

**A PHARMACOLOGICAL REVIEW ON THE TWO TRADITIONAL PLANTS USED IN BANGLADESH  
FOR JAUNDICE**

Rahman Bhuiyan, M. M.<sup>1</sup>; Jannatun Nahar, U.<sup>2\*</sup>

<sup>1</sup> University of Development Alternative, 80, Satmosjid Road, Dhanmondi, Dhaka - 1209, Bangladesh

<sup>2</sup> Jagannath University, 9-10, Chittaranjan Ave, Dhaka-1100, Bangladesh

[\\*nahar.jannatun58@gmail.com](mailto:nahar.jannatun58@gmail.com)

**Abstract**

Since the inception of the universe, nature has been enriched with abundant resources are being given away for the betterment of humanity. Nature has the plethora of biologically active compounds those are potential to prevent ailments. By virtue of natural favor, Bangladesh has been a domain with plenty of natural plants possessing great therapeutic significance. Over time myriad medicinal plants have been identified and used in Bangladesh to treat various types of diseases. Hepatic diseases are progressively increasing in Bangladesh and it has been a challenge to achieve expected compounds that elicit effective potential against these complications. Various traditional plants have been used in treating hepatic diseases and jaundice since ancient period. The current review has focused largely on the manifold aspects of pharmacological actions of *Asparagus racemosus* and *Terminalia chebula*, two indigenous traditional plants used in Bangladesh for the treatment of jaundice. Alongside it has also demonstrated the current traditional formulation of these plants used for the treatment of jaundice.

**Keywords:** *Asparagus racemosus*, *Terminalia chebula*, jaundice, pharmacological activities, Bangladesh.

## Introduction

Nature is considered as eternal sources for various medicinal agents and drug development. There are about 250000 species of plants in the universe among them 6% have been found to be biologically active and 15% have undergone phytochemical testing [1]. It is estimated that about 80% out of more than 4000 million people of the world depend on the traditional medicines to meet the demand for primary health care needs [2]. Whereas 70-80% people of developing Asian countries rely on the traditional medicine despite the availability of allopathic medicines in many areas of the community [3]. The utilization of plant-based medicines is known as ethnomedicine. The aim of ethnopharmacology is to facilitate the discovery of the bioactive compounds from natural sources [4]. Plants remain an excellent source of new drugs, and new chemical entities. Medicinal plants serve for the development of both natural products and their derivatives. For example anti-malarial drug arteetheris derived from artemisinin which is isolated from the traditional plant, *Artemisia annua* L. (Asteraceae). There are few more derivatives of artemisinin under clinical trials in Europe to be used as anti-malarial drugs [5]. Evidence shows that there was an involvement of the use of various traditional medicines in the early health care systems of developing countries [3]. The rapid increase of the knowledge regarding the optimistic effects of the plant-derived compounds accelerates using these agents to cure diseases [6].

Jaundice is not a disease but it is a sign that may result from conditions like viral hepatitis, gallstones, hemolytic anemia, pancreatic cancer, parasite infection of the liver, autoimmune hepatitis, or use of certain drugs etc. In recent days jaundice is very common in Bangladesh because of the rapid increase in the various hepatic diseases [6, 7]. There is no effective treatment for jaundice and hepatitis to be covered by current allopathic and homeopathic treatment procedure [6].

In this review article, the authors have tried to put together information of two exclusively used plants with their activity against jaundice along with other investigated pharmacological effects.

## Methods

*Asparagus racemosus* and *Terminalia chebula* were selected from searching literature, and then their inclusion was confirmed in the Ethnobotanical Database of Bangladesh (EDB), which ensures their indigenous identity in Bangladesh. Most of the information required to prepare the manuscript were collected by the comprehensive searching of literature. Google scholar was the preliminary tool for searching and identifying appropriate published works. Then these articles were accumulated from various reputed electronic sites like Elsevier, ScienceDirect, Springer, PubMed, PMC, Wiley Online Library, Hindawi, and Taylor & Francis etc.

## Traditional uses for the treatment of jaundice

Exploration of literature claimed that different parts of the plants are used for the treatment of jaundice and these parts of the plants include roots and fruits. In most of the cases, root of *A. racemosus* are taken as juice and fruits of *T. Chebula* are taken orally [8-11]. The common forms of use of these two selected plants are given in the table 1.

Table 1: Mode of use of reviewed plants in jaundice

### Pharmacological activities

#### 4.1. Pharmacological activities of *A. racemosus*

##### 4.1.1. Antitussive effect

Study reported that methanol roots extract of *A. racemosus* (AR) given at the dose of 200 and 400 mg/kg, orally showed the antitussive effects on SO<sub>2</sub> induced cough in mice model. The dose 200 and 400 mg/kg showed 40% and 58.5% cough inhibition respectively those were comparable to that of 10 and 20 mg/kg of codeine phosphate that resulted in 36% and 55.4% inhibition respectively when used as standard [12].

##### 4.1.2. Antiulcer activity

Crude extract of *A. racemosus* was administered at the dose of 100 mg/kg body weight, treatment improved condition of indomethacin plus pyloric ligation (PL)-induced gastric ulcer in rat models. The gastric ulcer reduction was also comparable to the standard antiulcer drug Ranitidine given at the dose of 30 mg/kg/day [13]. Methanol extract of *A.*

*racemosus* dose-dependently protected gastric ulcers when administered to test animals at the dose of 25–100 mg/kg twice daily for 5 days [14].

#### 4.1.3. Adaptogenic activity

Standardized methanol extract of *A. racemosus* (AR) was applied at various doses (50, 100, and 200 mg/kg, BW orally) to test animals for assessing physiological modulation of the stress pathways. Methanol extract of AR (MAR) decreased the plasma corticosterone and norepinephrine levels in a dose-dependent manner. Tissue level of monoamines (serotonin, dopamine, and norepinephrine) was increased in hypothalamus by MAR. The dose 100 mg/kg was found to show its ceiling effect at which it produced its maximum effect [15].

#### 4.1.4. Anticancer activity

The anticancer activity of the extract of *A. racemosus* was evaluated using HT-29 (human colon adenocarcinoma), MCF-7 (human breast cancer), and A-498 (human kidney cancer) cell lines. Oral administration of the test material at the doses of 250 and 500 mg/kg body weight caused significant reduction in tumor volume, packed cell volume, percent increase in body weight, viable tumor cell count, and increase in non-viable cell count compared to control group [16].

#### 4.1.5. Antidepressant activity

Study reported that methanol extract of *A. racemosus* (MAR) showed antidepressant activity when administered at the dose of 100, 200 and 400 mg/kg daily for 7 to test animals. The study results showed that MAR decreased immobility in forced swim test and increased avoidance response in learned helplessness test that could be interpreted as antidepressant activity [17].

#### 4.1.6. Antihypercholesterolemic activity

Study conducted on male albino rats suggested that the root powder of *A. racemosus* when provided with diet to hypercholesteremic rats showed reduction in both plasma and hepatic lipid profiles. Treatment also increased excretion of cholesterol in feces together with increased hepatic HMG-CoA reductase activity and bile acid content [18].

#### 4.1.7. Antiosteoporotic activity

*A. racemosus* revealed significant ( $P < 0.05$ ) impact on various processes of bone viz. ossification, mineralization, and suppression of ovariectomized rats. Treatment of ovariectomized rats with root extracts of *A. racemosus* caused reduction in serum alkaline phosphatase and serum calcium accompanied by inhibition of excessive loss of calcium in urine. In addition, extracts of *A. racemosus* improved hardness of 4th lumbar vertebra, length of femoral bone and its weight [19].

#### 4.1.8. Antioxidant activity

Crude and purified aqueous fraction of *A. racemosus* were investigated to assay its antioxidant activity against membrane damage of hepatic mitochondria of rats caused by the free radical generation during exposure to  $\gamma$ -radiation. Lyophilized polysaccharides (stated as P3) was found to be effective even at the concentration of 10  $\mu$ g/ml. Both the P3 fraction and the crude extract resulted in significant inhibition of protein oxidation and lipid peroxidation. The antioxidant effect of P3 fraction was more effective against lipid peroxidation, while the crude extract was measured more effective in inhibiting protein oxidation [20]. Methanol root extracts obtained from the *A. racemosus* significantly increased superoxide dismutase, ascorbic acid and catalase, whereas it decreased lipid peroxidase product in animal models [21].

#### 4.1.9. Enzyme inhibitory activity

In a study, methanol root extract of *A. racemosus* was administered at a dose of 50, 100 and 200 mg/kg orally, treatment significantly shortened the immobility periods in a dose dependent manner that can be measured by both tail suspension test and forced swim test. It was suggested that this might result from the reduction of Monoamine oxidase-A (MAO-A) and Monoamine oxidase-B (MAO-B) in brain after the administration of methanol extract of *A. racemosus* [22]. Another study reported that liposomal formulations of *A. racemosus* root extract (AR1-6) showed in vitro tyrosinase inhibitory activity [23].

#### 4.1.10. Hepatoprotective effect

Ethanol extract obtained from the roots of *A. racemosus* was co-administered orally with isoniazid at the dose of 50 mg/kg BW which showed hepatoprotective effects through free radical scavenging. Reduction of free radical generation resulted from the inhibition of hepatic enzyme (CYP2E1) activity and removal of free radical occurs due to induction of antioxidant enzymes and improvement of non-enzymatic thiol antioxidant glutathione [24]. Study conducted on Wistar rats revealed that pretreatment with the aqueous extract obtained from the roots of *A. racemosus* showed potential to prevent hepatocarcinogenesis caused by Diethylnitrosamine [25].

#### 4.1.11. Hypoglycemic activity

Ethanol extract of *A. racemosus* was administered to type 2 diabetic rats twice daily for 28 days. It significantly reduced the serum glucose levels and increased the level of serum insulin by 30% when compared with control. Moreover, the extract significantly decreased postprandial glucose level after ingestion [26].

#### 4.1.12. Immunomodulatory activity

One study reported that a unique polyhydroxylated steroidal sapogenin acid isolated from the roots of *A. racemosus* could booster the immune system of experimental animals with cyclosporine-A induced immune suppression [27]. A study postulated that treatment on experimental animal with (100 mg/kg b.w. p.o.) aqueous roots extract of *A. racemosus* revealed considerable increase in CD4/CD8+ and CD3+ counts that indicated activation of T cell. Animals model showed significant increase in Th1 (IL-2, IFN-g) and Th2 (IL-4) cytokines which suggested its mixed Th1/Th2 adjuvant potential [28].

#### 4.1.13. Neuroprotective effects

This study was performed using five groups of adult female Wistar rats where three ovariectomized (OVX) groups received ethanol root extract of *A. racemosus* at the dose of 100 mg/kg BW, 1000 mg/kg BW and 17 $\alpha$ -ethynylestradiol (EE) at the dose of 0.1 mg/kg BW during experimental period. The rat groups treated with ethanol root extract and 17 $\alpha$ -ethynylestradiol manifested significantly upper recognition index than that of

OVX group. It is suggested that the root extracts and EE reduced the neuronal loss in the brain and thereby improved cognitive function of ovariectomized rats [29]. Methanol root extract of *A. racemosus* was used against kainic acid (KA) induced hippocampal and striatal neuronal damage in female Swiss albino mice. Administration of kainic acid (KA) caused an increase in protein carbonyl content and lipid peroxidation and decrease in glutathione peroxidase (GPx) activity and reduced glutathione (GSH) content. The mice model treated with *A. racemosus* root extract displayed an improvement in GPx activity and GSH content but reduction in protein carbonyl and membranal lipid peroxidation [30].

#### 4.1.14. Prokinetic activity

Study showed that *A. racemosus* (Shatavari) and metoclopramide significantly reduced gastric emptying half-time (GE t<sub>1/2</sub>). The basal GE t<sub>1/2</sub> in volunteers was 159.9  $\pm$  45.9 min which was reduced to 101  $\pm$  40.8 min by Shatavari (P<0.001) and to 85.3  $\pm$  21.9 by metoclopramide (P<0.001). Both of the agent did not differ significantly in their effects [31].

#### 4.1.15. Teratogenic activity

When methanol extract of *A. racemosus* given at the dose of 100 mg/kg for 60 days to test animals showed teratological changes including increased resorption of fetuses, gross malformations including swelling in legs and intrauterine growth retardation with reduced placental size [32].

#### 4.1.16. Miscellaneous effects

Searching of various literatures revealed that various forms of extracts of *A. racemosus* showed distinctive pharmacological activities. Methanol extract of *A. racemosus* root showed antiuremic activity and anti-amnesic activity, ethanol extract of roots showed anti-urolithiatic activity, ethanol and aqueous extracts of *A. racemosus* root showed anti-diarrheal activity, aqueous solution of the crude alcoholic extract of the roots exhibited antiprotozoal activity, alcohol extract of the roots of *A. racemosus* produced positive inotropic and chronotropic effect, leaf extracts of *A. racemosus* were responsible for its antimicrobial effects, hydro-alcoholic extracts of roots of *A. racemosus* showed



meaningful aphrodisiac activity, aqueous roots extract of *A. racemosus* exhibited immunoadjuvant and antidiuretic activity, study conducted using incision and excision model revealed wound healing activity of *A. racemosus*, root extract of *A. racemosus* has been prescribed to augment the secretion of milk of lactating mother, lyophilized extracts of *A. racemosus* were found to be beneficial to improve fertility, alcoholic extract of rhizome of *A. racemosus* exhibited oosterogenic effects, ethanol leaf extract of *A. racemosus* showed anti-inflammatory effect, asparagine A of *A. racemosus* showed anti-abortifacient effect [33-47].

## 5.2. Pharmacological activities of *Terminalia chebula*

### 5.2.1. Adaptogenic activities

The ethanol extracts of *T. chebula* showed anti-stress activity by preventing stress-induced increased levels of various biochemicals like triglycerides, cholesterol, blood urea nitrogen (BUN), glucose plasma corticosterone. Treatment also reduced blood cell count (RBC and WBC) and prevented alteration of weights of organs like liver, spleen, testis, and adrenal glands etc. [48].

### 5.2.2. Antibacterial activity

Various extracts of *T. chebula* showed varying degree of antibacterial potential against specific bacterial strain i.e. ethanol extract was superior against *E. coli* and hot aqueous extract showed superiority against *S. aureus*. Whereas cold aqueous extract exhibited the least antibacterial activity against all the tested strains [49]. Study reported that ethanol fruit extract of *T. chebula* was highly effective against *Salmonella typhi* SSFP 4S, *Staphylococcus epidermidis* MTCC 3615, *Staphylococcus aureus* ATCC 25923, *Bacillus subtilis* MTCC 441 and *Pseudomonas aeruginosa* ATCC 27853 [50].

### 5.2.3. Anticlastogenic activity

The anticlastogenic activity of *T. chebula* extract has been reported against micronuclei formation and chromosomal aberration induced by cyclophosphamide in mice bone marrow cells. It dose dependently and significantly ( $P < 0.05$ ) inhibited micronuclei formation and chromosomal

aberration when compared with the standard cyclophosphamide group [51].

### 5.2.4. Antidiabetic activities

Ethyl Acetate-Soluble portion of ethanol extract of *T. chebula* fruit (EETC) improved streptozotocin induced diabetic mellitus in rats. High dose (500 mg/kg BW) of extract (HEETC) decreased the blood glucose and serum lipid levels, treatment also reduced serum and thoracic aorta concentrations of malondialdehyde in diabetic rats [52]. It was revealed that the oral administration of ethanol extract of the fruits at the dose of 200 mg/kg BW for 30 days significantly reduced levels of blood glucose and glycosylated hemoglobin in streptozotocin induced diabetic rats [53].

### 5.2.5. Anti-inflammatory activity

In vivo anti-inflammatory activity of *T. chebula* fruit was evaluated at different doses ranged from 50 to 500 mg/kg, p.o. against carrageenan-induced inflammation in rats. Study revealed that standardized extract at the dose of 250 mg/kg, p.o. caused 69.96% reduction in carrageenin-induced paw edema in rats [54].

### 5.2.6. Antioxidant activity

The polyphenolic extract of *T. chebula* fruits has been demonstrated to have antioxidant activity. It was authenticated by determining DPPH radical concentration ( $IC_{50}$  14  $\mu$ g/mL), the reducing power, total antioxidant capacity, nitric oxide radical concentration ( $IC_{50}$  30.51  $\mu$ g/mL) and hydrogen peroxide scavenging activity ( $IC_{50}$  265.53  $\mu$ g/mL) under in vitro conditions [55]. Study showed that aqueous extract of fruit of *T. chebula* (TCE) significantly reversed the t-BHP-induced cell cytotoxicity and lactate dehydrogenase leakage in rat models. TCE also exhibited in vitro ferric-reducing antioxidant activity and 2,2-diphenyl-1-picrylhydrazyl free radical-scavenging activities [56].

### 5.2.7. Anti-platelet activity

*T. chebula* fruit extract showed dose dependent inhibition of arachidonic acid induced human platelet aggregation. More than 30% inhibition was observed at 5 mg/ml dose while maximum inhibition of 95% was obtained at the dose of 10 mg/ml ( $IC_{50}$ : 6.74 mg/ml) [57].

#### 5.2.8. Antiulcer activity

Study demonstrated that pretreatment with hydroalcoholic extract of *Terminalia chebula* fruit at the doses of 200 and 500 mg/kg caused significant reduction in lesion index, total affected area and percentage of lesion in comparison with control group ( $P < 0.05$  and  $P < 0.01$ ) in the aspirin, ethanol and cold restraint stress-induced ulcer in rat models [58].

#### 5.2.9. Enzyme inhibitory activity

Acetylcholinesterase inhibitors are employed for the symptomatic treatment of Alzheimer's disease. Methanol extracts of *T. chebula* of ayurvedic herbal drug 'Triphala' showed acetylcholinesterase inhibitory activity. Gallic acid and ellagic acid, and the phenolic acids present in the fruits inhibited the enzyme acetylcholinesterase [60]. Another study reported to isolate 1,2,3,4,6-penta-O-galloyl- $\beta$ -D-glucose, a compound from *T. chebula* showed significant acetylcholinesterase (AChE) and butyrylcholinesterase (BChE) inhibitory activities with IC<sub>50</sub> values of  $29.9 \pm 0.3 \mu\text{M}$  and  $27.6 \pm 0.2 \mu\text{M}$ , respectively [61].

#### 5.2.10. Hepatoprotective effects

*T. chebula* water extract (TCW) ameliorated tert-butylhydroperoxide-(t-BHP-) induced acute liver injury in C57/BL6 mice model. Tert-butylhydroperoxide-(t-BHP-) dramatically elevated the serum AST, ALT, and LDH level, while animals pretreated with TCW showed significant reduction of these biomarkers [62].

#### 5.2.11. Hypocholesterolemic activity

Study reported Haritaki that is *Terminalia chebula*, when administered to atherogenic diet induced hyperlipidemic rat models, treatment caused reduction in total cholesterol, triglycerides, total protein and elevation of high density lipoprotein cholesterol. It was also suggested that Haritaki at concentrations of 1.05 and 2.10 mg/kg BW worked as an excellent lipid-lowering agent [63].

#### 5.2.12. Immunosuppressive activity

Study reported that gallic acid (GA) and chebulagic acid (CA) extracted from the fruits of the *T. chebula* revealed immunosuppressive activity.

They blocked the cytotoxic T lymphocyte mediated (CTL) mediated cytotoxicity. GA and CA were found to inhibit the activity of CD8+ CTL clone at IC<sub>50</sub> values of  $30 \mu\text{M}$  and  $50 \mu\text{M}$ , respectively [64].

#### 5.2.13. Neuroprotective effect

In vitro neuroprotective effect of various extracts (water, methanol, and 95% ethanol) of the *T. chebula* was evaluated by the inhibition of H<sub>2</sub>O<sub>2</sub>-induced rat pheochromocytoma cell (PC12) death. Application of H<sub>2</sub>O<sub>2</sub> induced a dose-dependent loss in viability of PC12 cells. Among the three extracts water extract showed the greatest neuroprotective activity [65].

#### 5.2.14. Miscellaneous effects

Oil fraction from *T. Chebula* possessed purgative activity, polyhedral formulation of *T. chebula* exhibited antiallergic activity, combination of *T. chebula* with other botanicals showed antiprotozoal activity, acetone extract of *T. chebula* showed antimutagenic activity, ethyl acetate, methanol and acetone extracts of *T. chebula* revealed anthelmintic activity, *T. chebula* extract showed prokinetic activity, methanol extract of *T. chebula* fruit showed anticancer activity, ethyl acetate and ether extract of fruits of *T. chebula* were found to be active against pathogenic fungal strains, ripe *T. chebula* fruits expressed Immunomodulatory activities, alcohol extract of *T. chebula* manifested cardioprotective effect, aqueous extract of *T. chebula* showed renoprotective, antiurolithiatic, anticaries and radioprotective activity, various organic and aqueous extracts of *T. chebula* showed wound healing activities, compounds of *T. chebula* showed activity against hepatitis C virus, water soluble fraction of *T. chebula* exhibited antianaphylactic effect, acetone extract of *T. chebula* fruits possessed antiarthritic activity, ethanol extract of *T. chebula* showed anticonvulsant activity [66-81].

### Discussion

Literature review revealed that both of the plants are used for the treatment of jaundice as folk medicine in Bangladesh. Besides this review also manifested many distinctive pharmacological activities of these plants. It is important to rationale and prioritize the promising pharmacological effects with a view to fitting contemporary treatment need.

This review is likely to provide new glimpse to the researcher of relevant fields to find out any lead candidate or its derivatives for the effective treatment of Jaundice or any other ailments which are postulated by the pharmacological activities of these medicinal plants.

### Acknowledgments

Authors are thankful to the Department of Pharmacy, University of Development Alternative for giving technical support.

### References

1. Verpoorte R. Pharmacognosy in the new millennium: leadfinding and biotechnology. *Journal of pharmacy and pharmacology*. 2000 Mar;52(3):253-62.
2. Farnsworth NR, Akerele O, Bingel AS, Soejarto DD, Guo Z. Medicinal plants in therapy. *Bulletin of the world health organization*. 1985;63(6):965.
3. Sheng-Ji P. Ethnobotanical approaches of traditional medicine studies: some experiences from Asia. *Pharmaceutical biology*. 2001 Jan 1;39(sup1):74-9.
4. Fabricant DS, Farnsworth NR. The value of plants used in traditional medicine for drug discovery. *Environmental health perspectives*. 2001 Mar;109(Suppl 1):69.
5. Balunas MJ, Kinghorn AD. Drug discovery from medicinal plants. *Life sciences*. 2005 Dec 22;78(5):431-41.
6. Abbasi AM, Khan MA, Ahmad M, Zafar M, Khan H, Muhammad N, Sultana S. Medicinal plants used for the treatment of jaundice and hepatitis based on socio-economic documentation. *African Journal of Biotechnology*. 2009;8(8).
7. Chowdhury AR, Rahmatullah M. Ethnomedicinal plants for treatment of jaundice by the folk and tribal medicinal practitioners of several districts in Bangladesh and review of their scientifically reported hepatoprotective activity. *American-Eurasian Journal of Sustainable Agriculture*. 2012 Oct 1:360-71.
8. Rahman AH, Anisuzzaman M, Haider SA, Ahmed F, Islam AK, Naderuzzaman AT. Study of medicinal plants in the Graveyards of Rajshahi city.

*Research Journal of Agriculture and Biological Sciences*. 2008;4(1):70-4.

9. Rahmatullah M, Mollik MA, Harun-or-Rashid M, Tanzin R, Ghosh KC, Rahman H, Alam J, Faruque MO, Hasan MM, Jahan R, Khatun MA. A comparative analysis of medicinal plants used by folk medicinal healers in villages adjoining the Ghaghot, Bangali and Padma Rivers of Bangladesh. *American-Eurasian Journal of Sustainable Agriculture*. 2010 Jan 1:70-86.

10. Rahman AM, Sultana N, Islam AR, Zaman AT. Study of Medical Ethno-botany at the Village Genda under Savar Upazilla of District Dhaka, Bangladesh. *Journal of Medicinal Plants*. 2013;1(5).

11. Rahmatullah M, Mollik MA, Islam MK, Islam MR, Jahan FI, Khatun Z, Seraj S, Chowdhury MH, Islam F, Miajee ZU, Jahan R. A survey of medicinal and functional food plants used by the folk medicinal practitioners of three villages in Sreepur Upazilla, Magura district, Bangladesh. *American Eurasian Journal of Sustainable Agriculture*. 2010 Sep 1;4(3):363-73.

12. Mandal SC, CK AK, Lakshmi SM, Sinha S, Murugesan T, Saha BP, Pal M. Antitussive effect of *Asparagus racemosus* root against sulfur dioxide-induced cough in mice. *Fitoterapia*. 2000 Dec 1;71(6):686-9.

13. Bhatnagar M, Sisodia SS. Antisecretory and antiulcer activity of *Asparagus racemosus* Willd. against indomethacin plus pyloric ligation-induced gastric ulcer in rats. *Journal of herbal pharmacotherapy*. 2006 Jan 1;6(1):13-20.

14. Sairam K, Priyambada S, Aryya NC, Goel RK. Gastroduodenal ulcer protective activity of *Asparagus racemosus*: an experimental, biochemical and histological study. *Journal of Ethnopharmacology*. 2003 May 1;86(1):1-0.

15. Krishnamurthy S, Garabadu D, Ranga Reddy N. *Asparagus racemosus* modulates the hypothalamic-pituitary-adrenal axis and brain monoaminergic systems in rats. *Nutritional neuroscience*. 2013 Nov 1;16(6):255-61.

16. Mitra SK, Prakash NS, Sundaram R. Shatavarins (containing Shatavarin IV) with anticancer activity from the roots of *Asparagus racemosus*. *Indian journal of pharmacology*. 2012 Nov;44(6):732.

17. Singh GK, Garabadu D, Muruganandam AV, Joshi VK, Krishnamurthy S. Antidepressant activity

of *Asparagus racemosus* in rodent models. *Pharmacology Biochemistry and Behavior*. 2009 Jan 1;91(3):283-90.

18. Visavadiya NP, Narasimhacharya AV. *Asparagus* root regulates cholesterol metabolism and improves antioxidant status in hypercholesteremic rats. *Evidence-Based Complementary and Alternative Medicine*. 2009;6(2):219-26.

19. Chitme HR, Muchandi IS, Burli SC. Effect of *Asparagus racemosus* Willd root extract on ovariectomized rats. *The Open Natural Products Journal*. 2009;2(1):16-23.

20. Kamat JP, Bolor KK, Devasagayam TP, Venkatachalam SR. Antioxidant properties of *Asparagus racemosus* against damage induced by  $\gamma$ -radiation in rat liver mitochondria. *Journal of ethnopharmacology*. 2000 Aug 1;71(3):425-35.

21. Kongkanermit L, Witoonsaridsilp W, Peungvicha P, Ingkaninan K, Waranuch N, Sarisuta N. Antioxidant activity and antiapoptotic effect of *Asparagus racemosus* root extracts in human lung epithelial H460 cells. *Experimental and therapeutic medicine*. 2011 Jan 1;2(1):143-8.

22. Dhingra D, Kumar V. Pharmacological Evaluation for Antidepressant like Activity of *Asparagus racemosus* Willd. mice. *Pharmacologyonline*. 2007;3:133-52.

23. Therdphapiyanak N, Jaturanpinyo M, Waranuch N, Kongkanermit L, Sarisuta N. Development and assessment of tyrosinase inhibitory activity of liposomes of *Asparagus racemosus* extracts. *asian journal of pharmaceutical sciences*. 2013 Apr 1;8(2):134-42.

24. Palanisamy N, Manian S. Protective effects of *Asparagus racemosus* on oxidative damage in isoniazid-induced hepatotoxic rats: an in vivo study. *Toxicology and industrial health*. 2012 Apr;28(3):238-44.

25. Agrawal A, Sharma M, Rai SK, Singh B, Tiwari M, Chandra R. The effect of the aqueous extract of the roots of *Asparagus racemosus* on hepatocarcinogenesis initiated by diethylnitrosamine. *Phytotherapy research*. 2008 Sep;22(9):1175-82.

26. Hannan JM, Ali L, Khaleque J, Akhter M, Flatt PR, Abdel-Wahab YH. Antihyperglycaemic activity of *Asparagus racemosus* roots is partly mediated by inhibition of carbohydrate digestion and absorption,

and enhancement of cellular insulin action. *British Journal of Nutrition*. 2012 May;107(9):1316-23.

27. Sharma P, Chauhan PS, Dutt P, Amina M, Suri KA, Gupta BD, Suri OP, Dhar KL, Sharma D, Gupta V, Satti NK. A unique immuno-stimulant steroidal saponogenin acid from the roots of *Asparagus racemosus*. *Steroids*. 2011 Mar 31;76(4):358-64.

28. Gautam M, Saha S, Bani S, Kaul A, Mishra S, Patil D, Satti NK, Suri KA, Gairola S, Suresh K, Jadhav S. Immunomodulatory activity of *Asparagus racemosus* on systemic Th1/Th2 immunity: implications for immunoadjuvant potential. *Journal of ethnopharmacology*. 2009 Jan 21;121(2):241-7.

29. Lalert L, Kruevaisayawan H, Amatyakul P, Khongsombat O. Neuroprotective effects of the *Asparagus racemosus* root extract on ovariectomized rats. *J Physiol Biomed Sci*. 2013;26(1):18-22.

30. Parihar MS, Hemnani T. Experimental excitotoxicity provokes oxidative damage in mice brain and attenuation by extract of *Asparagus racemosus*. *Journal of Neural Transmission*. 2004 Jan 1;111(1):1-2.

31. Dalvi SS, Nadkarni PM, Gupta KC. Effect of *Asparagus racemosus* (Shatavari) on gastric emptying time in normal healthy volunteers. *Journal of Postgraduate Medicine*. 1990 Apr 1;36(2):91.

32. Goel RK, Prabha T, Kumar MM, Dorababu M, Singh G. Teratogenicity of *Asparagus racemosus* Willd. root, a herbal medicine. 2006; 44(7): 570–573.

33. Roy S, Das K, Mandal S, Pradhan S, Patra A, Nandi DK. Crude root extract of *Asparagus racemosus* ameliorates acetaminophen induced uremic rats. *International Journal of Pharmaceutical Sciences and Research*. 2013 Aug 1;4(8):3004.

34. Jagannath N, Chikkannasetty SS, Govindadas D, Devasankaraiah G. Study of antiurolithiatic activity of *Asparagus racemosus* on albino rats. *Indian journal of pharmacology*. 2012 Sep;44(5):576.

35. Hussain A, Ahmad MP, Wahab S, Hussain MS, Ali M. A review on pharmacological and phytochemical profile of *Asparagus racemosus* Willd. *Pharmacologyonline*. 2011;3:1353-64.

36. Alok S, Jain SK, Verma A, Kumar M, Mahor A, Sabharwal M. Plant profile, phytochemistry and pharmacology of *Asparagus racemosus* (Shatavari): A review. *Asian Pacific journal of tropical disease*. 2013 Jun;3(3):242.



37. Battu GR, Kumar BM. Phytochemical and Antimicrobial Activity of Leaf Extract of *Asparagus racemosus* Willd. *Pharmacognosy Journal*. 2010 Aug 1;2(12).
38. Wani JA, Achur RN, Nema RK. Phytochemical screening and aphrodisiac activity of *Asparagus racemosus*. *studies*. 2011;8(9):21-6.
39. Gautam M, Diwanay S, Gairola S, Shinde Y, Patki P, Patwardhan B. Immunoadjuvant potential of *Asparagus racemosus* aqueous extract in experimental system. *Journal of ethnopharmacology*. 2004 Apr 1;91(2-3):251-5.
40. Kumar S, Rajput R, Patil V, Udupa AL, Gupta S, Rathnakar UP, Rao S, Benegal D, Benegal A, Shubha HV. Wound healing profile of *Asparagus racemosus* (Liliaceae) wild. *Current Pharma Research*. 2011;1(2):111-4.
41. Goyal RK, Singh J, Lal H. *Asparagus racemosus*-An update. *Indian journal of medical sciences*. 2003 Sep 1;57(9):408-14.
42. Thakur M, Thompson D, Connellan P, Deseo MA, Morris C, Dixit VK. Improvement of penile erection, sperm count and seminal fructose levels in vivo and nitric oxide release in vitro by ayurvedic herbs. *Andrologia*. 2011 Aug;43(4):273-7.
43. Kumar MC, Udupa AL, Sammodavardhana K, Rathnakar UP, Shvetha U, Kodancha GP. Acute toxicity and diuretic studies of the roots of *Asparagus racemosus* Willd in rats. *West Indian Medical Journal*. 2010 Jan;59(1):03-6.
44. Pandey SK, Sahay A, Pandey RS, Tripathi YB. Effect of *Asparagus racemosus* rhizome (Shatavari) on mammary gland and genital organs of pregnant rat. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*. 2005 Aug;19(8):721-4.
45. Ojha R, Sahu AN, Muruganandam AV, Singh GK, Krishnamurthy S. *Asparagus racemosus* enhances memory and protects against amnesia in rodent models. *Brain and cognition*. 2010 Oct 1;74(1):1-9.
46. Battu GR, Kumar BM. Anti-inflammatory activity of leaf extract of *Asparagus racemosus* Willd. *International Journal of Chemical Sciences*. 2010;8(2):1329-38.
47. Sharma K and Bhatnagar M. *Asparagus racemosus* (Shatavari): a versatile female tonic. *Int J Pharm Biol Arch* 2011;2(3).
48. Debnath J, Prakash T, Karki R, Kotresha D, Sharma P. An experimental evaluation of anti-stress effects of *Terminalia chebula*. *J Physiol Biomed Sci*. 2011;24(2):13-9.
49. Bag A, Bhattacharyya SK, Pal BN, Chattopadhyay RR. Evaluation of antibacterial properties of Chebulic myrobalan (fruit of *Terminalia chebula* Retz.) extracts against methicillin resistant *Staphylococcus aureus* and trimethoprim-sulphamethoxazole resistant uropathogenic *Escherichia coli*. *African Journal of Plant Science*. 2009 Feb 28;3(2):025-9.
50. Kannan P, Ramadevi SR, Hopper W. Antibacterial activity of *Terminalia chebula* fruit extract. *African Journal of Microbiology Research*. 2009 Apr 30;3(4):180-4.
51. Raja W, Pandey S, Agrawal RC. Studies on the anticlastogenic effect of *Terminalia chebula* extract on cyclophosphamide-induced micronucleus formation and chromosome aberrations in Swiss albino mice. *Int J Gen*. 2011;1(2):13-7.
52. Kim JH, Hong CO, Koo YC, Kim SJ, Lee KW. Oral administration of ethyl acetate-soluble portion of *Terminalia chebula* conferring protection from streptozotocin-induced diabetic mellitus and its complications. *Biological and Pharmaceutical Bulletin*. 2011 Nov 1;34(11):1702-9.
53. Kumar GP, Arulselvan P, Kumar DS, Subramanian SP. Anti-diabetic activity of fruits of *Terminalia chebula* on streptozotocin induced diabetic rats. *Journal of health science*. 2006;52(3):283-91.
54. Bag A, Kumar Bhattacharyya S, Kumar Pal N, Ranjan Chattopadhyay R. Anti-inflammatory, anti-lipid peroxidative, antioxidant and membrane stabilizing activities of hydroalcoholic extract of *Terminalia chebula* fruits. *Pharmaceutical biology*. 2013 Dec 1;51(12):1515-20.
55. Saha S, Verma RJ. Antioxidant activity of polyphenolic extract of *Terminalia chebula* Retzius fruits. *J Taibah Univ Sci*. 2015.
56. Lee HS, Won NH, Kim KH, Lee H, Jun W, Lee KW. Antioxidant effects of aqueous extract of *Terminalia chebula* in vivo and in vitro. *Biological and Pharmaceutical Bulletin*. 2005;28(9):1639-44.
57. Zia-Ul-Haq M, Shahid SA, Ahmed S, Ahmad S, Qayum M, Khan I. Anti-platelet activity of methanolic extract of *Grewia asiatica* L. leaves and

Terminalia chebula Retz. fruits. Journal of Medicinal Plants Research. 2012 Mar 16;6(10):2029-32.

58. Sharma P, Prakash T, Kotresha D, Ansari MA, Sahrm UR, Kumar B, Debnath J, Goli D. Antiulcerogenic activity of Terminalia chebula fruit in experimentally induced ulcer in rats. Pharmaceutical Biology. 2011 Mar 1;49(3):262-8.

59. Na M, Bae K, Sik Kang S, Sun Min B, Kuk Yoo J, Kamiryo Y, Senoo YI, Yokoo S, Miwa N. Cytoprotective effect on oxidative stress and inhibitory effect on cellular aging of Terminalia chebula fruit. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives. 2004 Sep;18(9):737-41.

60. Nag G, De BR. Acetylcholinesterase inhibitory activity of Terminalia chebula, Terminalia bellerica and Emblica officinalis and some phenolic compounds. Int J Pharm Pharm Sci. 2011;3(3):121-4.

61. Sancheti S, Um BH, Seo SY. 1, 2, 3, 4, 6-penta-O-galloyl- $\beta$ -D-glucose: A cholinesterase inhibitor from Terminalia chebula. South African Journal of Botany. 2010 Apr 1;76(2):285-8.

62. Choi MK, Kim HG, Han JM, Lee JS, Lee JS, Chung SH, Son CG. Hepatoprotective effect of Terminalia chebula against t-BHP-induced acute liver injury in C57/BL6 mice. Evidence-Based Complementary and Alternative Medicine. 2015;2015.

63. Maruthappan V, Shree KS. Hypolipidemic activity of Haritaki (Terminalia chebula) in atherogenic diet induced hyperlipidemic rats. Journal of advanced pharmaceutical technology & research. 2010 Apr;1(2):229.

64. Hamada SI, Kataoka T, Woo JT, YAMADA A, YOSHIDA T, NISHIMURA T, OTAKE N, NAGAI K. Immunosuppressive effects of gallic acid and chebulagic acid on CTL-mediated cytotoxicity. Biological and Pharmaceutical Bulletin. 1997 Sep 15;20(9):1017-9.

65. Chang CL, Lin CS. Phytochemical composition, antioxidant activity, and neuroprotective effect of Terminalia chebula Retzius extracts. Evidence-Based Complementary and Alternative Medicine. 2012;2012.

66. Bag A, Bhattacharyya SK, Chattopadhyay RR. The development of Terminalia chebula Retz.(Combretaceae) in clinical research. Asian

Pacific journal of tropical biomedicine. 2013 Mar;3(3):244.

67. Gupta PC. Biological and pharmacological properties of Terminalia chebula Retz.(Haritaki)-An overview. Int J pharm pharm Sci. 2012;4(Suppl 3):62-8.

68. Upadhyay A, Agrahari P, Singh DK. A review on the pharmacological aspects of Terminalia chebula. Int. J. Pharmacol. 2014 Aug 15;10(6):289-98.

69. Saleem A, Husheem M, Härkönen P, Pihlaja K. Inhibition of cancer cell growth by crude extract and the phenolics of Terminalia chebula retz. fruit. Journal of Ethnopharmacology. 2002 Aug 1;81(3):327-36.

70. Jagtap AG, Karkera SG. Potential of the aqueous extract of Terminalia chebula as an anticaries agent. Journal of Ethnopharmacology. 1999 Dec 15;68(1-3):299-306.

71. Sharma A, Meena S, Barman N. Efficacy of ethyl acetate and ether extract of Terminalia chebula Retz against some human pathogenic strains. Inter J Pharm Tech Res. 2011;3(7):4-7.

72. Aher V, Wahi A. Immunomodulatory activity of alcohol extract of Terminalia chebula retz combretaceae. Tropical Journal of Pharmaceutical Research. 2011;10(5):567-75.

73. Suchalatha S, Srinivasan P, Devi CS. Effect of T. chebula on mitochondrial alterations in experimental myocardial injury. Chemico-biological interactions. 2007 Sep 20;169(3):145-53.

74. Sivachandran M, Hariharan P. Renoprotective effect of Terminalia chebula on gentamicin induced toxicity in rats. International Journal of Veterinary Science. 2012;1(2):76-9.

75. Singh D, Singh D, Choi SM, Zo SM, Painuli RM, Kwon SW, Han SS. Effect of extracts of Terminalia chebula on proliferation of keratinocytes and fibroblasts cells: an alternative approach for wound healing. Evidence-Based Complementary and Alternative Medicine. 2014;2014.

76. Ajala OS, Jukov A, Ma CM. Hepatitis C virus inhibitory hydrolysable tannins from the fruits of Terminalia chebula. Fitoterapia. 2014 Dec 31;99:117-23.

77. Pawar AT, Gaikwad GD, Metkari KS, Tijore KA, Ghodasara JV, Kuchekar BS. Effect of Terminalia chebula fruit extract on ethylene glycol induced urolithiasis in rats. Biomedicine & Aging Pathology. 2012 Jul 1;2(3):99-103.

78. Shin TY, Jeong HJ, Kim DK, Kim SH, Lee JK, Chae BS, Kim JH, Kang HW, Lee CM, Lee KC, Park ST. Inhibitory action of water soluble fraction of Terminalia chebula on systemic and local anaphylaxis. *Journal of ethnopharmacology*. 2001 Feb 1;74(2):133-40.

79. Naik GH, Priyadarsini KI, Naik DB, Gangabhagirathi R, Mohan H. Studies on the aqueous extract of Terminalia chebula as a potent antioxidant and a probable radioprotector. *Phytomedicine*. 2004 Sep 20; 11(6): 530-8.

80. Ramani YR, Pradhan S. Antiarthritic activity of acetone extract of Terminalia chebula. *WebmedCentral Pharmacology*. 2012; WMC003057, 3(2): 1-9.

81. Debnath J, Sharma UR, Kumar B, Chauhan NS. Anticonvulsant activity of ethanolic extract of fruits of Terminalia chebula on experimental animals. *International Journal of Drug Development and Research*. 2010 Oct; 2(4): 764-8.

**Table 1.** Mode of use of reviewed plants in jaundice

Sl. No	Scientific name	Local name	Parts used	Mode of use
1.	<i>Asparagus racemosus</i>	Satamuli	Root	Juice made from the tuberous roots
2.	<i>Terminalia chebula</i>	Horitoki	Fruit	Fruits are taken orally