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Study on phytochemical profile and anti-ulcerogenic effect of *Cayratia pedatata Lam* in albino wistar rats

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Summary

The aim of this study is to evaluate the anti-ulcer activity of chloroform extract of leaves of *Cayratia pedata* (Lam). The chloroform extract of *Cayratia pedata* (CECP) was investigated for its anti-ulcer activity against pylorus ligation and ethanol induced gastric ulcers in rats. Ranitidine (50mg/kg,p.o.) and misoprostol (100ug/kg,p.o.) were used as standard drugs. A significant (p<0.01) anti-ulcer activity was observed in both the models. Both does of *Cayratia pedata* produced gastric anti-secretory effect in pylorus ligated rats and also showed gastric cytoprotective effect in ethanol induced gastric ulcers. Pylorus ligation showed significant (p<0.01) reduction in gastric volume. Free acidity, ulcer index as compared to control. It also showed significant (p<0.01) reduction in ulcer index in ethanol induced model with respect to control. This present study indicates that, *Cayratia pedata* leaves extract have potential anti-ulcer activity in this tested models.

Keywords: Cayratia pedata (Lam), Pylorus Ligation, Ethanol Induced Gastric Ulcers.

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Introduction

Peptic ulcer disease (encompassing gastric ulcer and duodenal ulcer) affect a large portion of the world population and are induced by several factors, including stress, smoking, nutritional deficiencies, and ingestion of non-steroidal anti-inflammatory drugs ^[1]. The pathophysiology of these ulcers involves an imbalance between offensive (acid, pepsin, and *Helicobacter pylori*) and defensive factors (mucin, prostaglandin, bicarbonate, nitric oxide and growth factors). Today, there are two main approaches for treating peptic ulcer. The first deals with reducing the production of gastric acid and the second with re-enforcing gastric mucosal protection ^[2,3]. There has been a rapid progress in the understanding of the pathogenesis of peptic ulcer. Modern approach to this includes proton pump inhibitors, histamine receptor blockers, drugs affecting the mucosal barrier and prostaglandin analog ^[4]. Development of tolerance and incidence of relapses and side effects on clinical evaluation make their efficacy arguable. This has been the basis for the development of new antiulcer drugs, which includes herbal drugs.

Cayratia pedatata lam (Family- Vitaceae) Endemic to southern western ghats in Kerala and Tamil Nadu. In Kerala, reported only from Palakkad forest in Silent Valley, where it is locally common. In Tamil Nadu, collected only from Nilgiris. It may possibly occur in the high ranges of Anamalais and Kodaikanal hills ^[5,6]. In folklore claim, it is used for treating diarrhea, burns, refrigerant, ulcers, uterine, flukes, hysteria (India) Astringent, ^[5,7], uterine reflexes (Sri Lanka) low diuretic activity and has been a reputed remedy for cough, bronchitis, asthma, joint pain and to check uterine reflexes ^[7,8]. From the source of literature documentation and relevant traditional approaches on plant drugs, the present investigation was carried out to investigate the constituents and anti-ulcer profile of the chloroform extract of *Cayratia pedatata lam* (CECP) is being reported here.

Materials and Methods

Collection and authentication of plant material

The leaves of *Cayratia pedata lam* were collected from the Palakadu forest in Kerala, in the month of September. The plant material was identified and authenticated by Mr. Chelladurai, Research officer- Botany, C.C.R.A.S. Govt of India, Tirunelveli. The voucher specimen of the plant was deposited in the college for further references.

Preparation of plant extract

Freshly collected leaves of *Cayratia pedata lam* were dried to get a coarse powder. A weighed quantity of the powder was passed through sieve number 40 and extracted using soxhlet apparatus with chloroform as a solvent at a temperature range of $60^{\circ}C^{47}$. The extract was concentrated by distilling off the chloroform and then evaporating to dryness on a water bath. Before and after every extraction the powder bed was completely dried and weighed. The percentage yield of chloroform extract was 18.4%W/W. The chloroform extract was administered to the animals by suspending each time in 1% CMC.

Phytochemical Screening

The phytochemical examination of chloroform extract of *Cayratia pedatata lam* was performed by the standard methods ^[9].

Experimental animals

Adult Wistar rats of either sex weighing 180-250 gms were used in pharmacological and toxicological studies. The inbred animals were taken from the animal house in C.L.Baid Metha College of Pharmacy, Thoraipakkam, Chennai-97. The animals were maintained in a well ventilated room with at 12:12 hr light, dark cycle in polypropylene cages and maintained at $22\pm1^{\circ}$ C with humidity at $55\pm5^{\circ}$. They were fed balanced rodent pellet diet from Poultry Research station, Nandanam, Chennai-35 and tap water ad *libitum* throughout the experimental period. The experimental protocol was approved by the Institutional Animal Ethics Committee (IAEC) of CPCSEA (Committee for the Purpose of Control and Supervision of Experimental Animals). (Regd.No.930/a/06/CPCSEA. Dt. 30-06-2006.

Acute toxicity study

The acute toxicity of chloroform extract of *Cayratia pedatata lam leaves* was determined as per the OECD guideline no. 423 (Acute Toxic Class Method). It was observed that the test extract was not lethal to the rats even at 2000mg/kg dose. Hence, 1/10th (200mg/kg) and 1/5th (400mg/kg) of this dose were selected for further study ^[10].

ANTIULCER ACTIVITY

Pylorus ligated model

Wistar rats of either sex weighing 180 to 250 g were divided into five groups of six animals each.

GROUP I Control animals which are pretreated with suspended 1% CMC, ml/100g.

- GROUP II Pylorus ligated animals which are pretreated with 1% W/V CMC, 1ml/100g.
- GROUP III Pylorus ligated animals which are pretreated with CECP (200mg/kg p.o) suspended in 1%w/v CMC.
- GROUP IV Pylorus ligated animals which are pretreated with CECP (400mg/kg p.o) suspended in 1%w/v CMC.
- GROUP V Pylorus ligated animals which are pretreated with Ranitidine (50mg/kg, p.o) suspended in 1%w/v CMC.

PROCEDURE

Groups III and IV animals were treated with CECP and group V were treated with ranitidine for the respective 5 days daily. On day 6^{th} after the last dose, all the groups were kept for 18h fasting and care was taken to avoid coprophagy ^[11]. Animals were anaesthetized using ether the abdomen was opened and pylorus ligation was done without causing any damage to its blood supply. The stomach was replaced carefully in the abdomen and the wound was sutured by interrupted sutures. After 6 hr⁵³, stomachs were dissected out and cut open along the greater curvature and the content were drained into small beakers, centrifuged and then subjected to analysis for gastric volume, free acid, total acid and protein estimation ^[12]. The mucosa was flushed with saline and the stomach was pinned on frog board and the ulcer score was calculated.

Ethanol Induced Ulcer

Wistar rats of either sex weighing 180 to 250 g were divided into five groups of six animals each.

GROUP I Control animals which are pretreated with suspended 1% CMC, 1ml/10)0g
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- GROUP II Ethanol (1ml/200g p.o) induced ulcer which are pretreated with 1% W/V CMC, 1ml/100g.
- GROUP III Ethanol (1ml/200g p.o) induced ulcer which are pretreated with CECP (200mg/kg /p.o) suspended in 1%w/v CMC.
- GROUP IV Ethanol (1ml/200g p.o) induced ulcer which are pretreated with CECP (400mg/kg /p.o) suspended in 1%w/v CMC.

GROUP V Ethanol (1ml/200g p.o) induced ulcer which are pretreated with Ranitidine (50mg/kg/p.o) suspended in 1%w/v CMC.

Procedure

Groups III and IV animals were treated with CCEP and group V animals were treated with ranitidine for the respective 14 days. On day 15th after the last dose, all the groups were kept for 24h fasting and care was taken to avoid coprophagy⁵⁶. The 100 % ethanol (1ml/200g p.o) was administered to the groups II, III, IV and V animals and after 1 h animals were sacrificed by cervical dislocation and stomachs were dissected out and cut open along the greater Curvature and examined for ulcer ^[13].

Measurement of ulcer index

The stomachs were excised and were examined for hemorrhagic lesions in glandular mucosa. Immediately after the animals were sacrificed, their stomachs were dissected out, cut along the greater curvature and the mucosa were rinsed with cold normal saline to remove blood contaminant, if any. The sum of the length (mm) of all lesions for each stomach was used as the ulcer index (UI), and the percentage of inhibition (%*I*) was calculated as described by Nguelefack et al. ^[14] using the following formula:

 $\%I = \frac{(USc - USt)}{USc} \times 100$

Where USc = ulcer surface area in control and USt = ulcer surface area in treated animals.

Histological examination

Two animals from each group were sacrificed and the stomach was isolated, washed with saline and preserved in 10% formalin for histopathological studies. The sections of the stomach, stained with hematoxylin and eosin, were assessed for histopathological changes such as congestion, edema, hemorrhage and necrosis. Histological measurement and photographs were taken (magnification 100 X).

BIOCHEMICAL ESTIMATIONS

Pylorus ligated model

Determination of gastric volume

After sacrificing the rat, the stomach portion was removed. The gastric contents were transferred in to the centrifuge tube, and centrifuged at 1000 rpm for 10 minutes. The supernatant liquid was then transferred to a measuring cylinder, and the volume was measured.

Determination of pH of gastric content

1 ml of the gastric juice was collected, and pH was directly measured by using pH strip^[15].

Determination of free acidity and total acidity

The total volume of gastric content was measured. The gastric contents were centrifuged and filtered. One ml of the gastric juice was pipetted out and the solution was titrated against 0.1N sodium hydroxide using 2 to 3 drops of Topfer's reagent as indicator, to the endpoint when the solution turned to yellowish orange colour was observed. This indicated the volume of NaOH required neutralizing the free hydrochloric acid present in the gastric juice. Then 2 to 3 drops of phenolphthalein solution was added and titration was continued until a definite red tinge

reappears. The difference between the two readings indicated the volume of NaOH required neutralizing the combined acid present in the gastric juice. The sum of the two titrations was the total acid present in the gastric juice ^[16]. Acidity was calculated by using formula;

m. Eq. /dl.

Vol. of NaOH X Normality of NaOH

Acidity =

Vol. of Gastric juice used

Estimation of total proteins

Reagents

Alkaline copper reagent

Solution A: 2% sodium carbonate in 0.1N sodium hydroxide

Solution B: 0.5% copper sulphate in 1% sodium potassium tartarate. 50 ml of solution A was mixed with 1 ml of solution B just before use. Folins phenol reagent. One volume of folins reagent was diluted with two volumes of distilled water just before use.

Standard bovine serum albumin

20 mg of bovine serum albumin was dissolved in 100 ml of distilled water. Few drops of NaOH was added to it aid complete dissolution of bovine serum albumin and to avoid frothing, it was allowed to stand overnight in a refrigerator.

Procedure

The dissolved proteins in gastric juice were estimated in the alcoholic precipitate obtained by adding 90 % of alcohol with gastric juice in 9: 1 ratio respectively. Then 0.1 ml of alcoholic precipitate of gastric juice was dissolved in 1 ml of 0.1 N NaOH and from this 0.05 ml was taken in another test tube. To this 4 ml of alkaline copper reagent was added and kept for 10 minutes. Then 0.5 ml of phenol reagent was added and again 10 minutes was allowed for color development. Reading was taken against blank prepared with distilled water at 640 nm. The protein content was calculated from standard curve prepared with bovine albumin and has been expressed in terms of $\mu g/ml$ of gastric juice ^[17].

Statistical Analysis

The data represents Mean \pm SEM. Results were analyzed statistically using one way ANOVA followed by Dunnet's 't' test. The minimum level of significance was set at (P<0.05).

Results

Phytochemical investigation

The results of preliminary phytochemical investigation of the chloroform extract of Cayratia pedatata lam leaves (CECP) shows the presence of carbohydrates, steroids, sterois, phenols, flavanoids, glycosides, terpenes and absence of alkaloids, tannins, gum and mucilage and Saponins.

Acute toxicity study

Acute toxicity study in which the animals treated with the CECP at a higher dose of 2000 mg/kg did not manifest any significant abnormal signs, behavioral changes, body weight changes, or

macroscopic findings at any time of observation. There was no mortality in the above-mentioned dose at the end of the 14 days of observation.

Effect of CECP on gastric ulcer induced by pylorus ligation Gastric volume

A significant (P<0.01) increase in the gastric volume level is observed in the pylorus ligated group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.01) decrease in the gastric volume level and ranitidine treated group have shown a significant (P<0.01) decrease in the gastric volume level when compared to the Pylorus ligated group. The results were shown in Table 1.

pН

A significant (P<0.01) decrease in the pH level is observed in the pylorus ligated group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.05) and (P<0.01) increase in the pH level and ranitidine treated group have shown a significant (P<0.01) increase in the pH level when compared to the Pylorus ligated group. The results were shown in Table 1.

Ulcer number

A significant (P<0.01) increase in the ulcer number is observed in the pylorus ligated group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.01) decrease in the ulcer number and ranitidine treated group have shown a significant (P<0.01) decrease in the ulcer number when compared to the Pylorus ligated group. The results were shown in Table 1.

Ulcer severity score

A significant (P<0.01) increase in the ulcer severity score is observed in the pylorus ligated group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.01) decrease in the ulcer severity score and ranitidine treated group have shown a significant (P<0.01) decrease in the ulcer severity score when compared to the Pylorus ligated group. The results were shown in Table 1.

Free Acidity

A significant (P<0.01) increase in the free acidity is observed in the pylorus ligated group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.01) decrease in the free acidity and ranitidine treated group have shown a significant (P<0.01) decrease in the free acidity when compared to the Pylorus ligated group. The results were shown in Table 1.

Total acidity

A significant (P<0.01) increase in the total acidity is observed in the pylorus ligated group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.05) and (P<0.01) decrease in the total acidity and ranitidine treated group have shown a significant (P<0.01) decrease in the total acidity when compared to the Pylorus ligated group. The results were shown in Table 1.

Total protein

A significant (P<0.01) increase in the total protein is observed in the pylorus ligated group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.05) and (P<0.01) decrease in the total protein and ranitidine treated group have shown a significant (P<0.01) decrease in the total protein when compared to the Pylorus ligated group. The results were shown in Table 1.

Effect of CECP on gastric ulcer induced by Ethanol Ulcer index

A significant (P<0.01) increase in the ulcer index is observed in the ethanol induced group when compared to the control group. CECP 200mg/kg and 400mg/kg treated group have shown a significant (P<0.01) decrease in the ulcer index and ranitidine treated group have shown a significant (P<0.01) decrease in the ulcer index when compared to the ethanol induced group. The results were shown in Table 2.

Discussion

The antiulcer activity of CECP was studied in different models LIKE ulcers induced by pylorus ligation and ethanol. Although the etiology of gastric ulcer is not known in most cases, it is generally accepted that it results from an imbalance between aggressive factors and the maintenance of mucosal integrity through the endogenous defense mechanisms^[18].

It is well known that pylorus ligation causes gastric hypersecretion due to poorly understood mechanisms. The activation of the vagus-vagal reflex by stimulation of pressure receptors in the antral gastric mucosa in pylorus ligature model is believed to increase gastric tonus and secretion ^[19]. Digestive effect of the accumulated gastric juice is belived to be responsible for producing ulcers in the pylorus ligated rats. Pylorus ligated induced ulcers are thought to be caused due to increased presence of acid and pepsin in the stomach. The essential criteria, which determine the status of mucosal defense barrier against the offensive assault of acid-pepsin is the quality and quantity of gastric mucus secretion. Increased mucus secretion by the gastric mucosal cells can prevent gastric ulceration by several mechanisms including lessening stomach wall friction during peristalsis and acting as an effective barrier to the back diffusion of hydrogen ions ^[20].

The CECP reduces the gastric volume, ulcer score, free and total acidity of gastric acid secretion of pylorus ligature induced gastric ulceration in rats. From results it is clear that the CECP exhibited a significant gastric acid secretion activity by reducing the secretory parameters when compared with the Pylorus ligated group. CECP pretreatment have shown a significant reduction in protein levels when compared with group II. Hence, the protection by CECP against gastric ulcers induced by Pylorus Ligated appears to be produced by the suppression of pepsin levels and strengthening of mucosal barrier. Further, it is clear that the antiulcer activity against this model was again found to be more with CECP extract.

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Table 1. Effect of CECP on acid secretary parameters in pylorus ligation induced gastric ulcer

Comparisons were made between: a- (Group I vs II), b- (Group II vs III, IV), C-(GroupII vs V)

Groups	Gastric Volume (ml/100g)	рН	Number of Ulcer	Ulcer severity score	Free acidity (mEq/dl)	Total acidity (mEq/dl)	Total protein (μg/ml)
I Control (CMC)	2.367 ± 0.15	2.91±0.15	0.1667±0.16	0.1667 ±0.1667	4.4480± 0.39	5.505 ± 0.0291	307.80 ± 8.01
II Pylorus Ligated	7.033± 0.28a**	2.16 ±0.10a**	4.500±0.22a**	9.667± 0.61a**	7.617± 0.21 a**	8.358 ± 0.21a**	496.10 ± 6.45 a **
III CECP 200mg/kg	6.333± 0.17b**	$2.417 \pm 0.15b*$	3.500±0.34b*	7.333 ±0.55b**	6.598±0.11b**	7.928 ±0.35b*	385.64 ±5.89 b*
IV CECP 400mg/kg	5.817 ± 0.11b**	$3.00 \pm 0.18b^{**}$	2.166±0.16b**	3.667±0.61b**	4.405± 0.28b**	$5.45 \pm 0.20b^{**}$	325.26 ± 8.45 b **
V Ranitidine50mg/kg	$3.717 \pm 0.13c^{**}$	$3.583 \pm 0.15c^{**}$	1.000±0.25c**	1.167± 0.30c**	3.058±0.11c**	$4.038 \pm 0.12c^{**}$	$262.45 \pm 6.76 \text{ b **}$

Values are expressed as mean \pm SEM of 6 animals. Statistical Significance test for comparison was done by ANOVA followed by Dunnet's 't' test. Symbols represent statistical significance: ** P < 0.01, * P < 0.05, ns- non a significant.

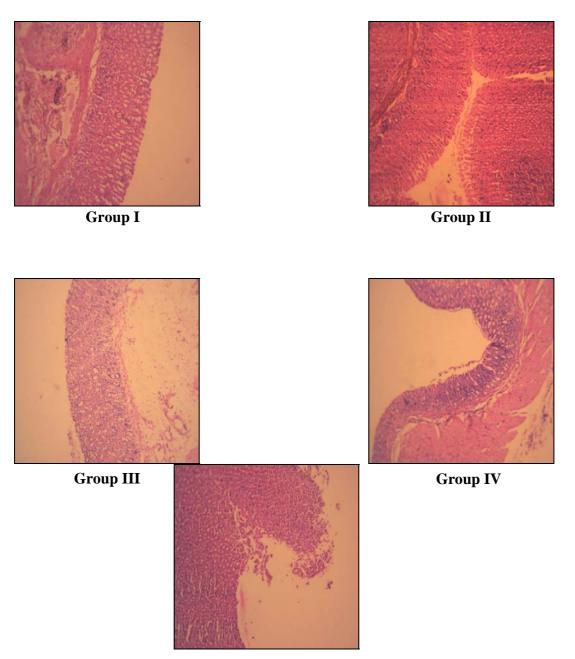
Groups	Ι	II	III	IV	V	
	Control (CMC)	Pylorus Ligated	CECP 200mg/kg	CECP 400mg/kg	Ranitidine 50mg/kg	
Ulcer index(mm ²)	0.333±0.33	41.50 ±1.4a**	$29.50 \pm 0.92a^{**}$	18.67 ± 1.92b**	9.000 ± 1.0C**	

Comparisons were between: a- (Group I vs II), b- (Group II vs III, IV), C-(GroupII vs V). Values are expressed as mean \pm SEM of 6 animals. Statistical Significance test for comparison was done by one way ANOVA followed by Dunnet's 't' test.

Symbols represent statistical significance: ** P < 0.01, * P < 0.05, ns- non a significant.

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Figure 1. Histopathology of pylorus ligated gastric ulcer stomach





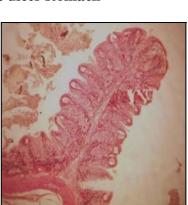
- I Showing No Ulcer in Gastric Mucosa
- II Ulcer induced Gastric Mucosa showing ulcerated focus with detatching mucosal fragments
- III shows stomach mucosa at ulcer site superficial epithelium normal submucosa shows oedema with congested vessels.
- IV Regenerating epithelium, compact glands .No sub mucosal oedema.
- V Hyperplastic Gastric Mucosa with no ulceration

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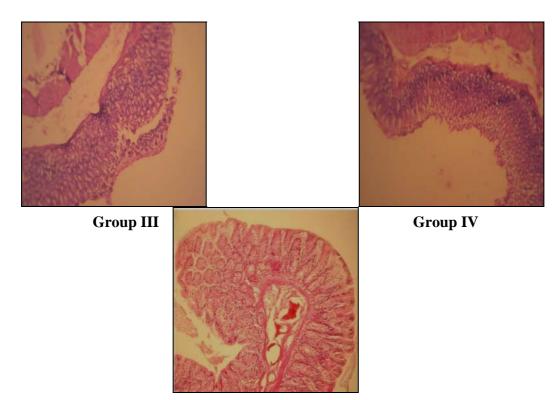
Figure 2. Histopathology of ethnol induced gastric ulcer stomach



Group I



Group II



Group V

- I II Showing No Ulcer in Gastric Mucosa
- Ulcer induced Gastric Mucosa showing ulcerated focus with detatching mucosal fragments. Submucosal oedema.
- III
- ulcer area showing regenerating epithelium. Submucosal oedema Present. The presence of regenerating epithelium. No congestion and inflammatory cells showing minimus IV gastric mucosal ulcer.
- V Hyperplastic Gastric Mucosa with mimimus ulceration

Oral treatment with ethanol causes focal hyperemia, edema, necrosis and submucosal hemorrhage as well as circulatory disturbances. The extent of ethanol-induced gastric mucosal damage in rats correlates with the number of degranulating mast cells. Since these cells are a source of several neuropeptides and inflammatory mediators, including histamine and leukotrienes ^[21]. Ethanol induced gastric lesion formation may be due to stress in the gastric blood flow which contributes to the development of hemorrhage and necrotic aspects of tissue injury. Ethanol also increases Na⁺ and K⁺ flux into the lumen and increases pepsin secretion along with the histamine release.HCL further deepens the necrosis and increases the tissue injury⁶⁵. Ethanol induced ulcer are more predominant in the glandular part of the stomach stimulates leukotrienes, 5-lioxygenase pathway, mast cell secretary product and breakdown of reactive oxygen species resulting in the damage of gastric mucosa.Another action promoted by ethanol is its ability to damage the gastric mucosa by mechanical injury^[22].

The results further indicate that CECP may enhance gastric mucosal defensive factor, such as mucus and or prostaglandin. Therefore protection afforded by CECP against ethanol induced gastric ulceration could also be due to inhibition of the 5-lipoxygenase pathway or to the antagonistic activity of leukotrienes. Flavonoids are capable of protecting the gastric mucosa from necrotizing substances and possible useful in the therapy of acute and chronic gastric ulcerations. plant-originated flavanoid substances such as Solon-Sophoradin root, seed extract of Amaranth, extract of grapefruit seeds - Citro, and capsaicin present in chilli pepper extract are beneficial and dose-dependent reduction in acute and chronic gastric lesions. Plant-originated flavonoid substances are highly gastroprotective ^[23].

The Antiulcer effect of the lipid components of M. azedarach fruits which is mainly due to the phytosterol fraction ^[24,25]. Sterols may even protect against peptic ulcer diseases even in the area of prevalence of H.pylori. Phytosterol esters and sterols, in horse gram an herb in the genus Dolichos cultivated in India for food and fodder were protective for ulcers ^[26].

Phytochemical studies of the chloroform extract of *Cayratia pedata Lam* revealed the presence of flavonoids, sterols and terpenes. Numerous flavonoids have shown reducing gastric acid secretion and protective properties in different experimental model. Phytosterols which have been shown various properties that is necessary for protection against ulcer induction. Since flavonoids and sterols shown to be present in the, chloroform extract of *Cayratia pedata Lam* these constituents may be responsible for the anti-ulcer activity of the chloroform extract of *Cayratia pedata* Lam.

Therefore, it can be concluded that this crude extract have a potential to be used as an antiulcer drug in combination with other drugs or alone. Though the mechanism of the Anti-ulcer action of the plant extract remains to be studied in detail. Isolation of the active constituents may be carried in future.

References

- 1. Nash J, Lambert L, Deakin M. Histamine H2-receptor antagonists in peptic ulcer disease. Evidence for a prophylactic use. *Drugs*. 1994; 47: 862–871.
- 2. Hoogerwerf WA, Pasricha PJ. Agents used for control of gastric acidity and treatment of peptic ulcers and gastroesophageal reflux disease. In: Hardman JG, Limbird LE, Goodman

Gilaman A (Eds.). The Pharmacological Basis of Therapeutics, tenth ed. Mc Graw-Hill, New York, 2001; 1005–1019.

- Valle DL. Peptic ulcer diseases and related disorders. In: Braunwald E, Fauci AS, Kasper DL, Hauser SL, Longo DL, Jameson JL. (Eds.), Harrison's Principles of Internal Medicine, vol. 16. McGraw-Hill, New York, 2005; 1746–1762.
- 4. Manonmani S, Viswanathan VP, Subramanian S, Govindasamy S. Biochemical studies on the antiulcerogenic activity of cauvery 100, an ayurvedic formulation in experimental ulcers. *Indian Journal of Pharmacology*. 1995; 27: 101–105.
- 5. Asia medicinal plant database, National Agro Biodiversity Center, RDA.
- 6. Environmental information center; FRLHT'S, Medicinal plants for conservation concern.
- 7. FRLHT'S-Encyclopaedia of Indian herbal medicine plants, 213.
- 8. Joy PP, Samuel Mathew, Baby P. Skaria, Kerala Agricultural University Aromatic and Medicinal Plants Research Station Odakkali, Asamannoor P.O Ernakulam District, Kerala, India.1998.
- 9. Harborne JP. Phytochemical methods, a guide to modern technique of plant analysis (*Chapmann and Hall, London*). 1973; 1-271.
- 10. OECD, 2002. Acute oral toxicity. Acute oral toxic class method guideline 423 adopted 23.03.1996. In: Eleventh Addendum to the, OECD, guidelines for the testing of chemicals organisation for economical co-operation and development, Paris, June, 2000.
- 11. Rao CHV, Verma R, Vijayakumar M, Rastogi S. Gastroprotective effect of Standardized Extract of *Ficus glomerata* Fruit on Experimental Gastric Ulcers in Rats. *Journal of Ethnopharmacology*, 2008; 115: 323–326.
- 12. Pallavi bafna, Subhash bodhankar. Gastrointestinal Effects of Mebarid an Ayurvedic Formulation in Experimental Animals. *Journal of Ethnopharmacology*, 2003; 86: 173–176.
- 13. Devendra shirode, Tushar patel, Samaresh pal Roy, Jyothi TM, rajendra SV, Prabhu, Ramachandra settyanti. Ulcer Properties of 70% Ehanolic Extract of Leaves of Albizzia Lebbeck. *Phcog Mag.*, 2008; 4: 228-231.
- 14. Nguelefack TB, Watcho P, Wansi SL, Nguelta MM, Ngamga D, Tane P, Kamanyi A. The antiulcer effect of the methanol extract of the leaves of *Aspilia africana* (Asteraceae) in rats. *African Journal of Traditional Complementary and Alternative Medicines*. 2005; 2: 233–237.
- 15. Arun M, Asha VV. Gastroprotective Effect of Dodonaea Viscose on Various Experimental Ulcer Models. *Journal of Ethnopharmacology*, 2008; 24: 1-6.
- 16. Muhammad Jan, Farah Faqir, Hamida Qureshi, Salman Akbar Malik, Muhammad Azhar Mughal. Evaluation of Effects of Extract from Seeds of Myristica Fragrans on Volume and Acidity of Stimulated Gastric Secretion, Liver and Kidney Function. The Journal of Postgraduate Medicinal Institute. 2004; 18: 644-650.
- 17. Lowery OH, Rosenbrough NJ, Farr A and Randall RJ Protein Measurment with Folin Phenol Reagent. *J Biol Chem.*, 1993; 265-275.
- 18. Ramakrishna D, Pavan Kumar K, Mukkanti K, Abedulla Khan K.Antiulcer Activity of The Seeds of *Entada Phaseoloides*. *Pharmacologyonline*, 2008; 3: 93-99.
- 19. Cristiane Hatsuko Baggio, Cristina Setim Freitas, Lia Rieck, Maria Consuelo, Andrade Marques. Gastroprotective effects of a crude extract of Baccharis illinita DC in rats, *Pharmacological Research*, 2003; 47: 93–98.

- 20. Manish Rachchh A, Sunita Jain M. Gastroprotective effect of Benincasa hispida fruit extract. *Indian J Pharmacol.*, 2009; 40: 271-5.
- 21. Leônia Maria Batista, Ana Beatriz Albino De Almeida, Luciana De Pietro Magri, Walber Toma, Tamara Regina Calvo, Wagner vilegas, Alba Regina Monteiro Souza Brito. Gastric Antiulcer Activity of Syngonanthus Arthrotrichus Silveira. *Biol. Pharm. Bull.*, 2004; 27: 328-332.
- 22. Hiruma-lima CA, Gracioso JS, Toma W, Almeida AB, paula. Gastroprotective effect of Aparisthman. A terpene Isolated from Aparisthmium Cordatum, on Experimental Gastric Ulcer Models in Rats and Mice. *Phytomedicine*, 2001; 8(2): 94-100.
- 23. Singh S, Khajuria A, Singh J, Taneja SC. The Gastric Ulcer Protective Effect of Boswellic Acids a Leukotrine Inhibitor from Boswellia Serrata in Rats. *Phytomedicine*, 2008; 15: 408-415.
- 24. Zayachkivska OS, Konturek SJ, Drozdowicz D, Konturek PC, Brzozowski T, Ghegotsky MR.Gastroprotective Effects of Flavonoids in Plant Extracts. *Journal of Physiology and Pharmacology*, 56, 2005; 56: 219-231.
- 25. Hanifa Moursi SA, Izaldin Al Khatib MH. Effect of Melia Azedarach Fruits on Gipsing-Restraint Stress-Induced Ulcers In Rats. *Japan J Pharmacol.*, 1984; 36: 527-533.
- 26. Alvin Berger, Peter Jones JH, Suhad Abumweis S. Plant Sterols: factors Affecting their Efficacy and Safety as Functional Food Ingredients, 2007.