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ANTIBACTERIAL AND FREE RADICAL SCAVENGING ACTIVITY OF METHANOL EXTRACT OF CRYSTELLA DENATATA (LEAVES)

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Abstract

Antibiotic resistance is getting global treat and also many diseases are associated with oxidative stress caused by free radicals. The present study evaluated the in vitro antioxidant and antibacterial activities of methanol extract of *Crystella denatata*. In vitro antioxidant activities of the plant extract were determined by DPPH scavenging method as compared with standard antioxidants. Extract showed significant radical scavenging activity as $IC_{50}.25.27 \mu g/ml$ comparable IC_{50} 8.687 $\mu g/ml$ of standard ascorbic acid. Mild antibacterial activities were comparable with standard drug, Kanamycin. The present study provides evidence that *Crystella denatata* prove to be potent natural antioxidants and could replace synthetic antioxidants. Plants can also be used against pathogenic bacterial strains.

Keywords: Crystella denatata, antibacterial, free radical, ascorbic acid, kanamycin

Introduction

In the past, ancient people greatly depended on local plants (flora) for their survival. They experimented on different part of the plants and sometimes some plant extract shows real and beneficial effect e.g. ancient Rhubarb plant which contained anthraquinones and it was used as a lead compound in the design of laxative drug named dantron^{[1].} A free radical is a molecules or molecular fragments which have one or more unpaired electrons in atomic or molecular orbital ^[2]. There is evidence that most of the degenerative disease which persecute humanity have their origin in detrimental free radical reaction. The disease caused by free radicals includes atherosclerosis, cancer, inflammatory joint disease, asthma, diabetes, senile dementia, biological ageing ^[3]. Free radicals are produced in cells as part of normal cellular functions. Therefore excess production of free radical in the body might play a role in much disease. In this case, antioxidants prevent tissue damage caused by free radical through preventing the formation of free radical, scavenging them and promoting their decompositions ^[4]. Plant contains different compounds which possess antioxidant property. Plant antioxidant are composed of a of compounds like varietv ascorbic acid. tocopherols, polyphenolic compounds which exert important function both in plant and human. Flavonoid is another polyphenolic compound which has a capacity to act as antioxidants [5-6]. Recently in "in vitro" test phenolics have been considered powerful antioxidants. It is proved to be more potent antioxidants than vitamin C and E and carotinoids [7-8]. Antibiotic resistant is considered as global health challenge. Globally 3.5% of new cases and 20.5% of previously treated cases are caused by tuberculosis and these strains are resistant to rifampicin and isoniazid ^[9]. Carbapenem resistant enterobacteriaceae species have isolated recently ^[10]. As day passes antibiotic resistant species of bacteria are increased. On the contrary new antibiotic are not developed. To face this great thread, plant source may be used to develop new antibiotic.

Methods

Sample collection and extraction

Leaves of Christella dentata were collected during March 2017 from Manikganj district, Bangladesh, and taxonomically identified at the Bangladesh Herbarium National (Accession Number43854). The undesirable materials were removed from the plants and sun dried for one week after cutting into small pieces. The dried plants were powdered with a mechanical grinder. The powder was extracted by using 98% methanol and the bottle was stand at room temperature and allowed for stand for 7 days. The extract was filtered off with vacuum filter to remove debris of plant. The filtrate (methanol extract) obtained was evaporated under ceiling fan and in water-bath until dried. After drying, the crude extract was stored at 4°C in refrigerator.

Phytochemical Screening Methods

Phytochemical studied was carried out for identification of different chemical groups which present as described [11-13].

DPPH radical scavenging assay

Antioxidant activity of the methanol leaves extract was estimated by using stable free radical DPPH (1, 1- Diphenyl-2-Pycrylhydrazyl) quantitatively ^[14-17]. The potential activity of methanol extract was calculated on the basis of their scavenging activity of stable DPPH radical. At first 25 mg extract was mixed with 25 ml of methanol to prepare 1000 µg/ml solution of extract as stock solution. Then five concentration of sample were prepared by serial dilution method .these concentration were 500, 250, 125, 62.5, 31.25 µg/ml. After that 1 ml. of solution of each concentration was taken into test tubes designed for each concentration. 3 ml. of 0.004% DPPH solution was added to each concentration and kept for 30 minutes at dark place to allow any reaction that is to be occurred. After 30 minutes, absorbance by UVwas measured spectrophotometer at 517 nm. Ascorbic acid was used as reference standard. The percent of inhibition was calculated by using following formula-

% of inhibition

 $=\frac{blank \ absorbance - sample \ absorbance}{blank \ absorbance} \times 100$

 $IC_{5^{\circ}}$ value was determined from % of inhibition vs. concentration graph which is discussed in result and discussions.

Antimicrobial Activity

Disc Diffusion Assay

Agar diffusion testing method was developed for assaying antimicrobial susceptibility [18]. Sterile blank discs (5mm) were saturated with the test extract at the doses of 500 µg/disc using micropipette. The desired concentration of extract was prepared by using methanol. Blank (control disc) were also prepared by using methanol. Both sample and control discs were dried. Sample containing discs, standard antibiotic discs (Kanamycin 30 µg/disc, Oxoid Ltd, UK) and control discs were placed in Petri dishes containing nutrient agar medium seeded with the test pathogens using sterile forceps. Then Petri dishes were transferred into incubator and incubated at 37 °C for 20 h. After incubation, zone of inhibition was measured using digital slide calliper [19-20].

Results

Phytochemical screening

DPPH scavenging assay

Anti-bacterial test

Discussion

The excessive use of antibiotic was contributed to the spread of antibiotic resistant bacteria in communities all over the world. Medicinal plant may be used to develop antibiotic against multidrug resistant bacteria ^[21]. Again antioxidant are prominent as prophylactic for certain disease like diabetes, cardiovascular cancer, disease, neurodegenerative disorders. These agents scavenge the reactive oxygenated species and prevent damage caused by them ^[22]. In the present investigation, extracts of C. dentata was evaluated for investigation of antimicrobial activity against some gram negative and gram positive bacteria.

The antioxidant activity of methanolic extract of *C. dentata* was evaluated by using standard DPPH free radical scavenging method. The IC_{50} of the *C. dentata* was 25.27µg/ml, whereas IC_{50} of Ascorbic Acid as a standard was 8.687 µg/ml. The extract showed antibacterial activity against all the tested

pathogen at the doses of 500 μ g/disc compare with standard kanamycin at the dose 30 μ g/disc which is shown at the table 5 with zone of inhibition. Our findings confirm that significant antioxidant activity and modest antimicrobial activity.

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References

- 1. Graham L. Patrick (an introduction to medicinal chemistry- fifth edition)
- Pala, Funda Sibel, and Hakan Gürkan. "The role of free radicals in ethiopathogenesis of diseases." Advances in Molecular Biology 1 (2008): 1-9.
- 3. Aust N Z J opthalmol.1995 Feb;23(1):3-7
- 4. I S Young, J V Woodside; Antioxidants in health and disease; Journal of Clinical Pathology, March 2001
- 5. J.Graßmann;Terpenoids as Plant Antioxidants;Vitamins & HormonesVolume 72, 2005, Pages 505-535
- 6. Shashank Kumar and Abhay K. Pandey;Chemistry and Biological Activities of Flavonoids: An Overview;The Scientific World Journal ,Volume 2013, Article ID 162750, 16 pages
- Rice-Evans, C.A.; Miller, N.J.; Bolwell, P.G.; Bramley, P.M.; Pridham, J.B. The relative antioxidant activities of plant-derived polyphenolic flavonoids. Free Radic. Res. 1995, 22, 375–383.
- 8. Rice-Evans, C.A.; Miller, N.J.; Paganga, G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. Free Radic. Biol. Med. 1996, 20, 933–956.
- 9. Global tuberculosis report 2014-WHO.
- 10. Carl Llor and Lars Bjerrum;Antimicrobial resistance: risk associated with antibiotic overuse and initiatives to reduce the problem;Ther Adv Drug Saf. 2014, Vol. 5(6) 229 -241.

- 11. Evans WC. Pharmacognosy: London, W.R. Saunders; 2002.
- 12. Mohammad Ali; Text book of Pharmacognosy, Second Edition 1998.
- 13. AbdulGhani,Practicalphytochemistry. First edition; 2005.
- 14. Talukder C, Saha S, Adhikari S, Mondal HK, Islam MK, Anisuzzman M. Evaluation of antioxidant, analgesic and antidiarrhoeal activity of Flacourtia jangomas (Lour.) Raeusch leaves. Pharmacologyonline 2012; Arch.3:2028.
- 15. Saha S, Hossain F, Anisuzzman M, Islam MK. Pharmacological evaluation of Musa seminifera Lour. fruit. J Integr Med 2013 Apr; Epub ahead of print.
- 16. Sadhu SK, Okuyama E, Fujimoto H, Ishibashi M. Separation of Leucas aspera, a medicinal plant of Bangladesh, guided by prostaglandin inhibitory and antioxidant activities. Chem Pharm Bull, 2003; 51: 595-598.
- 17. Sharma OP, Bhat TK. DPPH antioxidant assay revisited. Food Chem, 2009; 113:1202-1205.
- Mounir Balouiri, Moulay Sadiki, Saad Koraichi Ibnsouda; Methods for in vitro evaluating antimicrobial activity: A review; journal of pharmaceutical analysis; volume 6, issue 2; page 71-73.
- 19. Ríos JL, Recio MC, Villar A. Screening methods for natural products with antimicrobial activity: a review of the literature. J Ethnopharmacol, 1988; 23: 127-149.
- 20. Kelmanson JE, Jager AK and Vaan Staden J. Zulu medicinal plants with antibacterial activity.J.Ethanopharmacol, 2000; 69: 241-246
- 21. Prof Dr Ali Esmail Al-Snaf;iAntimicrobial effects of medicinal plants (part 3): plant based review;IOSR Journal Of Pharmacy;Volume 6, Issue 10 Version; PP. 67-92
- 22. D. Venkat Ratnam,D.D.Ankola,V.Bhardwaj,D.KSahana,

M.N.VRavi Kumar; Role of antoxidants in prophylaxis and therapy:a pharmaceutical purpose;journal of control release;volume 113;issue 3;page 189-207

Table 1: phytochemical tests of C.dentata

Tested groups	Methanol extract of C. dentata	
Alkaloid	+	
Reducing Sugar group	+	
Flavonoids	+	
Saponin	+	
Tannin	+	

(+) indicates the presence.

Concentration	% of DDDU acquerging by According acid	% of DDDU acquire give by C dout at a	
Concentration	% of DPPH scavenging by Ascorbic acid	% of DPPH scavenging by C. dentata	
500	86.86	90.87	
250	80.60	82.80	
125	75.78	74.21	
62.5 69.25		63.157	
31.25 60.55		51.75	

Table 2: % of scavenging by Ascorbic acid and extract C. dentata.

Table 3: IC_{50} values of the extracts of C.dentata and standard.

Test sample	Regression line	R ² value	IC ₅₀ (µg/ml)
Ascorbic acid	y = 9.2289ln(x) + 30.048	0.9891	8.687
Crystella dentata	y = 14.123ln(x) + 4.3679	0.9936	25.27

Bacterial strains	Type of bacteria	Diameter of zone of inhibition		
		Blank	Kanamycin(30µg/disc)	Extract (500 µg/disc)
Escherichia coli	Gram (-)	-	34 mm	8 mm
Salmonella typhi	Gram (-)	-	40 mm	8 mm
Staphylococcus epidermis	Gram (+)	-	32 mm	-
Bacillus subtilis	Gram (+)	-	38 mm	9 mm



Figure 1: Anti-oxidant activity of ascorbic acid and extract.