

GAS CHROMATOGRAPHY MASS SPECTROMETRIC ANALYSIS OF CARBOXYLIC ACIDS IN THE HERBS OF TWO DRACOCEPHALUM L. SPECIES

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Abstract

The genus *Dracocephalum* L. (*Lamiaceae* Martinov family) includes 72 species of essential oil bearing plants with promising biological activities. They belong to the category of unofficial medicinal plants. The most studied group of the bioactive compounds of *Dracocephalum* species are essential oils and polyphenols, while the other components have been investigated much less. The aim of the study was the gas chromatography mass spectrometric (GC-MS) analysis of carboxylic acids in the herbs of two *Dracocephalum* species (*D. moldavica* L. and *D. grandiflorum* L.). The plants were grown under the cultivation conditions of Ukraine. There were identified 14 short-chained carboxylic acids in the *D. moldavica* herb and 16 ones in the *D. grandiflorum* herb. Ferulic acid possessing prominent healing properties prevailed among the found aromatic acids in both species. 14 fatty acids were found in the *D. moldavica* herb and 13 of them in the *D. grandiflorum*. Unsaturated α -linolenic acid (ω_3) dominated among the long-chained carboxylic acids in the raw materials of both plants. Thus, the studied species could be regarded as the promising sources of some carboxylic acids.

Keywords: *Dracocephalum moldavica*, *Dracocephalum grandiflorum*, herb, short-chained carboxylic acid, fatty acid, GC-MS analysis

Introduction

Plants produce a diverse number of specialized secondary metabolites which are biosynthetically originated from the primary metabolites [1]. Secondary metabolites play decisive roles in plant adaptation to different environmental factors as well as they are widely used in medicine and human nutrition. The secondary metabolites are synthesized from the certain primary compounds (precursors), such as sugars, amino acids, and carboxylic acids, which are essential for the life of whole plant organisms. Many of the components of primary synthesis as well as secondary ones possess the proven therapeutic activity [2–7].

Short-chained carboxylic acids are quite diverse in structure and biological properties [3, 4]. They can exist in the free state or as the salts, esters, etc. Certain organic acids such as citric, succinic, ferulic, benzoic, and salicylic were defined as the pharmacologically active substances. Some of them are used in the food industry (malic, lemon, benzoic, fumaric acids) as well as in perfume manufacturing (for instance, esters of benzoic and phenyl acetic acids) [3–5].

Polyunsaturated fatty acids play an important role in functioning of the human body [4, 7]. Their deficiency leads to the dysfunction of cell membranes and cell homeostasis at all. So, the appropriate intake of polyunsaturated fatty acids can prevent various health disorders such as diabetes mellitus, chronic inflammation, atherosclerosis [8–13]. Polyunsaturated fatty acids are not synthesized in the human body. Thus, they must be constantly supplied with food or dietary supplements.

The genus *Dracocephalum* L. (*Lamiaceae* Martinov family) includes 72 species of essential oil bearing herbaceous plants and subshrubs distributed in temperate and sub-tropical regions [13, 14]. They belong to the unofficial medicinal plants with promising biological activities. The most studied group of their bioactive compounds are essential oils and polyphenols, while the other components have been investigated much less [14, 16–18].

There is a relatively small portion of scientific information on the contents of carboxylic acids in the raw materials of the *Lamiaceae* representatives, e.g. *Dracocephalum* species [8, 19, 20]. Thus, this

aspect of their chemical composition deserves much more attention.

The aim of the study was the gas chromatography mass spectrometric (GC-MS) analysis of carboxylic acids in the herbs of two *Dracocephalum* species (*D. moldavica* L. and *D. grandiflorum* L.) which are cultivated in Ukraine.

Methods

The herbs of investigated plants were harvested from the experimental plots in Temopil region (Ukraine) and shade dried for 6–8 days.

The sample preparation for GC-MS started from the weighing of an exact portion of grinded raw material (0.05 g). It was placed in a vial (2 mL). Then the internal standard (50 µg of tridecane in hexane) and 1.0 mL of methylating agent (14% boron trichloride in methanol, Supelco 3-3033) were added. The mixture was kept for 8 h in a hermetically sealed vial at a temperature of 65°C for the extraction of fatty oil, its hydrolysis and methylation. The reaction mixture was drained from the precipitate of plant material and diluted by adding 1.0 mL of distilled water. To remove the fatty acid methyl esters, 0.2 mL of dichloromethane was added and it was gently shaken several times during 1 h. Then the resulting mixture of methyl esters was chromatographed.

The component compositions of the obtained methyl esters of carboxylic acids were detected by GC-MS method according to [21]. Chromatograph Agilent Technologies 6890 N with mass spectrometric detector 5973 N was used in the analysis. The chromatographic capillary column was Innowax (30 m × 0.25 mm). The rate of helium as a carrier gas was 1.2 mL/min. The temperature of the thermostat have been programmed in the range 50–250 °C with a rate growing 4 °C/min. The library of mass spectra NIST 05 and WILEY 2007 was taken into account for the identification of components.

Results

The GC-MS method was used for the simultaneous investigation of short-chained carboxylic acids and short-chained fatty acids in the herbs of *D. moldavica* and *D. grandiflorum* (Tables 1, 2 and Figures 1, 2).

There were identified 14 short-chained carboxylic acids in the *D. moldavica* herb and 16 ones in the *D. grandiflorum* herb (Table 1). The total amount of the short-chained carboxylic acids was higher in the *D. moldavica* herb (8818 mg/kg) comparatively to *D. grandiflorum* (4977 mg/kg).

As can be seen from the data of Table 1, the aliphatic carboxylic acids prevailed over aromatic acids both quantitatively and in a variety of composition. The malonic, oxalic, malic and citric acids dominated in the aliphatic group. As it is known, the detected acids play an important role in human life [4, 6, 19, 22]. For instance, citric acid is used in medicine as a part of agents that improve energy metabolism; malic and oxalic acids are used in food industry; malonic acid is a precursor in the biosynthesis of flavonoids.

The hydroxycinnamic ferulic acid with prominent nootropic and antioxidant properties [23] prevailed among the found aromatic acids. Found benzoic acid is often used as a food additive due to its antioxidant activity [24]. Accumulation of aromatic acids in the raw materials of plants is of great interest for their further using as antioxidants and anti-inflammatory agents [23–25].

The composition of long-chained carboxylic acids known as fatty acids presented in Table 2. There were identified 14 fatty acids in the *D. moldavica* herb and 13 of them in the *D. grandiflorum* herb. The total amount of the fatty acids was similar: 3726 mg/kg in the *D. moldavica* herb and 3790 mg/kg in the *D. grandiflorum*.

As can be seen from Table 2, unsaturated α -linolenic acid (ω_3) dominated among the all fatty acids in the raw material of both plants. The detected predominant α -linolenic and linoleic polyunsaturated acids play an important role in the construction of cell membranes, regulation hormone metabolism, and can prevent the deposition of cholesterol on the blood vessels [10, 20]. Researchers also found the dominance of linoleic and linolenic acids in the aboveground parts of several *Lamiaceae* species from the *Mentha*, *Salvia*, *Satureja*, *Ocimum* genera [8, 19, 20]. The content of carboxylic acids in plant raw materials may vary dependently on the genetic peculiarities of plant, geographical location, cultivation season, method of harvesting, etc. [26, 27].

Conclusion

The conducted GC-MS analysis revealed the chromatographic profiles of carboxylic acids in the *Dracocephalum* herbs. There were detected 14 short-chained carboxylic acids in the *D. moldavica* herb and 16 ones in the *D. grandiflorum* herb. Ferulic acid prevailed among the found aromatic acids in both species. 14 fatty acids were found in the *D. moldavica* herb and 13 of them in the *D. grandiflorum*. Unsaturated α -linolenic acid dominated among the fatty acids in the raw materials of both plants. Thus, the herbs of the studied species could be regarded as promising sources of new food additives and medicines with preventing and healing properties.

References

1. Maeda, H.A. (2019). Evolutionary diversification of primary metabolism and its contribution to plant chemical diversity. *Front Plant Sci*, 10, 81.
2. Bakoglu, A., Kilic, O., & Kokten, K. (2016). Fatty acid composition of the leaves of some *Salvia* taxa from Turkey. *Chemistry of Natural Compounds*, 52(4), 676–678.
3. Kovalyova, A. M., Ilna, T.V., Osmadko, A. P., Koshovyi, O. M., & Grudko, I. V. (2020). Carboxylic acids from herbs of *Veronica austriaca*, *V. cuneifolia*, and *V. armena*. *Chemistry of Natural Compounds*, 56(6), 1111–1113.
4. Marchyshyn, S. M., Shanayda, M. I., Kernychna, I. Z., Demydiak, O. L., Dakhym, I. S. et al. (2016). Investigation of the qualitative composition and contents of organic acids in the overground part of plants Families *Lamiaceae*, *Asteraceae*, *Apiaceae* and *Chenopodiaceae*. *Int J of Medicine and Med. Research*, 2(1), 19–22.
5. Mykhailenko, O, Ivanauskas, L, Bezruk, I, Sidorenko, L., Lesyk, R., & Georgiyants, V. (2021). Characterization of phytochemical components of *Crocus sativus* leaves using HPLC-MS/MS and GC-MS: a new potential by-product. *Sci Pharm*, 89(2), 28–45.
6. Savych, A., & Nakonechna, S. (2021). Determination of amino acids content in two herbal mixtures with antidiabetic activity by GC-MS. *Pharmakeftiki*, 33(2), 116–123.
7. Savych, A., Marchyshyn, S., & Basaraba, R. (2020). Determination of fatty acid composition

- content in the herbal antidiabetic collections. *Pharmacia*, 67(3), 153–159.
8. Shanaida, M., Kernychna, I., & Shanaida, Yu. (2017). Chromatographic analysis of organic acids, amino acids, and sugars in *Ocimum americanum* L. *Acta Poloniae Pharmaceutica – Drug Research*, 74(2), 729–732.
 9. Upchurch, R. G. (2008). Fatty acid unsaturation, mobilization, and regulation in the response of plants to stress. *Biotechnol. Lett*, 30, 967–977.
 10. Gasmi, A., Mujawdiya, P. K., Shanaida, M., Ongenae, A., Lysiuk, R., et al. (2020). Calanus oil in the treatment of obesity-related low-grade inflammation, insulin resistance, and atherosclerosis. *Appl Microbiol Biotechnol*, 104(3), 967–979.
 11. Mykhailenko, O., Kovalyov, V., Kovalyov, S., Toryanik, E., Osolodchenko, T., & Buidin, Y. (2017). Fatty acid composition of lipids of *Iris sibirica*. *Ceska a Slovenska Farmacie*. 66(5), 220–227.
 12. Savych, A., & Mazur, O. (2021). Antioxidant activity *in vitro* of antidiabetic herbal mixtures. *PharmacologyOnLine*, 2, 17–24.
 13. Stechyshyn, I., Pavliuk, B., Chornij, N., Kritsak, M., & Prokopovych, O. (2021). Substantiation of attention around antioxidants of plant origin for use in complex pharmacotherapy of diseases of the oral cavity. *Polski Merkurusz Lekarski*, 49(291), 238–241.
 14. Kakasy, A. Z. (2006). New phytochemical data on *Dracocephalum* species. Theses of the PhD dissertation. Budapest, 14 p.
 15. The Plant List (2021): <http://www.theplantlist.org>
 16. Barchuk, O., Pryshlyak, A., Shanaida, M. (2021). Chemical compositions and sedative activities of the *Dracocephalum moldavica* L. and *Ocimum americanum* L. essential oils. *PharmacologyOnLine*, 2, 179–187.
 17. Ehsani, A., Alizadeh, O., Hashemi, M., Afshari, A., & Aminzare M. (2017). Phytochemical, antioxidant and antibacterial properties of *Melissa officinalis* and *Dracocephalum moldavica* essential oils. *Vet Res Forum*, 8(3), 223–229.
 18. Weremczuk-Jeżyna, I., Grzegorzczak-Karolak, I., Frydrych, B. et al. (2017). Rosmarinic acid accumulation and antioxidant potential of *Dracocephalum moldavica* L. cell suspension culture. *Not Bot Horti Agrobo*, 45(1), 215–219.
 19. Cacan, E., Kokten, K., & Kilic, O. (2018). Leaf fatty acid composition of some *Lamiaceae* taxa from Turkey. *Progress in Nutrition*, 20 (Suppl. 1), 231–236.
 20. Maffei, M., & Scannerini, S. (1993). Fatty acid variability from non-polar lipids in some *Lamiaceae*. *Biochemical Systematics and Ecology*, 21(4), 475–486.
 21. Carrapiso, A. I., García, C. (2000). Development in lipid analysis: some new extraction techniques and *in situ* transesterification. *Lipids*, 35(11), 1167–1177.
 22. Jarukas, L., Mykhailenko, O., Baranauskaite, J., Marksa, M., & Ivanauskas, L. (2020). Investigation of organic acids in saffron stigmas (*Crocus sativus* L.) extract by derivatization method and determination by GC/MS. *Molecules*, 25, 3427–3437.
 23. Michels, B., Zwaka, H., Bartels, R., Lushchak, O., Franke, K., et al. (2018). Memory enhancement by ferulic acid ester across species. *Sci Adv*, 24, 4(10).
 24. Mao, X., Yang, Q., Chen, D., Yu, B., & He, J. (2019). Benzoic acid used as food and feed additives can regulate gut functions. *BioMed research international*, 5721585.
 25. Guzyk, M. M., Dyakun, K. O., Yanytska, L. V., Pryvrotska, I. B., Krynytska, I. Ya. et al. (2017). Inhibitors of poly(ADP-Ribose) polymerase-1 as agents providing correction of brain dysfunctions induced by experimental diabetes. *Neurophysiology*, 49, 183–193.
 26. Kumar, V., Sharma, A., Bhardwaj, R., & Thukral, A.K. (2017). Analysis of organic acids of tricarboxylic acid cycle in plants using GC-MS, and system modeling. *J Anal Sci Technol*, 8, 20–29.
 27. Olennikov, D. N., Chirikova, N. K., Kashchenko, N. I., Gomostai, T. G., Selyutina, I. Yu., & Zilfikarov I. N. (2017). Effect of low temperature cultivation on the phytochemical profile and bioactivity of arctic plants: a case of *Dracocephalum palmatum*. *Int J Mol Sci*, 30, 18(12), 2579.

Table 1. Composition and contents of the short-chained carboxylic acids in the herbs of two *Dracocephalum* species, mg/kg

Acid	Retention time, min	<i>D. moldavica</i>	<i>D. grandiflorum</i>
Dimethoxyacetic	8.88	49	37
Oxalic	9.30	245	195
Malonic	11.59	4835	3067
Fumaric	12.34	172	107
Furfurylic	13.02	13	19
Succinic	13.42	159	54
Benzoic*	13.91	27	39
Phenylacetic*	16.88	10	4
Salicylic*	17.13	18	17
Malic	21.68	636	420
Azelaic	24.21	21	15
Citric	29.0	2525	895
Vanillic*	31.97	29	19
Syringic*	37.40	-	31
<i>p</i> -Hydroxycinnamic*	38.94	-	5
Ferulic*	39.80	79	53
Total amount		8818	4977

Note: * – aromatic acid; "-" – component was not detected.

Table 2. Composition and contents of the fatty acids in the herbs of two *Dracocephalum* species, mg/kg

Acid	Retention time, min	<i>D. moldavica</i>	<i>D. grandiflorum</i>
Myristic	21.92	124	114
Palmitic	25.86	1439	1400
Palmitoleic*	26.18	127	95
7- Hexadecenic*	26.69	8	-
14-Methylpalmitic	26.90	21	-
Heptadecanic	27.54	18	15
Stearinic	29.31	141	116
Oleic*	29.59	76	227
11-Octadecenic*	29.72	26	34
Linoleic*	30.37	381	327
α -Linolenic*	31.48	1275	1379
Arachidic	32.59	31	31
2-Oxipalmitic	32.65	35	17
Behenic	35.66	24	13
Lignoceric	38.52	-	9
Total amount		3726	3790

Note: * – unsaturated fatty acid; "-" – the component was not detected.

Abundance

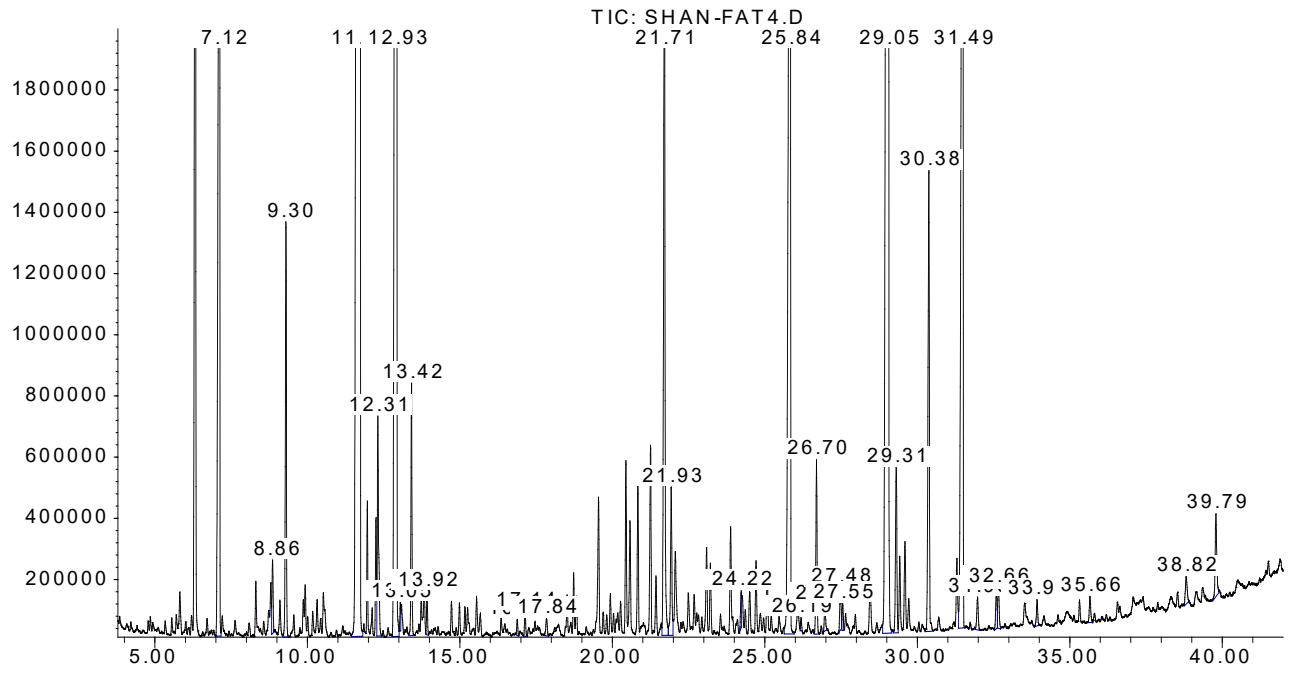


Figure 1. The GC-MS chromatogram of carboxylic acids methyl esters in the *D. moldavica* herb

Abundance

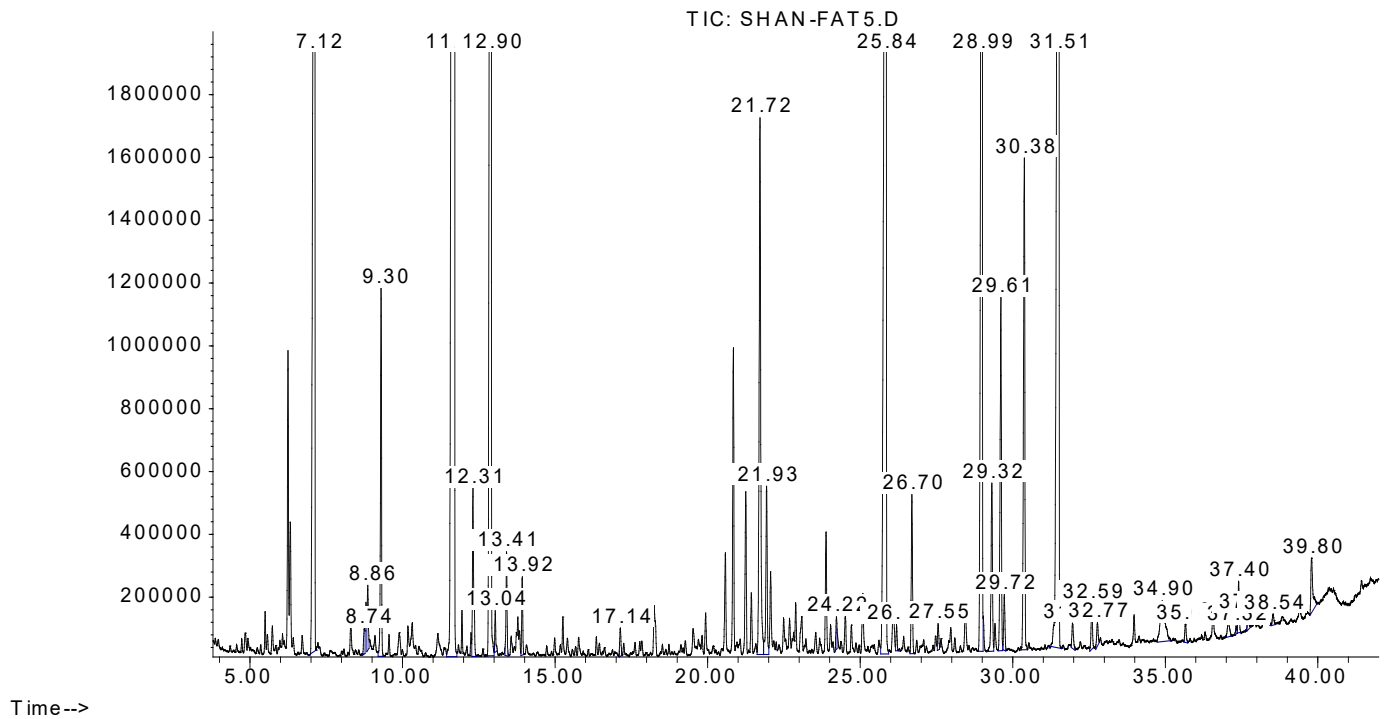


Figure 2. The GC-MS chromatogram of carboxylic acids methyl esters in the *D. grandiflorum* her