopathy in rats. Earlier work showed that andrographolide has multiple pharmacological activities such as inhibition of platelet activation², stimulation of cell differentiation⁶ and stimulation of immunity. Chiou *et al*^{7,8} reported that andrographolide inhibited nitric oxide production in macrophage cell line.

Peptic ulceration is a common, chronic, recurrent, and occasionally life-threatening disease of poorly defined pathogenesis in humans^{9,10}. The gastric mucosa is continuously exposed to potent injurious agents such as acid, pepsin, bile acids, food ingredients, bacterial products and drugs¹¹. Increase in gastric acid and pepsin secretion, decrease in gastric blood flow, suppression of endogenous generation of prostaglandins, inhibition of mucosal growth and cell proliferation, and alteration of gastric motility have been implicated in the pathogenesis of gastric ulcer¹². Hyperactivity of H⁺K⁺ ATPase has been claimed for the excess acidity and mucosal inflammation in gastric ulcer. Hence agents which suppress the activity of H⁺K⁺ ATPase activity are considered as a choice of antiulcer drugs.

Reactive oxygen species are generated by cells in some physiological and pathological circumstances. Any derangement between in ratio of pro-oxidants to antioxidants, in which pro-oxidants prevail leads to oxidative stress¹³. Insufficient antioxidant protection or excess production of reactive oxygen species can result this condition. Reactive oxygen species can react with all macromolecules such as lipids, proteins, nucleic acids and carbohydrates, particularly polyunsaturated fatty acids of cell membranes. Initial reaction with reactive oxygen species causes a continuous chain reaction resulting in cell injury and ultimately cell death¹⁴. Peptic ulcer is produced by the imbalance between gastroduodenal mucosal defense mechanism and offensive factors. Some studies revealed that ROS and lipid peroxidation are implicated in the pathogenesis of ethanol induced gastric lesions and gastrointestinal damage¹⁵⁻¹⁷.

Reactive oxygen species produced *in vivo* including superoxide radical (O₂⁻), hydrogen peroxide (H₂O₂) and hypochlorous acid (HOCl). H₂O₂ and O₂⁻ can interact in the presence of certain transition metal ions to yield a highly reactive oxidizing species, the hydroxyl radical (·OH).¹⁸ Free radicals are chemical species of atoms or molecules that possess an unpaired electron on their outermost orbit. These free radicals are highly unstable and can react with other molecules by giving out or accepting single electron. Although the human body continuously produces free radicals it possesses several defense systems constituting enzymes and radical scavengers¹⁹. Therefore antioxidants with free radical scavenging activities may have great relevance in the prevention and treatment of diseases in which oxidants or free radicals are involved²⁰.

Active components of plants such as flavonoids, triterpenes and tannins may be regarded as possible phytochemicals against gastric lesions by acting as protective factors or antioxidants. It is believed that the antioxidant activity of polyphenols is an important factor because reactive oxygen and free radicals are related to the occurrence of ulcers.

Herbal medicines are now widely used by nearly 50% of the world population, in a number of instances for the treatment or prevention of digestive disorders^{21,22}. In traditional medicine, numerous plants and herbs are used to treat gastrointestinal disorders. There has been renewed interest in identifying new anti ulcer drugs from natural sources²³.

Many plants such as *Cedrus libani*, *Centaurea Solstitialis ssp. Solstitialis*, *Cistus laurifolius*, *Hypericum scabrum*, *Plantago major*, *Sambucus ebulus* and *Spartium junceum*²⁴ have been proved to exhibit anti-ulcer properties in experimental animals. These plants and their products are proved gastroprotective and found to act by different mechanisms. The ever-increasing problem of ulcer due to advancement in food pattern and life style, demands the identification and evaluation of new plants with anti-ulcer and antioxidant properties. In the present investigation, various fractions of *Andrographis paniculata* and andrographolide were tested for free radical scavenging and H⁺K⁺ ATPase inhibiting activity. The effect was further confirmed by testing the drug in ulcer induced rats.

Materials and Methods

Plant collection and identification

The plant was purchased from the local market in Chennai and authenticated by Dr. Jayaraman, Taxonomist, Plant Anatomy Research Centre, Chennai (Voucher No. of the Specimen: PARC/ 2008/ 185). The aerial parts were washed, air dried and pulverized to powder form.

Preparation of plant extracts

The test material was subjected to serial extraction with solvents of different polarity in the order of petroleum ether, ethyl acetate, ethanol and hydroalcohol. Briefly, 15-20g of the dried and homogenous powder of *Andrographis paniculata* was soaked in 100ml of petroleum ether 60-80°C by cold percolation method and the extract was filtered. The residue was air dried and used for subsequent extraction with solvents like ethyl acetate, ethanol, and hydroalcoholic extract was prepared separately in a similar manner. The filtrate collected in each extraction was evaporated in a desiccator and the filtrate was concentrated. The concentrate was lyophilized and used for the study.

Isolation of Andrographolide

The dried leaves of *Andrographis paniculata* were macerated in methanol (4.01x2 times) and kept at room temperature for 3 days. After filtration, the methanol solution was evaporated under reduced pressure. The residue was partitioned between ethyl acetate and water. The water soluble portion was extracted with n-butanol and filtered. The crude andrographolide, a part of which was chromatographed on a silica gel column using chloroform/methanol (20/1) as a solvent to yield pure andrographolide²⁵.

Antioxidant activity

DPPH radical scavenging activity

The DPPH radical scavenging capacity of the extracts determined by the method described by Vitorro et al, 1999²⁶. 0.1ml of of extracts (5-25µg/ml) was mixed with 0.1ml of 1, 1'-diphenyl 1-2-dipicryl hydrazine (DPPH) in dimethyl sulphoxide (DMSO) (0.15%) and incubated for 10 min. The absorbance was then read at 450nm in ELISA strip reader (Qualigens). Percentage inhibition was determined and compared with that of similar concentration of ascorbic acid. IC₅₀ values were calculated for both test extracts and the standard.

Nitric oxide scavenging activity

The method of Sreejayan Rao 1997²⁷ was used to measure the nitric oxide scavenging of the extracts. Various concentrations of the test material (30-150 µg/ml) was dissolved in phosphate buffer (0.025M, pH 7.4) and incubated with sodium nitroprusside (5 µM) in standard phosphate buffer at 25°C for 5 hrs. After incubation, 0.5ml of the reaction mixture was added with 0.5ml of Griess reagent (equal volumes of 1% sulphanilamide in 2% phosphoric acid and 0.1% naphthyl ethylene diamine dihydrochloride in water). The absorbance of the Chromophore formed was read at 540nm. The activity was compared with that of rutin.

Superoxide anion scavenging activity

The method of Nishmiki *et al* 1972²⁸ was used to measure the superoxide anion scavenging activity of the extracts. Briefly 1ml of nitro blue tetrazolium (NBT) (156 µM NBT in 100mM phosphate buffer, pH 7.4, 1ml of nicotinamide adenine dinucleotide (NADH) in 100mM phosphate buffer, pH 7.4) were added to the 0.1ml of the test extract with varying concentrations (5-25µg/ml). The reaction was started by adding 100µl of phenazonium methosulphate (PMS) (60 µM in 100mM phosphate buffer, pH 7.4) and the reaction mixture was incubated at 25°C for 5 min. The decrease in absorbance was measured at 560nm against water blank. Curcumin was used as the positive control.

Hydroxyl radical scavenging activity

The assay was based on the method of Halliwell *et al.*, 1987²⁹. 1ml of the reaction mixture contained 100µl of 28mM 2-deoxy-2-ribose in phosphate buffer, pH 7.4, 500 µl of various concentrations of plant extract (10-80µg/ml), 200µl of 200µM ferric chloride and 1.04mM ethylenediamine tetra acetic acid (EDTA). The reaction mixture was incubated and deoxyribose degradation was measured by the thiobarbituric acid (TBA) reaction³⁰. The absorbance was measured at 532 nm. Vitamin E was used as the standard.

Reducing Power

Varying concentrations of different extracts (100-1000µg) were mixed with 2.5ml of phosphate buffer (0.2M, pH 6.6), 2.5ml of 1% potassium ferricyanide and incubated at 50°C for 20 min. 1.5 ml of 10% trichloroacetic acid (TCA) was then added to the reaction mixture and the contents were centrifuged at 3000rpm for 10 min. 0.5 ml of the supernatant was collected and mixed with 1 ml of distilled water and 0.5 ml 0.1% ferric chloride. Control was processed similarly with distilled water. Ascorbic acid was used as standard. Increase in absorbance at 700nm indicates increased reducing power³¹.

Preparation of Parietal cells

Proton potassium ATPase was prepared from mucosal scrapings of sheep stomach obtained from slaughter house and then homogenized in 200mM Tris-HCl buffer, pH 7.4, centrifuged for 10 mins at 5000xg. The resulting supernatant was subsequently centrifuged at 5000xg for 20 min³². The protein concentration in the supernatant was determined with bovine serum albumin as standard³³. The parietal cell extract was then employed to determine H⁺K⁺ ATPase activity.

Determination of H⁺K⁺ ATPase

The H⁺K⁺ ATPase activity in the presence of different concentrations of test extracts and omeprazole was assayed by the method of Reyes- Chilpa *et al* 2006³⁴. The enzyme source was preincubated with different concentration of the test material (10-80µg) for 30min. The assay was conducted in a mixture contained an aliquot of *Andrographis paniculata* treated enzyme in 20mM tris-HCl, pH 7.4, 2mM magnesium chloride (MgCl₂) and 2mM potassium chloride (KCl). The reaction was started with the addition of 2mM adenosine-5'-triphosphate (ATP) and incubated for 30 mins at 30°C and terminated by the addition of 10% trichloroacetic acid followed by centrifugation at 2000xg. The amount of inorganic phosphorous released from adenosine-5'-triphosphate (ATP) was determined spectrophotometrically at 640nm. The enzyme source was also treated similarly with the standard drug omeprazole and the enzyme activity was measured.

HPLC analysis

A flavonoids profile assay was performed using HPLC-UV chromatogram. It was used at 40°C with linear gradient mobile phase (methanol, water and phosphoric acid) in the ratio of 100:100:1 and solvent mixture (alcohol, water and hydrochloric acid) in the ratio 50:20:8 with flow rate set of 1.5/min. Equal volumes of standard (20µl) and test solutions were injected into chromatograph. The major peaks were determined according to the retention times obtained from authentic standards run at identical conditions.

In vivo antiulcer study

Animals

Male albino Wistar rats (120-140g) were obtained from Kings Institute, Chennai, India. They were acclimatized to animal house conditions, fed commercial pelleted rat chow (Hindustan Lever Ltd., Bangalore, India) and water *ad libitum*. Animals were maintained according to the rules and regulations laid down by the Institutional Ethics Committee.

Treatment protocol for antiulcer activity

Rats were divided into VII groups of six animals each.

Group I (a-c) - Rats treated with ulcerogenic agents.

Group II (a-c) -Rats pretreated with 200mg/kg b wt of petroleum ether extract of *Andrographis paniculata* for 30days and then subjected to ulcer induction.

Group III (a-c)-Rats pretreated with 200mg/kg b wt of ethyl acetate extract of *Andrographis paniculata* for 30days and then subjected to ulcer induction.

Group IV (a-c)- Rats pretreated with 200mg/kg b wt of ethanol extract of *Andrographis paniculata* for 30days and then subjected to ulcer induction.

Group V (a-c) -Rats pretreated with 200mg/kg b wt of hydroalcoholic extract of *Andrographis paniculata* for 30 days and then subjected to ulcer induction.

Group VI (a-c)- Rats pretreated with 3mg/kg b wt of andrographolide for 30 days and then subjected to ulcer induction.

Group VII (a-c)- Rats pretreated with 30mg/kg b wt of omeprazole for 30 days and then subjected to ulcer induction.

Ulcer induction

By ethanol (a)

Gastric ulcer was produced by administering ethanol orally 1ml/200g to rats after 12hr fasting. The animals were sacrificed after 1hr of ethanol administration³⁵.

By aspirin (b)

Aspirin was administered at the dose of 200mg/kg b.wt and ulcer score was determined after 4h. The stomach was cut open along the greater curvature and ulcer index was scored by a person unaware of the experiment protocol in the glandular portion of the stomach³⁶.

By pylorus ligation (c)

Drugs were administered for a period of 30 days. On day 30, after the last dose, the rats were kept for 18h fasting. Rats were anaesthetized using diethyl ether, the abdomen was opened and pylorus ligation was done without causing any damage to its blood supply. After replacing the stomach carefully the abdomen wall was closed in two layers with interrupted sutures. The animals were deprived of water during the postoperative period³⁷. After 4h, stomach was dissected out and the contents collected.

Determination of ulcer score

The ulcer index of gastric mucosal lesions was evaluated by the score system reported by Nie et al., 2003³⁸. Briefly, after collecting gastric juice, the stomach was opened along the greater curvature and rinsed with 0.1mol/L ice-cold PBS. The stomach was then examined under microscope (100x) to observe erosions and made scores as 1-5: 1. small round hemorrhagic erosion, 2. haemorrhagic erosion<1mm, 3. Hemorrhagic erosion = 1-2mm, hemorrhagic erosion = 2-3mm, 5. haemorrhagic erosion>4mm. The score was multiplied by 2 when the width of the erosion is larger than 1mm.

Statistical analysis

Data were analyzed by using a commercially available statistics software package (SPSS for window V.7.5). Student's t test was performed and results were presented as mean \pm S.E.M.

Results & Discussion

Reactive oxygen species and other free radicals have been implicated in the pathology of many disorders like neurodegenerative diseases, inflammation, cataract, liver cirrhosis and gastrointestinal disorders including ulcer³⁹. Herbal drugs containing radical scavengers are widely applied for the prevention and treatment of various diseases. Larson⁴⁰ reported that phenolic compounds, flavanoids and tannins are the major constituents in most of the medicinal plants to possess antioxidant activity and responsible for the free radical scavenging effect.

The scavenging activity of different extracts of *Andrographis paniculata* on DPPH radicals is shown in table1. The IC₅₀ value of petroleum ether, ethyl acetate, ethanol, hydroalcoholic extracts and the isolated andrographolide were found to be 8.7 μ g/ml, 7.4 μ g/ml, 6.5 μ g/ml, 5.3 μ g/ml and 5.2 μ g/ml respectively.

Table no. 1 DPPH scavenging activity of various fractions of *Andrographis paniculata* and andrographolide.

| Various Fractions | %inhibition | | | | |
|-------------------|--------------------------|-----------------------|-----------------------|------------------------|------------------------|
| | 5 | 10 | 15 | 20 | 25 |
| | (Concentration in µg/ml) | | | | |
| Petroleum ether | 10 ± 1.0 | 15 ± 1.6 | 20 ± 2.2 | 25 ± 2.8 | 28 ± 3.2 |
| Ethyl acetate | 15 ± 1.5 | 20 ± 2.1 | 23 ± 2.5 | 29 ± 3.3 | 36 ± 4.0 |
| Ethanol | 25 ± 2.5 | 33 ± 3.4 | 40 ± 4.2 | 45 ± 4.8 | 58 ± 6.2 |
| Hydroalcoholic | 29 ± 2.9* | 36 ± 3.7 [#] | 48 ± 5.0 [#] | 52 ± 5.5 ^{\$} | 59 ± 6.3 ^{\$} |
| Andrographolide | 31 ± 3.1* | 40 ± 4.1 [#] | 50 ± 5.2 [#] | 59 ± 6.2 ^{\$} | 65 ± 6.9 ^{\$} |
| Ascorbic acid | 33 ± 3.3* | 45 ± 4.6 [#] | 69 ± 7.1 [#] | 75 ± 7.8 ^{\$} | 77 ± 8.1 ^{\$} |

Values are expressed as mean ± SD for six individual experiments. Statistically significant difference is expressed as ^{\$}p<0.001, [#]p<0.01, *p<0.05.

DPPH is a stable free radical, rapid and sensitive way to survey the antioxidant activity of a specific compound or plant extracts ⁴¹. When DPPH encounters a proton – donating substance such as an antioxidant, the radical would be scavenged and the absorbance reduced. Based on this principle, the antioxidant activity of a substance can be expressed as an ability to scavenge the DPPH radical. Among the various extracts tested, andrographolide and hydroalcoholic extract were found to exhibit greater DPPH radical scavenging activity. The IC₅₀ values of andrographolide and hydroalcoholic extracts were found to be nearer to that of standard ascorbic acid 5.0µg/ml.

The superoxide scavenging activity of the extracts with their IC₅₀ values are shown in table 2. IC₅₀ values of petroleum ether, ethyl acetate, ethanol, hydroalcoholic and andrographolide were found to be 11µg/ml, 9.3µg/ml, 8.5µg/ml, 7µg/ml and 6µg/ml respectively.

Table no. 2 Superoxide scavenging activity of various fractions of *Andrographis paniculata* and andrographolide.

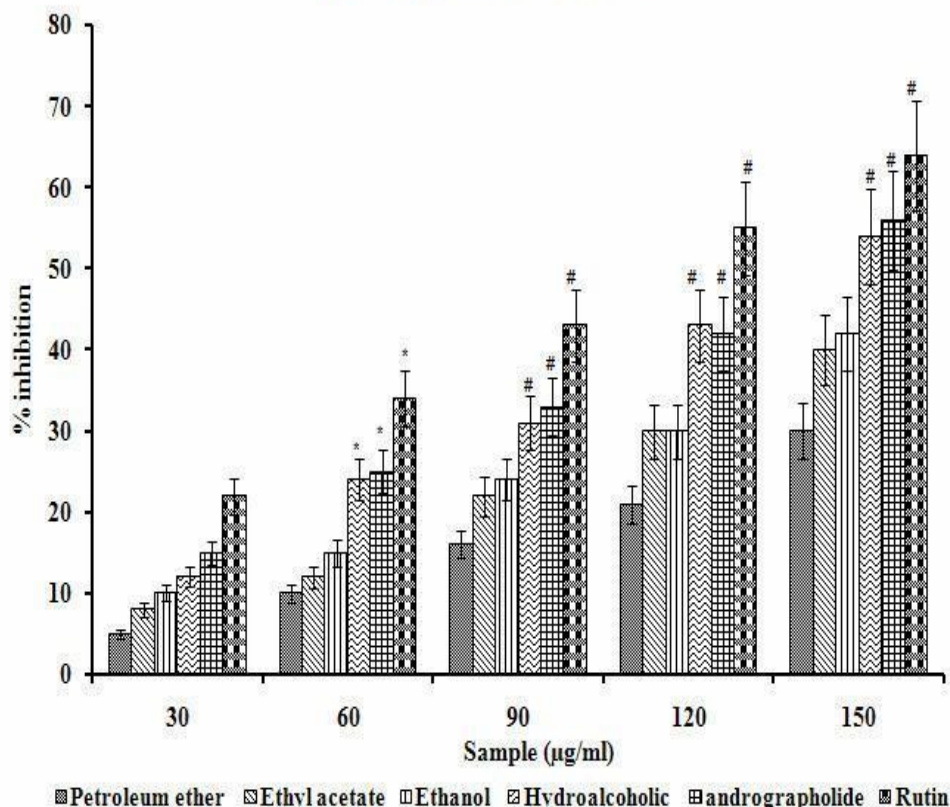
| Various Fractions | %inhibition | | | | |
|--------------------------------------|---------------|---------------|---------------------------|----------------------------|----------------------------|
| | 5 | 10 | 15 | 20 | 25 |
| (Concentration in $\mu\text{g/ml}$) | | | | | |
| Petroleum ether | 12 \pm 1.2 | 19 \pm 2.0 | 22 \pm 2.4 | 25 \pm 2.8 | 40 \pm 4.4 |
| Ethyl acetate | 15 \pm 1.5 | 21 \pm 2.2 | 26 \pm 2.8 | 32 \pm 3.5 | 42 \pm 4.6 |
| Ethanol | 19 \pm 1.9 | 28 \pm 2.9 | 36 \pm 3.8 | 45 \pm 4.8 | 52 \pm 5.6 |
| Hydroalcoholic | 21 \pm 2.1* | 36 \pm 3.7* | 41 \pm 4.3 [#] | 52 \pm 5.5 ^{\$} | 56 \pm 6.0 ^{\$} |
| Andrographolide | 29 \pm 2.9* | 38 \pm 3.9* | 43 \pm 4.5 [#] | 60 \pm 6.3 ^{\$} | 64 \pm 6.8 ^{\$} |
| Curcumin | 32 \pm 3.2* | 40 \pm 4.1* | 45 \pm 4.7 [#] | 68 \pm 7.1 ^{\$} | 70 \pm 7.4 ^{\$} |

Values are expressed as mean \pm SD for six individual experiments. Statistically significant difference is expressed as ^{\$}p<0.001, [#]p<0.01, *p<0.05.

Superoxide anion is a free radical created during energy metabolism in the human body. Superoxide anion is toxic to cells and tissues and can act as precursors to other reactive oxygen species⁴². It was found that all the extracts exhibited superoxide scavenging activity but more significantly hydroalcoholic extract and andrographolide. The IC₅₀ value of andrographolide (6 $\mu\text{g/ml}$) was found to be comparable to that of standard curcumin (5.7 $\mu\text{g/ml}$).

Nitric oxide is a free radical that exhibits numerous physiological properties and it is also implicated in several pathological states⁴³. The nitric oxide scavenging activity of various solvent extracts is shown in fig.1. The NO scavenging activity is seen in the order of petroleum ether, ethyl acetate, ethanol, hydroalcoholic and of andrographolide with the IC₅₀ values of 85 $\mu\text{g/ml}$, 80 $\mu\text{g/ml}$, 74 $\mu\text{g/ml}$, 70 $\mu\text{g/ml}$ and 66 $\mu\text{g/ml}$ respectively.

Fig 1. Nitric oxide scavenging activity of various extracts of *Andrographis paniculata* and andrographolide

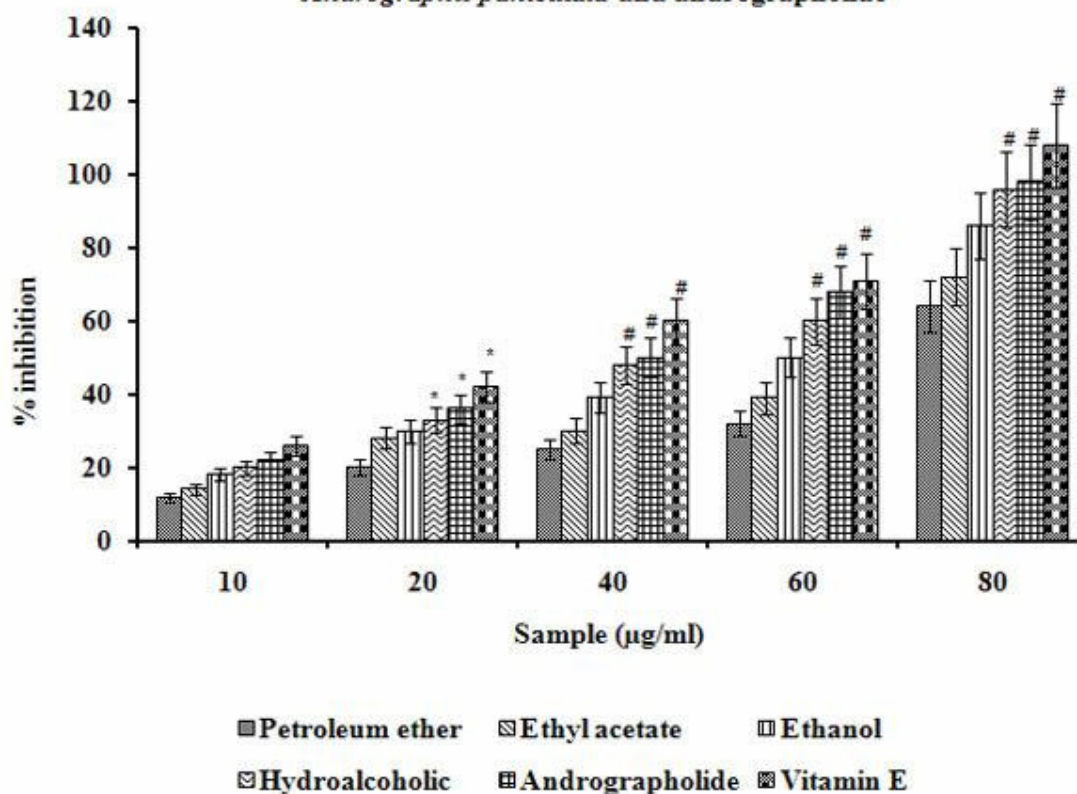


Values are expressed as mean \pm SD for six individual experiments. Statistically significant difference is expressed as # $p < 0.01$, * $p < 0.05$ when compared to minimal dose.

Excess production of nitric oxide is associated with several diseases. NO is produced in various cells including neurons, endothelial cells and neutrophils by three isoforms of NO synthase, from L-arginine⁴⁴. The nitric oxide scavenging activity of the various extracts shows that the isolated compound andrographolide exhibited maximum nitric oxide scavenging activity. When comparing the IC_{50} value of rutin 60 μ g/ml, andrographolide showed the highest nitric oxide scavenging activity when compared to the other solvent extracts.

The hydroxyl radical is extremely reactive in biological systems and has been implicated as highly damaging species in free radical pathology capable of damaging biomolecules of the living cells^{45, 46}. Fig. 2 show the hydroxyl radical scavenging activity of various solvent extracts. The isolated andrographolide showed maximum scavenging activity with the IC_{50} value of 38 μ g/ml. The IC_{50} values of petroleum ether, ethyl acetate, ethanol and hydroalcoholic extracts were found to be 60 μ g/ml, 54 μ g/ml, 47 μ g/ml and 40 μ g/ml respectively. The IC_{50} value of vitamin E was found to be 34 μ g/ml.

Fig 2. Hydroxyl radical scavenging activity of various fractions of *Andrographis paniculata* and andrographolide



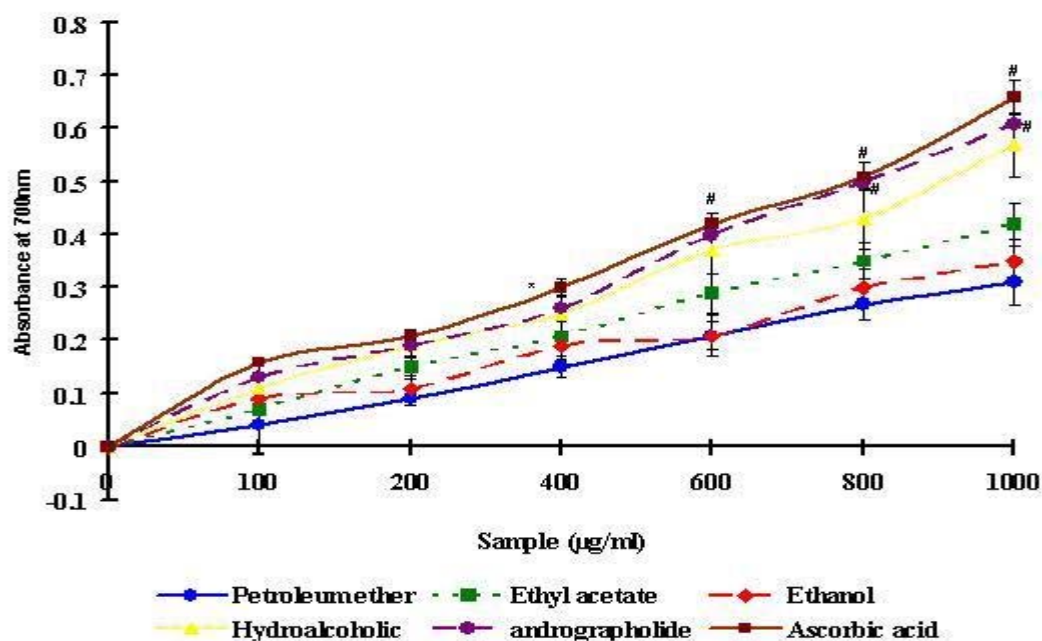
Values are expressed as mean \pm SD for six individual experiments. Statistically significant difference is expressed as # $p < 0.01$, * $p < 0.05$ when compared to minimal dose.

Some compounds are capable of redox cycling the metal ions required for hydroxyl ions generation, thus increasing the radical production, exhibiting a pro-oxidant activity⁴⁷. All the extracts have shown inhibitory activity against hydroxyl radical and there was a significant difference in their activities at similar concentrations. Among all the test compounds, andrographolide exhibited greatest scavenging activity. The IC_{50} value of andrographolide was comparable with that of Vitamin E.

Antioxidant activity of medicinal plant products has been reported to be attributed to their reducing power. The reducing power of the various extracts is shown in fig. 3 and the effect was more pronounced in andrographolide. The reducing capacity of a compound may serve as a significant indicator of its potential antioxidant activity.

The reducing ability of a compound generally depends on the presence of reductants⁴⁸, which exhibit antioxidative property by donating a hydrogen atom and breaking the free radical chain events⁴⁹. The reducing ability was present in all the fractions but significantly in hydroalcoholic extract and in andrographolide.

Fig 3. Reducing power exhibited by various fractions of *Andrographis paniculata* and andrographolide



Values are expressed as mean \pm SD for six individual experiments. Statistically significant difference is expressed as # $p < 0.01$, * $p < 0.05$ when compared to minimal dose.

H^+K^+ ATPase is a prime enzyme that influences secretion of acid in the stomach to aid digestion of proteins. We have found that andrographolide isolated from *Andrographis paniculata* exhibits maximum H^+K^+ ATPase inhibitory action and comparable to that of standard antiulcer drug omeprazole at similar concentrations.

Table 3 shows the H^+K^+ ATPase activity of sheep gastric parietal cells treated with different solvent extracts of *Andrographis paniculata* and omeprazole. Andrographolide exhibited maximum inhibition with the IC_{50} value of $27\mu g/ml$ when compared to that of other extracts. The IC_{50} value of petroleum ether, ethyl acetate, ethanol and hydroalcoholic extracts, andrographolide and omeprazole were found to be $18\mu g/ml$, $23\mu g/ml$, $25\mu g/ml$, $26\mu g/ml$, $27\mu g/ml$ and $29.5\mu g/ml$ respectively.

Table no. 3 H⁺K⁺ ATPase inhibitory activity of various fractions of *Andrographis paniculata* and andrographolide.

| Various Fractions | %inhibition | | | | | | | |
|-------------------|--------------------------|--------------------------|---------------------------|--------------------------|--------------------------|--------------------------|-------------------------|-------------------------|
| | 10 | 20 | 30 | 40 | 50 | 60 | 70 | 80 |
| | (Concentration in µg/ml) | | | | | | | |
| Petroleum ether | 10.6 ± 1.06 | 19.5 ± 1.96 | 23.6 ± 2.37 | 26.8 ± 2.71 | 32 ± 3.6 | 33 ± 3.9 | 34 ± 4.0 | 35.2 ± 4.2 |
| Ethyl acetate | 14 ± 1.4 | 23.6 ± 2.37 | 25.2 ± 2.53 | 27 ± 3.0 | 33.6 ± 3.36 | 34 ± 3.5 | 38 ± 3.9 | 38.9 ± 4.1 |
| Ethanol | 16.8 ± 1.68 | 24.9 ± 2.5 | 26.4 ± 2.7 | 27.6 ± 2.8 | 34.9 ± 3.5 | 35 ± 3.6 | 40 ± 4.0 | 42.1 ± 4.3 |
| Hydroalcoholic | 17.7 ± 1.7* | 25.3 ± 2.6* | 28.7 ± 2.9* | 29.6 ± 3.0* | 35.4 ± 3.6* | 36 ± 3.8* | 45.4 ± 4.6* | 46.4 ± 4.6* |
| Andrographolide | 19.4 ± 1.94 [#] | 26 ± 2.7 [#] | 29.4 ± 3.0 [#] | 32.4 ± 3.4 [#] | 36.9 ± 3.9 [#] | 40.2 ± 4.2 [#] | 46.3 ± 4.7 [#] | 47.2 ± 4.8 [#] |
| Omeprazole | 20 ± 2.0 ^{\$} | 26.2 ± 2.7 ^{\$} | 30.13 ± 3.2 ^{\$} | 34.7 ± 3.5 ^{\$} | 37.5 ± 3.8 ^{\$} | 45.4 ± 4.6 ^{\$} | 48 ± 4.9 ^{\$} | 55 ± 5.6 ^{\$} |

Values are expressed as mean ± SD for six individual experiments. Statistically significant difference is expressed as ^{\$}p<0.001, [#]p<0.01, *p<0.05.

H^+K^+ ATPase is the proton pump responsible for gastric acid secretion^{50, 51}, and the final common pathway mediating secretion of hydrochloric acid by gastric parietal cells⁵². The enzyme typically located in the parietal cells, mediates the electroneutral exchange of intracellular H^+ and extracellular K^+ to achieve acid secretion when parietal cells are under the stimulation of secretagogues⁵³. Gastric H^+K^+ ATPase is a membrane bound enzyme located in the apical membrane of parietal cells which pump protons into the gastric lumen using energy derived from the hydrolysis of ATP⁵⁰. Inhibition of this enzyme results in the inhibition of acid secretion. Thus, H^+K^+ ATPase inhibition test constitutes the best pharmacological target to screen new drugs for ulcer.

In this investigation, it was found that the activity of H^+K^+ ATPase was inhibited *invitro* by the andrographolide in a significant manner when compared to the other extracts. This is on par with omeprazole indicating the potential effect of test material as an inhibitor of acid secretion in stomach parietal cells which can be accounted for its antiulcer activity. The hydroalcoholic extract also was found to inhibit the enzyme significantly when compared to other extracts.

Antioxidants are vital substances which possess the ability to protect the body from free radical induced oxidative stress. Polyphenols present in certain medicinal and dietary plants have helped in preventing oxidative damage thus increasing interest towards natural antioxidants⁵⁴. Phytochemicals are required from a spectrum of sources for their antioxidant role to protect tissues from activities that manifest themselves into chronic diseases⁵⁵.

The quantitative determination of phytochemicals in the hydroalcoholic extract of *Andrographis paniculata* was found to contain flavonoids (total), alkaloids, xanthenes, tannins and triterpenes at the level of 0.0791%, 0.0989%, 0.1621% and 2.156%. Triterpenes were present abundantly (2.156) in *Andrographis paniculata* as shown in table 4.

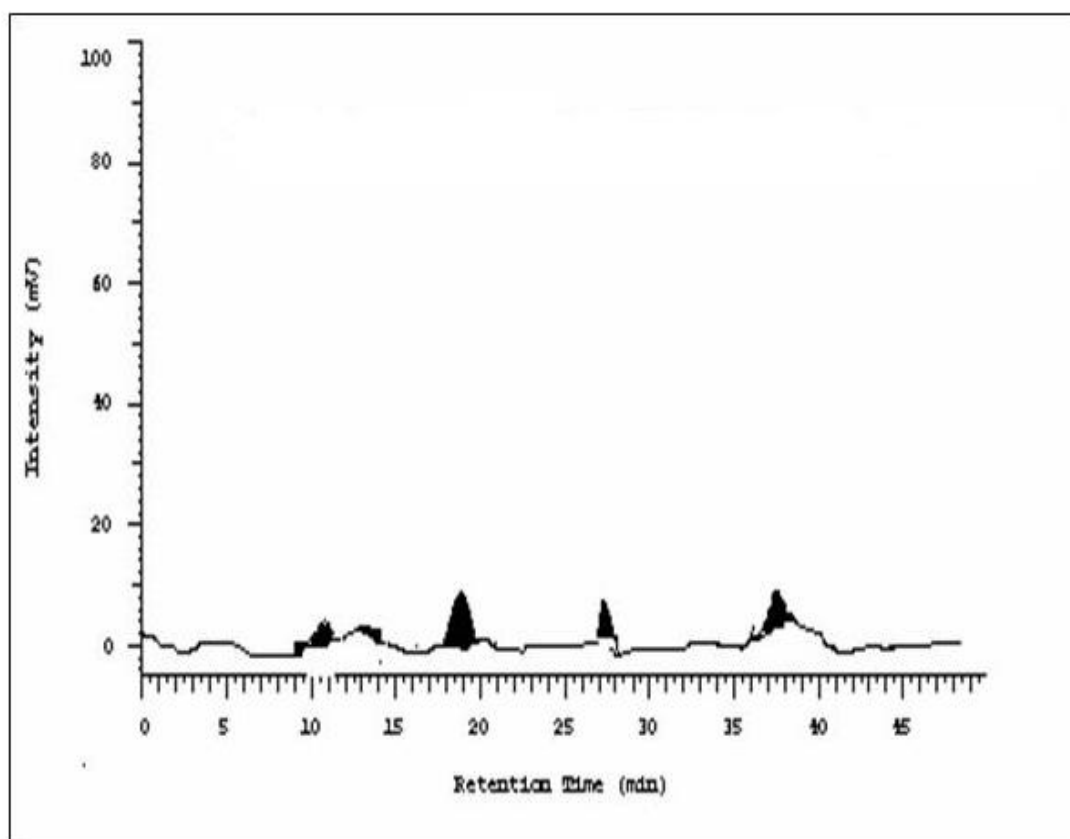
Flavonoids that contain multiple OH⁻ stimulation have very strong antioxidant activities against peroxy radicals⁵⁶. Recent reports and extensive literature survey indicate that many flavonoids and phenolic anti-oxidants possess anti-ulcerogenic and wound healing activity⁵⁷. Some of the flavonoids have been shown to increase the mucosal content and prostaglandin synthesis⁵⁸. Diterpenoids and flavonoids are the main chemical constituents of *Andrographis paniculata* and these compounds are believed to be responsible for the medicinal activities of the plant^{59, 60}.

Table no.4 Quantitative determination of phytochemicals in the hydroalcoholic extract of *Andrographis paniculata*

| | |
|-------------------------|---|
| DESCRIPTION | A BROWN COLOURED LIQUID |
| IDENTIFICATION | POSITIVE FOR FLAVONOIDS, ALKALOIDS, XANTHONES, TANNINS AND TRITERPENES |
| FLAVONOIDS (TOTAL) | 0.0791% W/W BIOCHIN A - 0.0178% QUERCETIN - 0.056% FORMONONETIN - 0.011% |
| ALKALOIDS | 0.0989% W/W |
| XANTHONES | 0.0856% W/W |
| TANNINS | 0.1627% W/W |
| TRITERPENES | 2.156% W/W |

A typical HPLC-UV chromatogram of hydroalcoholic extract of *Andrographis paniculata* was shown in fig.4. Four components were observed in the chromatogram as peaks. Two were identified as Quercetin (18.08 R.T) and Formononetin (39.44) and their retention time being similar with those of authentic standards.

Fig.4 Detection of flavonoids by HPLC-UV chromatogram



Increased gastric acidity and inflammation are considered to be important contributing factors in the pathogenesis of gastric ulcers⁶¹. Table 5 presents the ulcer score measured in the gastric wall of the experimental animals. The ulcer score was significantly reduced in rats pretreated with 3mg/ kg b wt of andrographolide when compared with other solvent extracts. The ulcer reducing effect was comparable with that of the standard drug ranitidine. Ulcer score was reduced significantly in all the experimental model of ulcer.

Table no.5 Ulcer score in ulcerogen animals pretreated with andrographolide and various solvent extracts of *Andrographis paniculata*.

| Treatment protocol | Ulcerogens (Ulcer score) | | |
|--|-----------------------------|--------------------|-------------------|
| | Ethanol | Aspirin | Pylorus ligation |
| Rats treated with ulcerogens | 4.63 ± 0.46 | 4.01 ± 0.44 | 4.70 ± 0.52 |
| Rats pretreated with 200mg/kg b wt of petroleum ether extract of <i>Andrographis paniculata</i> for 30 days +Ulcerogen | 2.6 ± 0.27* (44%) | 3.1 ± 0.34* (23%) | 2.5 ± 0.27* (47%) |
| Rats pretreated with 200mg/kg b wt of ethyl acetate extract of <i>Andrographis paniculata</i> for 30 days +Ulcerogen | 2.7 ± 0.31* (42%) | 2.9 ± 0.34* (28%) | 3.0 ± 0.36* (36%) |
| Rats pretreated with 200mg/kg b wt of ethanol extract of <i>Andrographis paniculata</i> for 30 days +Ulcerogen | 1.8 ± 0.18* (61%) | 2.0 ± 0.22* (50%) | 1.9 ± 0.21* (60%) |
| Rats pretreated with 200mg/kg b wt of hydroalcoholic extract of <i>Andrographis paniculata</i> for 30 days +Ulcerogen | 1.0 ± 0.10* (78%) | 1.2 ± 0.13* (70%) | 1.1 ± 0.12*(77%) |
| Rats pretreated with 3mg/kg b wt of andrographolide for 30 days +Ulcerogen | 0.8 ± 0.08* (83%) | 0.7 ± 0.07* (83%) | 0.9 ± 0.09*(81%) |
| Rats pretreated with 30mg/kg b wt of omeprazole for 30 days +Ulcerogen | 1.0 ± 0.11* (78%) | 1.10 ± 0.12* (73%) | 1.15 ± 0.13*(76%) |

Values are expressed as mean ± SD for six animals in each group. Statistically significant difference is expressed as *p<0.001. (Percentage in parenthesis) Groups are compared as: Ulcerogens vs Rats pretreated with 200mg/kg b wt of petroleum ether extract, ethyl acetate extract, hydroalcoholic extract of *Andrographis paniculata*, 3mg/kg b wt of andrographolide & 30mg/kg b wt of omeprazole for 30 days +Ulcerogen.

Studies suggest that the ethanol induced damage to the GI mucosa starts with microvascular injury, namely a disruption of the vascular endothelium resulting in increased vascular permeability, edema formation and epithelial lifting⁶². Administration of aspirin produces severe gastric haemorrhagic erosions and has not been found to increase the aggressive factors (acid and pepsin) but significantly decreased the gastric output because of so called back diffusion of hydrochloric acid through the broken barrier inhibition of mucosal blood flow and acute inflammation^{63,64}. In pylorus ligated rats gastric acid is associated with severe ulceration of the rat gastric mucosa⁶⁵. Pepsin and hydrochloric acid (HCl) are important for the formation of pylorus ligated ulcers⁶⁶. The rats treated with various solvent extracts / 3mg/kg b wt of andrographolide shows significant reduction in the ulcer score.

In vivo study revealed that andrographolide and the hydroalcoholic extract of *Andrographis paniculata* acted as potent ulcer reducing agents and the effect was comparable to that of standard drug omeprazole. The ulcer score was reduced significantly in all the experimental ulcer models studied. However the detailed study on the level of prostaglandins and gastric mucin in drug treated animals is essential to confirm the antiulcer property.

Conclusion

It can be concluded that that the hydroalcoholic fraction of the aerial parts of *Andrographis paniculata* and andrographolide isolated from the plant possess potent H^+K^+ ATPase inhibitory activity *in vivo* as well as *in vitro*. The potent free radical scavenging activity of the test drugs may probably influence the antiulcer property by preventing the formation and the harmful action of toxic oxygen free radicals on gastric mucosa. The H^+K^+ ATPase inhibitory activity may also be accounted for gastroprotective activity.

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References

1. Shen YC, Chen CF, Chiou WF. Andrographolide prevents oxygen radical production by human neutrophils: possible mechanism(s) involved in its anti-inflammatory effect. *Br J Pharmacol* 2002; 135: 399-406.
2. Amroyan E, Gabrielian E, Panossian A, Wikman G, Wagner H. Inhibitory effect of andrographolide from *Andrographis paniculata* on PAF-induced platelet aggregation. *Phytomedicine* 1999; 6: 27-31.
3. Chang RS, Ding L, Chen GQ, Pan QC, Zhao ZL, Smith KM. Dehydroandrographolide succinic acid monoester as an inhibitor against the human immunodeficiency virus. *Proc Soc Exp Biol Med* 1991; 197: 59-66.
4. Madav S, Tripathi HC, Tandan SK, Mishra S. Analgesic, antipyretic and antiulcerogenic effect of andrographolide. *Indian J Pharm Sci* 1995; 57: 121-125.

5. Vedavathy S, Rao KN. Antipyretic activity of six indigenous medicinal plants of Tirumala Hills, Andhra Pradesh, India. *J Ethnopharmacol* 1991; 33: 193-196.
6. Matsuda T, Kuroyanagi M, Sugiyama S, Umehara K, Ueno A, Nishi K. Cell differentiation-inducing diterpenes from *Andrographis paniculata* Nees. *Chem Pharm Bull (Tokyo)* 1994; 42: 1216-1225.
7. Chiou WF, Lin JJ, Chen CF. Andrographolide suppresses the expression of inducible nitric oxide synthase in macrophage and restores the vasoconstriction in rat aorta treated with lipopolysaccharide. *Br J Pharmacol* 1998; 125: 327-334.
8. Chiou WF, Chen CF, Lin JJ. Mechanisms of suppression of inducible nitric oxide synthase (iNOS) expression in RAW 264.7 cells by andrographolide *Br J Pharmacol* 2000; 129: 1553-1560.
9. Katz J. The course of peptic ulcer disease. *Med Clin North Am* 1991; 75: 831-840.
10. O'Connor HJ. The role of *Helicobacter pylori* in peptic ulcer disease. *Scand J Gastroenterol* 1994; 201: 11-15.
11. Peskar BM, Maricic N. Role of prostaglandins in gastroprotection. *Dig Dis Sci* 1998; 43: 23S-29S.
12. Konturek PC, Brzozowski T, Sliwowski Z, Pajdo R, Stachura J, Hahn EG, Konturek SJ. Involvement of nitric oxide and prostaglandins in gastroprotection induced by bacterial lipopolysaccharide. *Scand J Gastroenterol* 1998; 33: 691-700.
13. Halliwell B, Gutteridge JMC. *Free radicals in Biology and Medicine* (Oxford University Press, New York) 1989.
14. Kannan K, Jain SK. Oxidative stress and apoptosis. *Pathophysiology* 2000; 7: 153-163.
15. Cho CH, Pfeiffer CJ, Misra HP. Ulcerogenic mechanism of ethanol and the action of sulphanyl fluoride on the rat stomach in-vivo. *J Pharm Pharmacol* 1991; 43: 495-498.
16. Lutnicki K, wrobel J, Ledwozyw A, Trebas-Pietras E. The effect of calcium ions and the intensity of peroxidation processes and the severity of ethanol-induced injury to the rat's gastric mucosa. *Arch Vet Pol* 1992; 32: 125-132.
17. Bast A, Haenen GR, Doelman CJ. Oxidants and antioxidants: state of the art. *Am J Med* 1991; 91: 2S-13S.
18. Aruoma OI, Halliwell B, Hoey BM, Butler J. The antioxidant action of N-acetylcysteine: its reaction with hydrogen peroxide, hydroxyl radical, superoxide, and hypochlorous acid *Free Rad Biol Med* 1989; 6: 593-597.
19. Naik SR. Antioxidants and their role in biological functions: An overview. *Indian Drugs* 2003; 40: 501-515.
20. Soares JR, Dinis TCP, Cunha AP, Almeida LM. Antioxidant activities of some extracts of *Thymus zygis*. *Free Rad Res* 1997; 26: 469-478.
21. Langmead L, Rampton DS. Review article: herbal treatment in gastrointestinal and liver disease -- benefits and dangers. *Aliment Pharmacol Ther* 2001; 15: 1239-1252.

22. Piper DW, Stiel DD. Pathogenesis of chronic peptic ulcer, current thinking and clinical implications. *Med Prog* 1986; 2: 7-10.
23. Brito ARMS, Cota RHS, Nunes DS. Gastric antiulcerogenic effects of *Dalbergia monetaria*L in rats. *Phytother Res* 1997; 11: 314-316.
24. Yesilada E, Sezik E, Fujita T, Tanaka S, Tabata M. Screening of some Turkish medicinal plants for their antiulcerogenic activities. *Phytother Res* 1993; 7: 263-265.
25. Batkhuu J, Hattori K, Takano F, Fushiya S, Oshiman K, Fujimiya Y. Suppression of NO production in activated macrophages in vitro and ex vivo by neoandrographolide isolated from *Andrographis paniculata*. *Biol Pharm Bull* 2002; 25: 1169-1174.
26. Viturro C, Molina A, Schmeda-Hischmann G. Free radical scavengers from *Mutisia friesiana* (Asteraceae) and *Sanicula graveolens*(Apiaceae). *Phytother Res* 1999; 13: 422-424.
27. Sreejayan Rao MN. Nitric oxide scavenging by curcuminoids. *J Pharm Pharmacol* 1997; 49: 105-107.
28. Nishkimi M, Appaji N, Yagi K. The occurrence of superoxide anion in the reaction of reduced phenazine methosulphate and molecular oxygen. *Biochem Biophys Res Commun* 1972; 46: 849-854.
29. Halliwell B, Gutteridge JM, Aruoma OI. The deoxyribose method: a simple “test tube” assay for determination of rate constants for reaction of hydroxyl radicals. *Anal Biochem* 1987; 165: 215-219.
30. Bouchet N, Barrier L, Fauconneau B. Radical Scavenging activity and antioxidant properties of tannins from *Guiera senegalensis* (Combretaceae). *Phytother Res* 1998; 12: 159-162.
31. Oyaizu M. Studies on product of browning reaction prepared from glucose amine. *Jap J Nut* 1986; 44: 307-315.
32. Cheon HG, Lim H, Lee DH. Biochemical properties of a newly synthesized H (+)/K (+) ATPase inhibitor, 1-(2-methyl-4-methoxyphenyl)-4-. *Eur J Pharmacol* 2001; 41: 181-186.
33. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 1976; 72: 248-254.
34. Reyes-Chilpa R, Baggio CH, Alavez- Solano D, Estrada-Muniza E, Kauffmann FC, Sanchez RI, et al., Inhibition of gastric H⁺, K⁺-ATPase by flavonoids, coumarins and xanthenes isolated from Mexican medicinal plants. *J Ethnopharmacol* 2006; 105: 167-172.
35. Hollander D, Tarnawski A, Krause WJ, Gergely H. Protective effect of scuralfate against alcohol induced gastric mucosal injury in the rat. Macroscopic, histologic, ultrastructural, and functional time sequence analysis. *Gastroenterology* 1985; 88: 366-374.
36. Goel RK, Chakrabarti A, Sanyal AK. The effect of biological variables on the anti-ulcerogenic effect of vegetable plantain banana. *Planta Med* 1985; 2: 85-88.

37. Sanyal AK, Pandey BL, Goel RK. The effect of cyproheptadine on gastric activity, an experimental study, In: C. J. Pfeiffer (Ed.), Peptic ulcer, Munksguard, Copenhagen 1971; 312-318.
38. Nie SN, Qian XM, Wu XH, Yang SY, Tang WJ, Xu BH, *et al.*, Role of TFF in healing of stress induced gastric lesions. *World J Gastroenterol* 2003; 9: 1772-1776.
39. Halliwell B. Free radicals, antioxidants, and human disease: curiosity, cause, or consequence? *Lancet* 1994; 344: 721-724.
40. Larson RA. The antioxidants of higher plants. *Phytochemistry* 1988; 27: 969-978.
41. Koleva II, van Beek TA, Linssen JP, de Groot A, Evstatieva LN. Screening of plant extracts for antioxidant activity: a comparative study on three testing methods. *Phytochem Anal* 2002; 13: 8-17.
42. Korycka-Dahl M, Richardson T. Photogeneration of superoxide anion in serum of bovine milk and in model systems containing riboflavin and aminoacids. *J Dairy Sci* 1978; 61: 400-407.
43. Moncada S, Palmer RM, Higgs EA. Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol Rev* 1991; 43: 109-142.
44. Sessa WC, Harrison JK, Luthin DR, Pollock JS, Lynch KR. Genomic analysis and expression patterns reveal distinct genes for endothelial and brain nitric oxide synthase. *Hypertension* 1993; 21: 934-938.
45. Halliwell B. Antioxidants and human diseases: a general introduction. *Nutr Rev* 1997; 55: S44-S52.
46. Walling C. Fenton's reagent revisited. *Acc Chem Res* 1975; 8: 125-131.
47. Li C, Xie B. Evaluation of the antioxidant and pro-oxidant effects of tea catechin oxypolymers. *J Agric Food Chem* 2000; 48: 6362-6366.
48. Duh PD, Tu YY, Yen GC. Antioxidant activity of water extract of Harng Jyur (*Chrysanthemum moifolium* Ramat). *Lebensm-Wiss Technol* 1999; 32: 269-277.
49. Gordan MH. The mechanism of the antioxidant action in vitro, In: *Food Antioxidants* (Elsevier London) 1990, 1-18.
50. Sachs G, Chang HH, Rabon E, Schackman R, Lewin M, Saccomani G. A nonelectrogenic H⁺ pump in plasma membranes of hog stomach. *J Biol Chem* 1976; 251: 7690-7698.
51. Wallmark B, Larsson H, Humble L. The relationship between gastric acid secretion and gastric H⁺,K⁺-ATPase activity. *J Biol Chem* 1985; 260: 13681-13684.
52. Yang DH, Tsuyama S, Murata F. The expression of gastric H⁺-K⁺-ATPase mRNA and protein in developing rat fundic gland. *Histochem J* 2001; 33: 159-166.
53. Yao X, Forte JG. Cell biology of acid secretion by the parietal cell. *Annu Rev Physiol* 2003; 65: 103-131.

54. Silva BA, Ferreres F, Malva JO, Dias ACP. Phytochemical and antioxidant characterization of *Hypericum perforatum* alcoholic extracts. *Food Chem* 2005; 90: 157-167.
55. Sheety and Wahlqvist ML. A model for the role of the praline-linked pentose-phosphate pathway in phenolic phytochemical bio-synthesis and mechanism of action for human health and environmental applications. *Asia Pac J Clin Nutr* 2004; 3: 1-24.
56. Alanko J, Riutta A, Holm P, Mucha I, Vapatalo H, Metsa –Ketela T. Modulation of arachidonic acid metabolism by phenols: relation to their structure and antioxidant/prooxidant properties. *Free Radic Biol Med* 1999; 26: 193-201.
57. Soumarmon A, Lewin M J. Gastric (H⁺,K⁺)-ATPase. *Biochimie* 1986; 68: 1287-1291.
58. Shanmugapriya E, Shanmugasundaram E, Venkataraman S. Antiulcerogenic effect of *Justicia prostrata* Gamble. *Indian J Exp Biol* 2005; 43: 181-186.
59. Tang W, Eisenbrand G. Chinese drugs of plant origin: Chemistry, pharmacology and use in traditional and modern medicine. Springer Verlag, Berlin 1992; 97-103.
60. Saxena S, Jain DC, Bhakuni RS, Sharma RP. Chemistry and pharmacology of andrographis species. *Indian Drugs* 1998; 35:458-467.
61. Goa KL, Monk JP. Enprotil: a preliminary review of its pharmacodynamics and pharmacokinetic properties and therapeutic efficacy in the treatment of peptic ulcer diseases. *Drugs* 1987; 3: 539–559.
62. Szabo S, Trier JS, Brown A, Schnoor J. Early vascular injury and increased vascular permeability in gastric mucosal injury caused by ethanol in the rat. *Gastroenterology* 1985; 88:228-236.
63. Akthar MS, Munir M. Evaluation of antiulcerogenic effect of *Solanum nigrum*, *Brassica oleracea* and *Ocimum basilicum* in rats. *J Ethnopharmacol* 1989; 27: 163-172.
64. Tanaka H, Kosaka N, Tomaru A, Shuto K, Oghihara T, Sato N. Augmentation of the gastric mucosal defence mechanism induced by KW - 5805, a novel antiulcer agent. *Scand J Gastroenterol* 1989; 24: 170-173.
65. Martin MJ, Motilva V, Alarcon de la Lastra C. Quercetin and naringenin: effects on ulcer formation and gastric secretion in rats. *Phytotherapy Research* 1993; 7: 150-153.
66. Tan PV, Penlap VB, Nyasse B, Nguemo JD. Antiulcer actions of the bark methanol extract of *Voacanga africana* in different experimental ulcer models in rats. *J Ethnopharmacol* 2000; 73: 423-428.

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