

IN-SILICO STRUCTURE-ACTIVITY RELATIONSHIP AND MOLECULAR DOCKING STUDY OF LEVOFLOXACIN AND ITS MONO SUBSTITUTED ANALOGUES AGAINST THE ESCHERICHIA COLI AMINOGLYCOSIDE PHOSPHOTRANSFERASE

¹Sani Shedrach Bello, ²Itepu Victor E, *³Ogara Amaechi L. ⁴Uzoeto Henrietta Onyinye, ⁵Orum Terese Gabriel, ¹Nelson Christian, *⁶Durojaye Olanrewaju Ayodeji

¹Department of Biochemistry, University of Nigeria, Nsukka, Enugu State, Nigeria

²Department of Medical Biochemistry, College of Medical Sciences Edo State University Iyamho

³Department of Science Laboratory Technology University of Nigeria, Nsukka

⁴Department of Biological Sciences, Coal City University, Emene, Enugu State, Nigeria

⁵Department of Veterinary Public Health, University of Ibadan, Ibadan, Oyo State, Nigeria

⁶Department of Chemical Sciences, Coal City University, Emene, Enugu State, Nigeria

Email address: amaechi.ogara@unn.edu.ng; lanre.durojaye@yahoo.com

Abstract

Resistance to aminoglycoside antibiotics has had a profound impact on clinical practice. Despite their powerful bactericidal activity, aminoglycosides were one of the first groups of antibiotics to meet the challenge of resistance. The most prevalent source of clinically relevant resistance against these therapeutics is conferred by the enzymatic modification of the antibiotics. Aminoglycoside phosphotransferases provide an important means for high-level resistance to aminoglycoside antibiotics. Levofloxacin is used to treat a number of bacterial infections including acute bacterial sinusitis, pneumonia, urinary tract infections, chronic prostatitis, and some types of gastroenteritis. Along with other antibiotics it may be used to treat tuberculosis, meningitis, or pelvic inflammatory disease. Extensive structure activity relationship study was carried out with levofloxacin and its monosubstituted analogues which were designed by substituting the levofloxacin F group for C=O, CH₃, NH₂ and OH functional groups using the Marvin Sketch software. This was followed by a molecular docking study to target the experimental ligands against the E. coli aminoglycoside phosphotransferase active site. The result of the molecular docking study showed that the OH analogue of levofloxacin might be the better antibiotics to combat the resistance exhibited by the E. coli against aminoglycoside antibiotics.

Keywords: *Aminoglycoside; Resistance; Antibiotics; Levofloxacin; E. coli*

Introduction

The *Escherichia coli* (or *E. coli*) is the most prevalent infecting organism in the family of gram-negative bacteria known as enterobacteriaceae [1, 2]. *E. coli* bacteria were discovered in the human colon in 1885 by German bacteriologist Theodor Escherich who also showed that certain strains of the bacterium were responsible for infant diarrhea and gastroenteritis, an important public health discovery [3, 4].

The *E. coli* that are responsible for the numerous reports of contaminated foods and beverages are those that produce Shiga toxin, so called because the toxin is virtually identical to that produced by *Shigella dysenteriae* type 1 [5, 6]. The best-known and also most notorious *E. coli* bacteria that produce Shiga toxin is *E. coli* O157:H7 [7]. Shiga toxin-producing *E. coli* (STEC) cause approximately 100,000 illnesses, 3,000 hospitalizations, and 90 deaths annually in the United States. Most reported STEC infections are caused by *E. coli* O157:H7 [8, 9].

Enzymatic modification is the most common type of aminoglycoside resistance. Over 50 different enzymes have been identified [10]. Enzymatic modification results in high-level resistance [11, 12]. The genes encoding for aminoglycoside modifying enzymes are usually found on plasmids and transposons. Most enzyme-mediated resistance in gram-negative bacilli is due to multiple genes [13]. It is hypothesized that the enzymes are derived from organisms that make the aminoglycoside or from the mutation of genes that encode the enzymes involved in cellular respiration [14].

Levofloxacin is the levo isomer of the racemate ofloxacin, another quinolone antimicrobial agent [15]. Levofloxacin, a chiral fluorinated carboxyquinolone, is the pure (-)-(*S*)-enantiomer of the racemic ofloxacin [16]. Distinct functional groups on these molecules include a hydroxyl group, carbonyl group, and an aromatic ring [17]. Since levofloxacin is the *S*-enantiomer, it binds more effectively to the DNA gyrase enzyme and topoisomerase IV than its counterpart enantiomer. The substance is used as the hemihydrate, which has the empirical formula $C_{18}H_{20}FN_3O_4 \cdot \frac{1}{2} H_2O$ and a molecular mass of 370.38 g/mol [18].

The aim of this study is to evaluate the inhibitory effect of the modified analogues of levofloxacin against the *E. coli* aminoglycoside phosphotransferase in order to provide potent alternatives for the aminoglycoside antibiotics resistance exhibited by the *E. coli*.

Materials and Methods

Protein preparation

The crystal structure of the *Escherichia coli* aminoglycoside phosphotransferase, was obtained from the Protein Data Bank, PDB 3WoO (Figure 13). The protein structure was subjected to a refinement protocol using the Pymol viewer [19].

Designing of 6-Gingerol structural analogues

The structure of levofloxacin (Figure 1) was drawn with the Marvin Sketch software [20]. The structural analogues of levofloxacin were developed with structural modifications and different substituents [21]. The F substituent of levofloxacin was replaced with CH₃, OH, NH₂ and C=O groups. The structures were built with the Marvin Sketch software and minimized using the Chimera software [22, 23].

Molecular docking

Molecular docking was performed using AutoDock Vina Software [24]. Physicochemical, lipophilicity, solubility, pharmacokinetics and Lipinski drug likeness of levofloxacin and its analogues were determined using SwissADME Server [25].

Results and Discussion

The physicochemical property of proteins is critical for sustainability, efficiency, and stability in a biological system. Various physico-chemical parameters of proteins such as amino acid composition, extinction coefficient, instability index, grand average of hydropathicity (GRAVY), aliphatic index, theoretical pI, atomic composition and molecular weight allows us to understand the stability, activity and nature of the protein [26]. The result from the physicochemical property analysis of the *E. coli* aminoglycoside phosphotransferase showed that the enzyme is an acidic enzyme with a theoretical pI value in the acidic range of 5.16 [27]. The protein is made up of a total of 349 amino acids

as shown in table 1 with an estimated molecular weight of 39032.87 daltons.

The hydrophobicity scale produces a descriptive value for the relative hydrophobicity of residues of amino acids. The more positive or negative the value, the more hydrophobicity or hydrophilicity of the sequences located in that protein region [28]. The GRAVY calculator used in predicting the hydrophobicity assigned to the protein a value of -0.252.

The docking structures of all the compounds showed that they bind in a very similar pattern with the active site of *Escherichia coli* aminoglycoside phosphotransferase, as is evident from the superposition of the levofloxacin and all its 4 analogues in Figure 1. The interaction between levofloxacin and the different monosubstituted analogues with *Escherichia coli* aminoglycoside phosphotransferase shows steric interaction with the amino acid residues. The calculated free energy of binding of the levofloxacin and its analogues were -8.2, -8.7, -8.6, -8.2, and -7.6 Kcal/mol (Table 1). This confirms that the structural modification implemented in this study is significantly related to the increased antibiotic activity exhibited by the OH analogue of levofloxacin [29, 30]. This also proved the reliability of the docking results [31].

The solubility of a compound in water could improve its biotransformation and elimination as a drug [32]. Levofloxacin and all the substituted analogues were soluble in water (Table 1). The molecular weight of all the substituted derivatives including levofloxacin were less than 500g/mol, showing that they can be considered as drug. A compound can also be considered drug-like if it is characterized by high lipophilicity (less than 5) [33]. This is expressed as Log Po/w. The lipophilicity values of levofloxacin and all the monosubstituted compounds are less than 5 and are most likely to be drugs.

Lipinski's rule of 5 [34] helps in distinguishing between drug-like and non-drug-like molecules. It predicts high probability of success or failure due to drug likeness for molecules complying with 2 or more of the following rules: Molecular mass less than 500g/mol; High lipophilicity (expressed as Log Po/w less than 5); Less than 5 hydrogen bond

donors; Less than 10 hydrogen bond acceptors; Molar refractivity should be between 40-130. These filters help in early preclinical development and could help avoid costly late-stage preclinical and clinical failures [32]. Levofloxacin and all the monosubstituted analogues violated none of the Lipinski's rule and therefore are likely to be drugs (Table 1).

High penetration is needed for most of the drugs targeting the central nervous system (CNS), whereas blood brain barrier (BBB) penetration should be minimized for non-CNS drugs to avoid undesired side-effects [35]. Pharmacokinetically, the gastrointestinal drug absorption of all the substituents was high and could not cross the blood brain barrier (BBB). This shows that they cannot cause any problem to the brain.

For synthetic accessibility, values of 5 to 10 means that the drug could be synthesized [32]. Levofloxacin and all its analogues showed values less than 4. This means that the compounds can easily be synthesized in the laboratory. Synthetic studies followed by pre-clinical studies are further recommended.

Conclusion

An In-Silico Structure Activity Relationship and molecular docking study was carried out on the *Escherichia coli* aminoglycoside phosphotransferase, using levofloxacin and four of its structurally diverse analogues as the experimental compounds. The results obtained indicated that the OH analogue might have a better functional activity having shown a lower binding energy score against the target enzyme. These analogues also pose no threat to the Central Nervous System (CNS) as they do not penetrate the blood brain barrier.

Synthesis and pre-clinical studies of the OH analogue of levofloxacin, targeted at the *Escherichia coli* aminoglycoside phosphotransferase active site is therefore recommended.

References

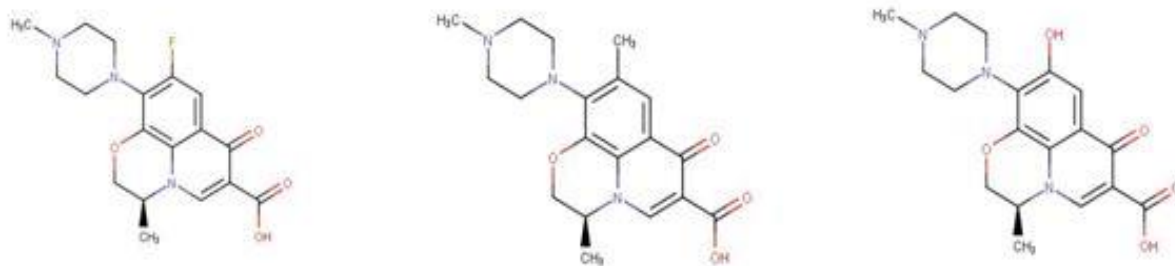
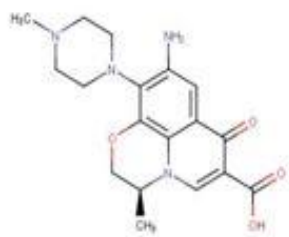
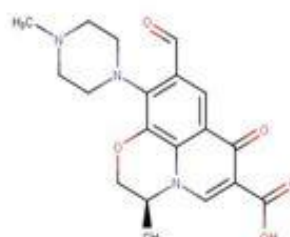
1. Umezawa, H.; Okanishi, M.; Kondo, S.; Hamana, K.; Utahara, R.; Maeda, K.; Mitsunashi, S. Phosphorylative inactivation of aminoglycosidic antibiotics

- by *Escherichia coli* carrying R factor. *Science* 1967, 157, 1559–1561. [CrossRef] [PubMed]
2. Doi, O.; Miyamoto, M.; Tanaka, N.; Umezawa, H. Inactivation and phosphorylation of kanamycin by drug-resistant *Staphylococcus aureus*. *Appl. Microbiol.* 1968, 16, 1282–1284. [PubMed]
 3. Miller, G.H.; Sabatelli, F.J.; Naples, L.; Hare, R.S.; Shaw, K.J. The changing nature of aminoglycoside resistance mechanisms and the role of isepamicin—A new broad-spectrum aminoglycoside. The Aminoglycoside Resistance Study Groups. *J. Chemother.* 1995, 7, 31–44. [PubMed]
 4. Mitscher, L.A. *Antibiotics and Antimicrobial Agents*; Lippincott Williams & Wilkins: Baltimore, PA, USA, 2002; pp. 788–791.
 5. Weinstein, M.J.; Luedemann, G.M.; Oden, E.M.; Wagman, G.H. Gentamicin, a new broad-spectrum antibiotic complex. *Antimicrob. Agents Chemother.* 1963, 161, 1–7. [PubMed]
 6. Martin, C.M.; Ikari, N.S.; Zimmerman, J.; Waitz, J.A. A virulent nosocomial *Klebsiella* with a transferable R factor for gentamicin: Emergence and suppression. *J. Infect. Dis.* 1971, 124, 24–29. [CrossRef]
 7. Benveniste, R.; Davies, J. R-factor mediated gentamicin resistance: A new enzyme which modifies aminoglycoside antibiotics. *FEBS Lett.* 1971, 14, 293–296. [CrossRef]
 8. Woo, P.; Dion, H.; Bartz, Q. Butirosins A and B, aminoglycoside antibiotics. III. structures. *Tetrahedron Lett.* 1971, 12, 2625–2628. [CrossRef]
 9. Hayashi, S.F.; Norcia, L.J.; Seibel, S.B.; Silvia, A.M. Structure activity relationships of hygromycin A and its analogs: Protein synthesis inhibition activity in a cell free system. *J. Antibiot.* 1997, 50, 514–521. [CrossRef] [PubMed]
 10. Shaw, K.J.; Rather, P.N.; Hare, R.S.; Miller, G.H. Molecular genetics of aminoglycoside resistance genes and familial relationships of the aminoglycoside-modifying enzymes. *Microbiol. Rev.* 1993, 57, 138–163. [PubMed]
 11. Hotta, K.; Zhu, C.B.; Ogata, T.; Sunada, A.; Ishikawa, J.; Mizuno, S.; Kondo, S. Enzymatic 20 -N-acetylation of arbekacin and antibiotic activity of its product. *J. Antibiot.* 1996, 49, 458–464. [CrossRef] [PubMed]
 12. Labby, K.J.; Garneau-Tsodikova, S. Strategies to overcome the action of aminoglycoside-modifying enzymes for treating resistant bacterial infections. *Future Med. Chem.* 2013, 5. [CrossRef] [PubMed]
 13. Gillings, M.R.; Paulsen, I.T.; Tetu, S.G. Genomics and the evolution of antibiotic resistance. *Ann. N. Y. Acad. Sci.* 2017, 1388, 92–107. [CrossRef] [PubMed]
 14. Wolf, E.; Vassilev, A.; Makino, Y.; Sali, A.; Nakatani, Y.; Burley, S.K. Crystal structure of a GCN5-related N-acetyltransferase: *Serratia marcescens* aminoglycoside 3-N-acetyltransferase. *Cell* 1998, 94, 439–449. [CrossRef]
 15. Morrissey, I.; Hoshino, K.; Sato, K.; Yoshida, A.; Hayakawa, I.; Bures, MG.; Shen, LL. (August 1996). "Mechanism of differential activities of ofloxacin enantiomers" (PDF). *Antimicrob Agents Chemother.* 40 (8): 1775–84. doi:10.1128/AAC.40.8.1775. PMC 163416. PMID 8843280. Archived (PDF) from the original on 11 June 2011.
 16. Kannappan, Valliappan; Mannemala, Sai Sandeep (7 June 2014). "Multiple Response Optimization of a HPLC Method for the Determination of Enantiomeric Purity of S-Ofloxacin". *Chromatographia.* 77 (17–18): 1203–1211. doi:10.1007/s10337-014-2699-4.
 17. Mouzam, M. I.; Dehghan, M. H. G.; Asif, S.; Sahuji, T.; Chudiwal, P. Preparation of a novel floating ring capsule-type dosage form for stomach specific delivery. *Saudi Pharmaceutical Journal* 2011, 19, 85-93.
 18. McGregor, J. C.; Allen, G. P.; Bearden, D. T. Levofloxacin in the treatment of complicated urinary tract infections and

- acute pyelonephritis. Therapeutics and clinical risk management 2008, 4, 843-853.
19. WL DeLano, Pymol: An open-source molecular graphics tool. CCP4 Newsletter On Protein Crystallography. 2002,40: 82-92.
 20. OToure, CGDussap, A. Lebert. Comparison of Predicted pKa Values for Some Amino-Acids, Dipeptides and Tripeptides, Using COSMO-RS, ChemAxon and ACD/Labs Methods". Oil & Gas Science and Technology – Review. IFP Energies nouvelles.2013. 68 (2): 281–291. doi:10.2516/ogst/2012094.
 21. R McBride. ChemAxon opens shop in 'heart' of Boston biotech hub".2012, Retrieved 11 May 2014.
 22. TD Goddard, C CHuang, E C Meng, EF Pettersen, GS Couch, JH Morris, TE Ferrin. UCSF chimera: meeting modern challenges in visualization and analysis". Protein Science, 2017,27 (1).
 23. EF Pettersen, TDGoddard, CC Huang, GS Couch, DM Greenblatt, EC Meng, TE Ferrin. UCSF Chimera—a visualization system for exploratory research and analysis. Journal of Computational Chemistry.2004, 25 (13): 1605–12.
 24. O Trott, AJ Olson, AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading, Journal of Computational Chemistry. 2010, 31 (2): 455–461, doi:10.1002/jcc.21334.
 25. A Daina, O, Michielin, V Zoete. A free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. Scientific Report, 2017,7: 42717.
 26. Gasteiger E., Hoogland C., Gattiker A., Duvaud S., Wilkins M.R., Appel R.D., Bairoch A. (2005). Protein Identification and Analysis Tools on the ExPASy Server; (In) John M. Walker (ed): The Proteomics Protocols Handbook, Humana Press (2005). 571-607.
 27. Shi Q, Zhou Y, Sun Y. Influence of pH and ionic strength on the steric mass-action model parameters around the isoelectric point of protein. Biotechnol Prog. 2005;21:516–23. [PubMed]
 28. Kyte, J. and Doolittle, R.F. (1982) A simple method for displaying the hydropathic character of a protein. J. Mol. Biol. 157, 105-132. [PubMed: 7108955]
 29. DB Kitchen, H Decornez, J R Furr, J Bajorath. Docking and scoring in virtual screening for drug discovery: methods and applications. Nature Review of Drug Discovery. 2004, 3(11):935–949.
 30. N Moitessier, P Englebienne, D Lee. J Lawandi, C R Corbeil. Towards the development of universal, fast and highly accurate docking/scoring methods: a long way to go. British Journal of Pharmacology. 2008,153(1): 7–26.
 31. BQ Wei. "Testing a flexible-receptor docking algorithm in a model binding site". Journal of Molecular Biology 337:5, 2004, 1161-1182.
 32. O.A Durojaye. U.I. Njoku. S. Cosmas. E.N Akpan. M.M Gayam. (2018). In-Silico Structure–Activity Relationship and Molecular Docking study of Levofloxacin and its Monosubstituted Analogues against the Escherichia coli DNA Gyrase. International Journal of chemistry and pharmaceutical Science, (2018), 6(4) 121-126.
 33. Durojaye, O.A, Ajuluchukwu, P. S, Cosmas S, Igomu, R. O, Okagu, I. U, S.B, Sani. (2018). Binding Energy Prediction and Molecular Docking Studies of Falcarindiol and its Monosubstituted Analogues against Aspergillus fumigatus Chitinase; The In Silico Pharmacokinetics. International Journal of Scientific and Engineering Research, 9(6) 271-283.
 34. CA Lipinski. Lead- and drug-like compounds: the rule of-five revolution. Drug Discovery Today: Technologies.2004 1(4): 337-341.
 35. DE Clark, In silico prediction of blood-brain barrier permeation. Drug Discovery Today, 2003, 8: 927–933.

Table 1: Physicochemical, lipophilicity, solubility, pharmacokinetics and Lipinski druglikeness of Levofloxacin and its monosubstituted analogues

Parameters	Levofloxacin	CH ₃ analogue of Levofloxacin	OH analogue of Levofloxacin	NH ₂ analogue of Levofloxacin	C=O analogue of Levofloxacin
Molecular weight g/mol	361.37	357.40	359.38	358.39	371.39
Docking score Kcal/mol	-8.2	-8.2	-8.6	-8.2	-7.6
Num. H-Bond acceptors	6	5	6	5	6
Num. H-Bond donors	1	1	2	2	1
Molar Refractivity	101.83	106.84	103.90	106.28	107.26
Lipophilicity Consensus Log P _{o/w}	1.10	1.12	0.45	0.29	0.52
Water Solubility Class	Very Soluble	Soluble	Very Soluble	Very Soluble	Very Soluble
GI absorption	High	High	High	High	High
BBB permeant	No	No	No	No	No
P-gp substrate	Yes	Yes	Yes	Yes	Yes
CYP1A2 inhibitor	No	No	No	No	No
CYP2C19 inhibitor	No	No	No	No	No
CYP2C9 inhibitor	No	No	No	No	No
CYP2D6 inhibitor	No	Yes	No	No	No
CYP3A4 inhibitor	No	No	No	No	No
Lipinski Druglikeness	Yes; 0 violation	Yes; 0 violation	Yes; 0 violation	Yes; 0 violation	Yes; 0 violation
Synthetic accessibility	3.63	3.66	3.65	3.66	3.67

Figure 1: Structural formula of levofloxacin and its modified analogues**Levofloxacin****CH₃ analogue of Levofloxacin****OH analogue of Levofloxacin****NH₂ analogue of Levofloxacin****C=O analogue of Levofloxacin**

Structural formula

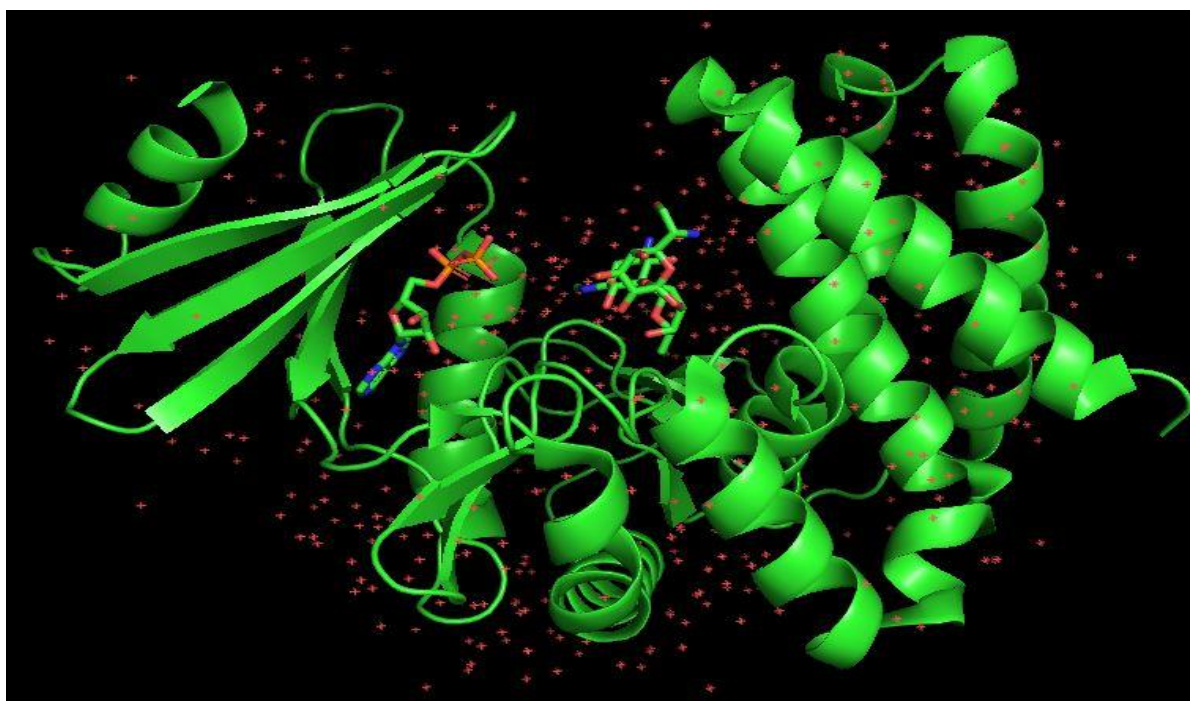
Figure 2: Crystal structure of the *E. coli* aminoglycoside phosphotransferase PDB 3Wo0

Figure 3: Levofloxacin in complex with *E. coli* aminoglycoside phosphotransferase.

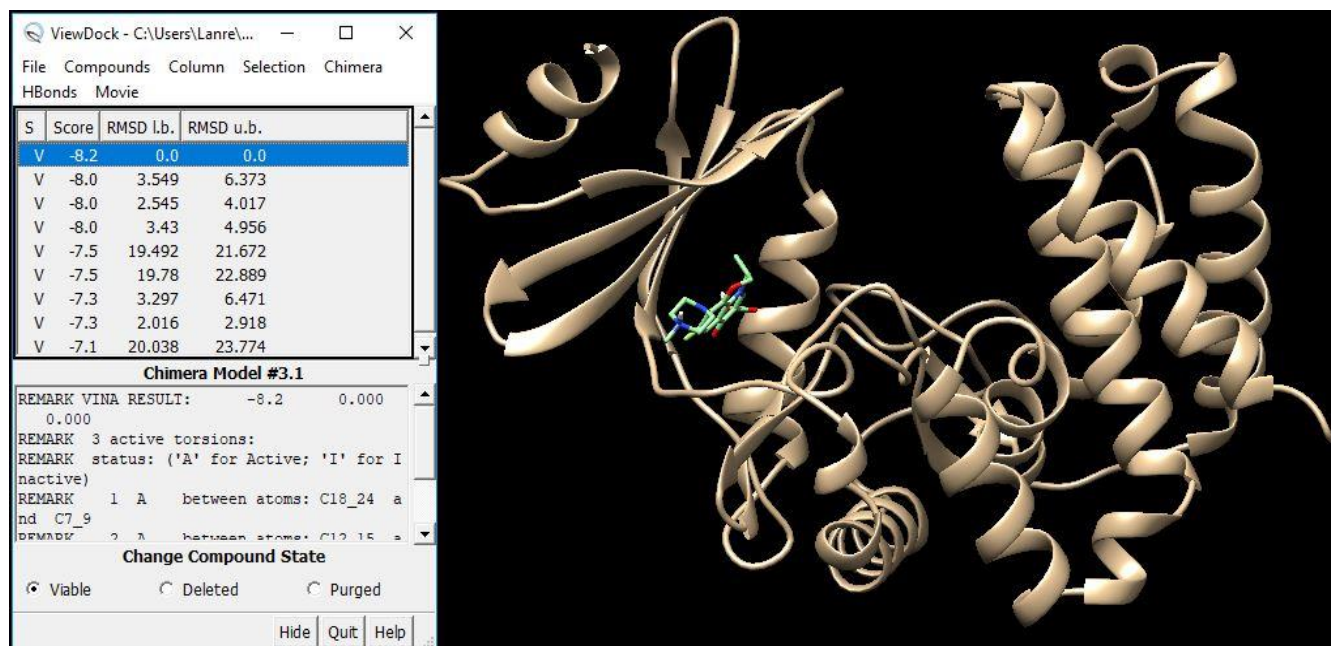


Figure 4: OH analogue of levofloxacin in complex with *E. coli* aminoglycoside phosphotransferase.

