

## COMPARATIVE ANTIOXIDANT POTENTIAL OF MYRTUS COMMUNIS LINN LEAVES AQUEOUS AND ORGANIC EXTRACTS

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### Abstract

The total phenol content, flavonoid, condensed tannins and free radical scavenging potential of aqueous, chloroform, n-hexane and methanol extracts of *Myrtus communis* (*M. communis*) leaves were evaluated. Folin-Ciocalteu method was applied to determine phenolic compound, colorimetric method was used to quantify flavonoids and Vanillin assay procedure was used for condensed tannin estimation. While antioxidant activity was measured by standards In Vitro antioxidant test systems i.e. H<sub>2</sub>O<sub>2</sub> scavenging activity, Superoxide dismutase (SOD)-like activity and NO scavenging activity uses ascorbic acid as standard. The highest total phenolic content (gallic acid equivalents/g of extract), total flavonoids (rutin equivalents/g of extract) and condensed tannins (catechin equivalents in mg CE/g) were measured in methanol extract i.e. 120±3 mg, 6.5±1 mg and 14±1 mg respectively.

Methanol extract at 250µg/mL remarkably inhibited hydrogen peroxide scavenging activity (84±1%), superoxide dismutase-like activity (85±0.5%) and nitric oxide scavenging activity (75±1%). These results indicated that *Myrtus communis* leaves extracts have potent antioxidant activities that would have beneficial effects on human health and methanol extracts are superior with better antioxidant potential.

**Keywords:** *Reactive oxygen species, diseases, natural antioxidant, leaves extracts, phenols and flavonoids.*

**Introduction:**

Reactive species such as OH<sup>-</sup>, H<sub>2</sub>O<sub>2</sub>, O<sup>-2</sup> radicals and singled 1O<sub>2</sub> are biological metabolites synthesized by aerobic microorganisms as a product of the metabolism of oxygen. These Reactive oxygen species (ROS) are highly reactive and unstable. Too much production of such molecular components can cause damage to DNA, proteins, cell membranes and lipids through chain reactions [1]. Antioxidants, molecules with a radical-scavenging capacity, are considered to exercise a defensive consequence against free radical injury. These molecular components may add to prevent many chronic diseases like as cancer, hepatitis, asthma, atherosclerosis, arthritis, heart disease and diabetes [2]. Currently many synthetic antioxidants are also used. These antioxidants have been supposed to damage liver and cause cancer [3]. Due to side effects and toxicity to health, there utilization is restricted [4]. The consumption of plant based additives as antioxidant increases day by day [5]. The herbal based antioxidant demand increases in the market, as they are used in food, cosmetic and pharmaceutical industries to replace synthetic antioxidants. Several studies have shown that medicinal plants contain various natural products possessing antioxidant properties with useful health effects.

Among natural antioxidants, phenolic substances have been of special interest because they are found extensively in plants [6]. Natural products containing phenolic substances have antioxidant potential, mostly because of their redox ability, which enable them to behave as hydrogen donor, single oxygen quenchers and reducing agents as well as they have a potential to chelate metals [4]. The phenolic substances have taken part in the body cell protection from harm by H<sub>2</sub>O<sub>2</sub>, lipid peroxide, unsaturated fatty acid, neutralizing and absorbing free radicals[7].

*Myrtus communis* (*M. communis*) is an evergreen small tree. It is mostly distributed in America, Africa, Asia and Europe. A refreshing and aromatic smell is emitted by its leaves and its taste is strongly bitter, quite unpleasant and intensive [8]. This herb is conventionally utilized for the treatment of urinary tract infection, dry coughs, vaginal discharge, stomach problems, sinusitis and respiratory system.

The leaves are balsamic, haemostatic, tonic and flavoring agent in dishes [9].

Currently the research interest is increased in herbal medicine because they have an excellent source of natural antioxidants and to utilize it in pharmaceutical and food industries to substitute artificial antioxidants. The current study was aimed to assess the phenol, flavonoids, condensed tannins and scavenging potential of n-hexane, methanol, aqueous and chloroform extracts of *M. communis* leaves.

**Materials and Methods:**

Fresh leaves of *M. communis* were collected from Medicinal Botanical Garden of Pakistan Council of Scientific and Industrial Research (PCSIR) Labs Complex, Peshawar-Pakistan. The collected leaves were dried in an Air Cabinet Dryer (England) at 35 °C for 3 days and were ground in a Waring® Commercial Laboratory Blender USA. Leaves powdered material was kept in airtight plastic bags at 4°C.

**Solvent extraction:**

100 gram leaves powder was extracted using one liter of methanol, n-hexane and chloroform separately at room temperature for seven days. While for water extraction preparation, dried sample 100 gram was extracted using one liter of hot water for twenty minutes on water bath. This process of extraction was repeated three times for each sample material. The liquid extracts prepared in different solvents were filtered through Watman No. 1 filter paper and the Buchi Heating Bath B-490, Buchi Rotavapor R-200, Switzerland rotary evaporator was used to remove solvents and a dry extract was obtained. For further processing, dried extracts were kept at 4 °C.

**Phenolic content determination**

Phenolic quantification in dry extract was estimated through Ultra violet Spectrophotometer (U-2900 Hitachi Tokyo Japan). 100 microgram extract/ml solvent of plant extract was prepared and from this sample took 0.5 ml solution, added 2.5 ml Folin-Ciocalteu reagent to it and then two milliliter of Na<sub>2</sub>CO<sub>3</sub> (75 g/l) was added after 0.5 to 8 min interval. The mixture sample was kept in incubator at 50 °C for five minute. As a control, distilled water 0.5 ml was used. Absorbance was taken at 760 nm, when the solution becomes blue-colored. Gallic acid

dissolved in methanol and then measured reading to draw the standard curve. The average of three readings ( $\pm$ SD) was expressed as GAE (gallic acid equivalents) in mg/g dry myrtle extract [4].

#### Total flavonoid content

In a test tube add 1.25 ml of distilled water and mix individually with 0.25 ml rutin (15-250 mg/ml) and plant extract (0.625– 5 mg/ml) and after that five percent sodium nitrite of 75  $\mu$ l was added. Subsequent to six minutes added 150  $\mu$ l AlCl<sub>3</sub> (10%) and this blend was kept for five minutes to react, after that added 0.5 ml of sodium hydroxide solution (1M). Before mix well added distilled water to the mixture to prepared the mixture up to 2.5 ml. At 510 nm absorbance was measured by Ultra violet Spectrophotometer (U-2900 Hitachi Tokyo Japan). The mean standard deviation results for three consecutive measurements were expressed as RE (rutin equivalents) in mg/g of plant extract [4].

#### Determination of Condensed Tannins

The modified Vanillin assay was used to determine the condensed tannin in plant leaves extract. [10]. From the 10% sample obtained 250  $\mu$ L of the sample was blended with 1.5 ml of 4% Vanillin solution, 750  $\mu$ L of 12N HCl solution and 750  $\mu$ L of H<sub>2</sub>O. This reactant was reserved for twenty minutes in a dark cabinet and its absorbance was measured at 500 nm by UV Spectrophotometer (U-2900 Hitachi Tokyo Japan). For standard catechin was used and quantification was expressed as mg/g in terms of catechin

#### In Vitro Antioxidant Activities

##### Hydrogen peroxide scavenging activity

Two millimoles of H<sub>2</sub>O<sub>2</sub> and each extract/standard at different concentrations (50-250  $\mu$ g/mL) were mixed in ratio (1:0.6) and kept in incubator for 10 min at 25°C. A control solution using a phosphate buffer without H<sub>2</sub>O<sub>2</sub> was prepared. At 230 nm with UV Spectrophotometer (U-2900 Hitachi Tokyo Japan) absorbance was taken and calculated by the following formula [11].

Scavenging Activity%=

$$\frac{(\text{Control Absorbance} - \text{Sample Absorbance}) \times 100}{\text{Control Absorbance}}$$

##### Superoxide dismutase (SOD)-like activity

The 0.2 mL of extracts/standard (50-250  $\mu$ g/mL) was added to 0.2 mL of pyrogallol (7.2 mM), 2.6 mL (pH 8.5) of tris buffer (10 mM EDTA, 50 mM tris) and at 25°C for ten minutes kept in an incubator. Then 0.1mL HCl (1N) was added and at 420 nm absorbance was measured using U-2900 Hitachi spectrophotometer (Tokyo Japan). The following equation was used to calculate SOD-like activity [12].

%SOD like activity=

$$[1 - (\text{Extract absorbance} / \text{Control absorbance})] \times 100.$$

NO Radical Scavenging Potential

Each standard and extracts (50-250  $\mu$ g/mL) was prepared and from these diluted samples one milliliter solution was taken and blended with sodium nitroprusside one milliliter (10 mmol/L) reaction solution using 20 mmol/L phosphate buffer (pH=7.0). At 37 °C incubation of reaction mixture was carried for 1hrs and mixed 0.5 mL of Griess reagent with 0.5 mL of incubated mixture. At 540 nm absorbance was measured by U-2900 Hitachi UV Spectrophotometer (Tokyo Japan). NO radical scavenging potential was calculated as [13].

Scavenging Activity % =

$$\frac{(\text{Control Absorbance} - \text{Sample Absorbance}) \times 100}{\text{Control Absorbance}}$$

Analysis of Statistics:

All the measurements were noted in triplicate form and results were expressed in average  $\pm$  SD. A significant value considered as  $p < 0.05$ .

#### Results and Discussion

The bioactive compounds in *M. communis* leaves extracts are shown in table 1. Methanol extract showed the highest active components as compared to the other extracts.

It is considered that H<sub>2</sub>O<sub>2</sub> is not poisonous, but it is dangerous because of its capacity to make the hydroxyl ions, in this manner call attention to the significance of its removal. The H<sub>2</sub>O<sub>2</sub> antioxidant activity of *M. communis* leaves extracts shows in table 2. The dissimilarities of hydrogen peroxide scavenging potential between the extract may be because of structural properties of its energetic

locations, which decide the donating potential of its electron, solvent extraction procedures and percent values of extractions.

To find out whether the *M. communis* extracts have superoxide-scavenging factors, we measured the ability of leaves to scavenge superoxide radicals. The *M. communis* leaves extract SOD-like activity is shown in table 3. The SOD-like activity results was found very good, which increased slowly with a rise in concentration (50–250 µg/mL) as compared to standard. At 250µg/mL extract the methanol and vitamin C were found the same SOD-like activity (85%). These studies support that the myrtle leaves extracts have active compounds scavenging superoxide radicals, dominantly present in the extract prepared in methanol.

Nitric oxide scavenging activity of various extracts of *M. communis* L. is illustrated in table 4. Increase in the leaves concentration shows increase in antioxidant activities. The results showed that 250 µg/mL vitamin C and methanol extract had the highest scavenging activity of 75±1.

Hydrogen peroxide has a potential to directly deactivate some enzymes, although H<sub>2</sub>O<sub>2</sub> is a weak oxidizing molecule [14]. Hydrogen peroxide is converted into •OH which have poisonous effects, due to the occurrence of redox active iron and copper active transition metals [11]. So it is necessary to monitor the H<sub>2</sub>O<sub>2</sub> amount accumulated in the tissue. *M. communis* scavenged H<sub>2</sub>O<sub>2</sub> at a considerable rate in comparison to vitamin C, because myrtle leaves contained high amount of flavonoids.

A powerful intermediary in many physiological phenomena like regulation of cell mediated toxicity, inhibition of platelet aggregation, neuronal signaling and smooth muscle relaxation is nitric oxide. Like a diffusible reactive oxygen species NO have many activities, like an effectors agent in numerous life processes including antitumor, antimicrobial, vasodilatation and neuronal messenger activities [15].

Antioxidant defense mechanisms in cells against oxidative damage include various enzymatic and non-enzymatic antioxidants. SOD is a primary enzyme in enzymatic antioxidant defenses. Removal of superoxide anion radicals is considered important

because they serve as precursors of other activated oxygen including H<sub>2</sub>O<sub>2</sub> and •OH [12]. Therefore, the presence of factors having strong SOD-like activity in *M. communis* could effectively eliminate the toxic effects of ROS. Due to the reactivity of phenol moiety, phenol compounds act as antioxidants [16]. Dissimilarities in the total phenol content observed in extracts of different solvents appear to be due to the difference in solubility of the phenolic compounds [17]. It was previously reported that antioxidant activities, nutritional, color, sensory and color qualities of foods have been related with phenolic acids [18].

### Conclusion

The present study concluded that leaves of *M. communis* extract have high polyphenol concentration and have good antioxidant properties. Their curative potential against a large number of diseases can be used effectively, especially those related with oxidative stress. The data obtained through this study revealed that all the leaves extracts are likely to have the effect of scavenging free radicals and can be used in pharmaceuticals, food and cosmetics for healthy skin and/or anti-ageing products.

### Acknowledgment:

We (authors) are very thankful to Mr. Faqir Hussain (Senior Technician) of Food Technology Center of PCSIR Laboratories Complex Peshawar-Pakistan for assistance during sample collection.

### Conflict in Interests

We (authors) have no competitive interest.

### References

1. Gamze, Y., Egemen, D., Sukran, D. 2014. Comparison of the Antioxidative Components of some Marine Macroalgae from Turkey. *Pak J Bot.* 46(2): 753-757.
2. Anelise, SNF., Carla, RFV., Matheus, S., Claudia, ALC., Maria, CV., Zefa, VP. 2014. Evaluation of Antioxidant Activity, Total Flavonoids, Tannins and Phenolic Compounds in Psychotria Leaf Extracts. *Antioxidants.* 3: 745-757.
3. Camelia, P., Cristiana, D., Nicorescu, V. 2008. Antioxidant activity of sea buckthorn (*Hippophae*

Rhamnoides) extracts compared with common food additives. *Rom Biotech Lett.* 13(6): 4049-4053.

4. Amensoura M., Sendrab E., Abrinia J., Perez-Alvarez JA., and Fernandez-Lopez J. Antioxidant activity and total phenolic compounds of myrtle extracts. *CyTA – Journal of Food.* Vol. 8, No. 2, August 2010, 95–101.

5. Moure, A., Cruz, J.M., Franco, D., Dominguez, J.M., Sineiro, J., Dominguez, H., et al. (2001). Natural antioxidants from residual sources. *Food Chemistry*, 72, 145–171.

6. Dai, J. and Mumper RJ. 2010. Plant phenolics: extraction, analysis, and their antioxidant and anticancer properties. *Molecules*, 15, 7313-7352.

7. Sroka, Z., and Cisowski, W. 2003. Hydrogen peroxide scavenging, antioxidant and anti-radical activity of some phenolic acids. *Food and Chemical Toxicology*, 41, 753–758.

8. Olga G., Stavros Lalas • Ioanna Chinou • John Tsaknis. Reevaluation of bioactivity and antioxidant activity of *Myrtus communis* extract before and after encapsulation in liposomes. *Eur Food Res Technol.* DOI 10.1007/s00217-007-0592-1.

9. Tuba M, Tuğçe F, Bijen K, Tansel O. 2008. Antimicrobial and Cytotoxic Activities of *Myrtus communis* L. *J. Fac. Pharm, Ankara.* 37 (3) 191 - 199.

10. Arvind, KG., Tanmayee, M., Malay, B.A., Pallab, K., Arnab, S. 2013. Evaluation of phytochemical constituents and antioxidant activity of selected actinorhizal fruits growing in the forests of Northeast India. *J Biosci.*, 38(4): 797–803.

11. Price, ML. and Scoyoc, SV and Butler, LG. 1978. A critical evaluation of the vanillin reaction as

an assay for tannin in sorghum grain. *Journal of Agricultural and Food Chemistry*, 26 (5): 1214-1218.

12. Jong, HJ., Ji, WL., Kyoung, SK., Ju-Sung, K., Sang, NH., Chang, YY., Ju, KL., Yong, SK., Myong, JK. 2010. Antioxidant and antimicrobial activities of extracts from a medicinal plant, Sea Buckthorn. *J Korean Soc Appl Biol Chem.*, 53(1): 33-38.

13. Otakar, R., Sezai, E., Jiri, M., Tunde, J., Ignac, H. 2014. Antioxidant and radical scavenging activities in fruits of 6 sea buckthorn (*Hippophae rhamnoides* L.) cultivars. *Turk J Agric For.*, 38: 224-232.

14. Hazra, B., Sarkar, R., Biswas, S., Mandal, N. 2010. Comparative study of the antioxidant and reactive oxygen species scavenging properties in the extracts of the fruits of *Terminalia chebula*, *Terminalia bellerica* and *Emblica officinalis*. *BMC Complement Altern Med.*, 10: 20-26.

15. Acharya, S., Sahu, A.R., Satyajit, MS. 2010. Free radical scavenging activity of thalamus of *Nymphacea stellata* Willd. *Int J Pharmacy Pharm Sci.*, 2: 61–63.

16. Shahidi, F., Wanasundara, PK. 1992. Phenolic Antioxidants. *Crit Rev Food sci Nutr.*, 32: 67- 72.

17. Singh, AK., Attrey, D.P., Naveed, T. 2013. Bioactivity guided extraction of Seabuckthorn (*Hippophae rhamnoides* L. ssp. *Turkestanica*) leaves. *J Sci Ind Res.*, 72: 307-311.

18. Shivani, C., Chandresh, V. 2012. The Profile of Bioactive Compounds in Seabuckthorn: Berries and Seed oil Hydroxyl radical scavenging activity. *Int J Theor Appl Sci.*, 4(2):216-220.

**Table 1.** Table 1 Bioactive Components in *M. communis* Leaves Extracts.

Extracts	Total phenolic content	Total flavonoids	Condensed Tannins
n-Hexane	85±02	5.5±0.7	12±0.5
Methanol	120±03	6.5±01	14±01
Aqueous	95±02	5.2±0.5	10±0.5
Chloroform	80±0.5	4.5±0.4	07±0.3

Results are average of three replicates ± SD. Phenolic quantity (gallic acid (mg) equivalents/g of sample). Flavonoids quantification (Rutin (mg) equivalents/g of sample), Condensed tannin contents (as catechin equivalents in mg CE/g)

**Table 2** H<sub>2</sub>O<sub>2</sub> Scavenging Activity (%) of *M. communis* Leaves Extracts.

Extracts	Concentrations (µg/mL)				
	50	100	150	200	250
n-Hexane	28±0.5	48±0.1	54±0.4	68±1.2	80±01
Methanol	30±0.3	52±0.5	62±0.3	70±1.3	84±01
Aqueous	28±0.2	48±0.2	62±0.2	65±0.2	78±02
Chloroform	18±0.1	32±0.3	45±0.2	52±1.1	62±01
Vitamin C	35±0.6	60±0.5	70±0.5	78±0.5	85±02

Data are average of three replicate ± SD.

**Table 3** Percent inhibition rate of SOD-like activity of *M. communis* Leaves Extracts.

Extracts	Concentrations (µg/mL)				
	50	100	150	200	250
n-Hexane	20±0.5	52±1	62±0.6	70±0.4	75±0.2
Methanol	22±1.4	62±1	70±1.5	78±0.1	85±0.5
Aqueous	15±0.8	40±2	58±0.3	68±0.6	74±0.4
Chloroform	12±0.1	25±1	34±0.5	52±0.2	60±00
Vitamin C	25±0.3	60±2	70±1.2	80±0.6	85±0.7

Data are average of three replicate ± SD.

**Table 4** NO Scavenging Potential of *M. communis* Leaves Extracts.

Extracts	Concentrations ( $\mu\text{g/mL}$ )				
	50	100	150	200	250
<b>n-Hexane</b>	20 $\pm$ 0.2	40 $\pm$ 0.5	50 $\pm$ 1	70 $\pm$ 0.1	72 $\pm$ 0.9
<b>Methanol</b>	25 $\pm$ 01	54 $\pm$ 0.5	64 $\pm$ 1	68 $\pm$ 01	75 $\pm$ 01
<b>Aqueous</b>	10 $\pm$ 0.7	35 $\pm$ 0.1	48 $\pm$ 0.7	65 $\pm$ 0.4	68 $\pm$ 0.2
<b>Chloroform</b>	12 $\pm$ 1.5	28 $\pm$ 0.2	38 $\pm$ 0.8	44 $\pm$ 0.2	65 $\pm$ 0.1
<b>Vitamin C</b>	25 $\pm$ 0.5	48 $\pm$ 0.9	65 $\pm$ 0.1	70 $\pm$ 0.5	75 $\pm$ 0.8

Data are average of three replicate  $\pm$  SD.