

TROPANE ALKALOIDS: AN OVERVIEW^a

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Summary

Tropane alkaloids are a group of secondary metabolites containing a 8-azabicyclo[3.2.1]octane nucleus skeleton as a key structural element. They have hallucinogenic features and are active on Central Nervous System. Some of them, such as atropine and scopolamine, are used in medicine as antimuscarinic drugs. Tropane alkaloids are abundant above all in Solanaceae and Erythroxylaceae, where they are the most important alkaloids, but they were extracted also from other families of higher plants, e.g. Proteaceae, Euphorbiaceae, Rhizophoraceae, Convolvulaceae, Cruciferae and Moraceae. All these secondary metabolites, containing the tropane core as main structure, have as a common biosynthetic origin the amino acid L-ornitine. This compound, after several biosynthetic steps, is converted in hygrine, from which α -tropine, β -tropine, ecgonine and cuscohygrine origin and constitute the bases for the biosynthesis of all the tropane alkaloids known. The diffuse presence of tropane alkaloids among higher plants, can support the idea that they can be considered as a chemical link among different families of plants.

Keyword: Chemotaxonomy, Tropane Alkaloids, Atropine, Scopolamine.

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Introduction

Chemotaxonomy, or chemosystematics, is the attempt to classify plants according to the differences in their biochemical features. Recently, it becomes more evidently the importance of a chemosystematic approach in chemical and/or pharmacological studies on biologically active plants.

One of the most fascinating field of research is constituted by plants containing compounds active on Central Nervous System and, among them by plants containing alkaloids. In this paper we attempt to consider a link between tropane alkaloids pharmacological activity and their distribution in different families of higher plants.

All tropane alkaloids contain the 8-azabicyclo[3.2.1]octane nucleus skeleton as key structural element (Fig.1).

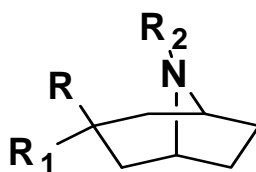


Figure 1. 8-azabicyclo [3,2,1]octane nucleus.

Since tropane alkaloids have a quite simple chemical structure and are easy to be extract, they have been identified not only in Solanaceae and Erythroxylaceae, where they are the most abundant alkaloids, but also in other families, e. g. Proteaceae, Euphorbiaceae, Rhizophoraceae, Convolvulaceae, Cruciferae and Moraceae (1).

Plants containing tropane alkaloids have been medicinally used for centuries for their different pharmacological activities. Among the most important alkaloids, while cocaine (**1**) acts as an anaesthetic and a sympathomimetic, classical tropane alkaloids, as atropine (**2**) and scopolamine (**3**), are parasympatholytic; in fact, they act as competitive antagonists, by blocking the action of acetylcholine at parasympathetic sites in smooth muscle, secretory glands and CNS. For this reason, they are classified as antimuscarinic drugs. But it is also important to remind the hallucinogenic characteristics of some tropane alkaloids extract above all from the Solanaceus genera *Brugmansia* and *Datura*. In fact, in most native American cultures, along with medicinal plants, these plants play a central role in therapeutic rites based on “magical plants”, which are seen as intermediaries between the human world and that of supernatural forces (2).

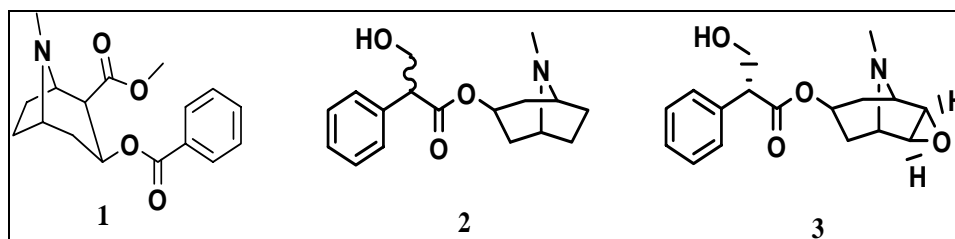


Figure 2. Cocaine (**1**); (+/-)-hyoscyamine or atropine (**2**); hyoscyine or scopolamine (**3**).

In this paper we present an overview on the chemotaxonomy of plant families containing tropane alkaloids and on their biological activities.

Results

Solanaceae

Solanaceae is the family where tropane alkaloids are really abundant. This family is usually divided by botanists in different subfamilies and tribes, characterized by chemical differences in their composition. (1).

In the subfamily Solanoideae, the tribe Datureae, and specifically *Datura* and *Brugmansia* genera, contains the greatest range of tropane alkaloids. The aerial parts of *Datura* genus contain above all hyoscyamine (2) but also scopolamine (3), littorine (4) and the base 6 β -(2-methylbutanoyloxy)tropan-3 α -ol (5), while the roots yield valtropine, a tropane alkaloid common in *Duboisia* species (3).

Recently, polyhydroxylated tropane alkaloids have been reported in *Datura* species: in the leaves of *Datura wrightii* Regel and in tubers of *Solanum* spp., calystegine A₃ (1 β ,2 α ,3 β -trihydroxynortropane) (6) and calystegine B₂ (1 β ,2 α ,3 β ,4 α -tetrahydroxynortropane) (7) have been found, while calystegines C₁ (1 β ,2 β ,3 β ,4 β ,6 β -pentahydroxynortropane) (8) and C₂ (1 β ,2 α ,3 β ,4 α ,6 β -pentahydroxynortropane) (9) have been isolated from *Duboisia leichhardtii* F. Muell. (3).

The aerial parts of some species belonging to *Brugmansia* genus contain scopolamine (3) and/or hyoscyamine (2) as principal alkaloids, with smaller amounts of derivatives of these bases (10), while the roots contain in addition a large number of esters formed from dihydroxytropane and teloidine (10) (9).

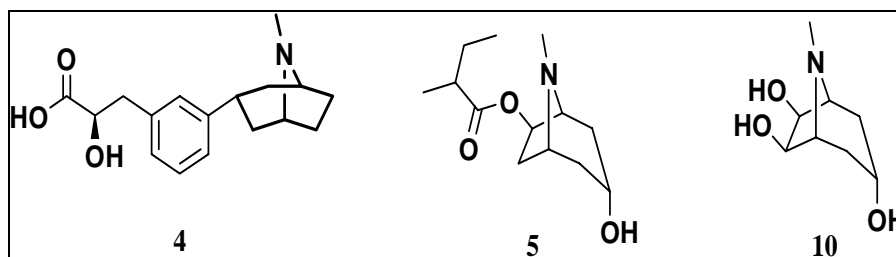


Figure 3. Littorine (4); 6 β -(2-methylbutanoyloxy)tropan-3 α -ol (5); teloidine (10).

The genus *Solandra*, (tribe Solandreae) contains atropine (2) and/or hyoscyamine (3) and their nor-derivatives as principal alkaloids (4).

Atropa belladonna L. (tribe Solaneae) contains hyoscyamine (2) as the principal alkaloid but also scopolamine (3), apoatropine (11), hyoscyamine (2) and scopolamine *N*-oxides (5). Leaves of *A. belladonna* and of *Hyosciamus* spp. contain also a few quantity of calystegines A₃ (6), B₁ (12) and B₂ (7). Hyoscyamine (2) and tigloidine (13) have been found in some species of the minor genera *Latua* and *Acristus*. Tigloidine (3 β -tigloyloxytropane) and 3 α -tigloyloxytropane were isolated as root alkaloids (6-7). Some secotropane alkaloids have been isolated, as major alkaloids, from the roots and have been subsequently identified as (+)-physoperuvine (14), racemic physoperuvine and (+)-*N,N*-dimethyl-physoperuvinium salt (8). Extracts from leaves and roots contain 3 β -acetoxytropane and *N*-methylpyrrolidinylhygrine (two isomers), but also 3 β -tigloyloxytropane (15), hygrine (16), physoperuvine (14), tropine (17) and cuscohygrine (18) (9). Roots of *Physalis alkekengi* L. yielded tigloidine (13), 3 α -tigloyloxytropane (15), cuscohygrine (18), the hygrine dimer phygrine and some unidentified bases. Calystegines A₃ (6), B₂ (7), A₅ (1 β ,3 β ,4 α -trihydroxynortropane) (19) and B₃ (1 β ,2 β ,3 β ,4 α -tetrahydroxynortropane) (20) were extracted from the roots of *Physalis alkekengi* L.(9), while from *Lycium chinense* Mill. calystegines C₂ (9), B₄ (21), A₇ (22) and *N*-methylcalystegines were isolated (10).

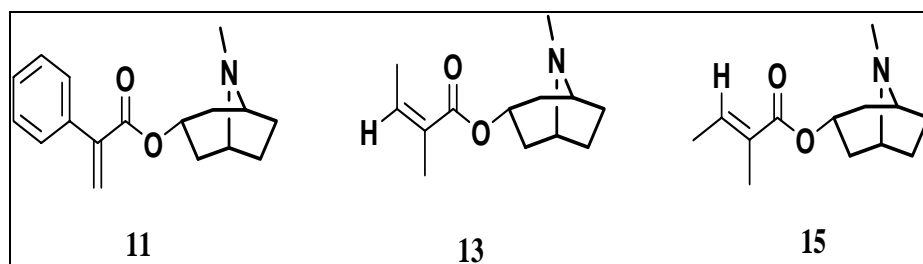


Figure 4. Apoatropine (**11**); tigloidine (**13**); tigloyloxytropane (**15**).

The large genus *Solanum* does not contain the usual tropane alkaloids (**11**): calystegine A₃ (**6**) has been found along with calystegine B₂ (**7**) in the leaves of some *Solanum* species, including *S. tuberosum* L., *S. dulcamara* L., *S. melongena* L. and in herbarium fragments of *S. kwebense* N. E. Br. ex C. H. Wright from Southern Africa (12).

In conclusion, calystegines are located in *A. belladonna* (all parts but mainly upper leaves), *Mandragora officinarum* L. (likewise, mainly spring leaves), *Scopolia carniolica* Jacq. (spring leaves and flowers), *Hyoscyamus niger* L. (traces in all plant parts) and *Solanum tuberosum* L. (sprouts only). Specifically, *Hyoscyamus niger* L. contains also an alkaloid with a bridgehead aminogroup, calystegine N₁ (**23**) (13).

For many centuries *Mandragora* roots have been considered very important as folk medicines. The former *Mandragora officinalis* Mill. comprises two distinct species, *M. autumnalis* Bertol. and *M. vernalis* Bertol., which contain hyoscyamine (**2**), scopolamine (**3**), cuscohygrine (**18**), apoatropine (**11**), 3 α -tigloyloxytropane (**15**) and 3,6-ditigloyloxytropane. The roots contain small quantities of cuscohygrine (**18**), pseudotropine (**24**), tropine (**17**) and hyoscyamine (**2**). *Withania somnifera* Dunal likewise contains cuscohygrine (**18**) and 3 α -tigloyloxytropane (**15**), but also tropine (**17**) and pseudotropine (**24**) (14). The main base from the roots of *Cyphomandra betacea* Cav. is N,N'-bis-(4-dimethylaminobutyl)-hexamide. Other bases included tropinone (**25**), cuscohygrine (**18**), hyoscyamine (**2**), tigloidine (**13**), tropine (**17**) and pseudotropine (**24**) (5).

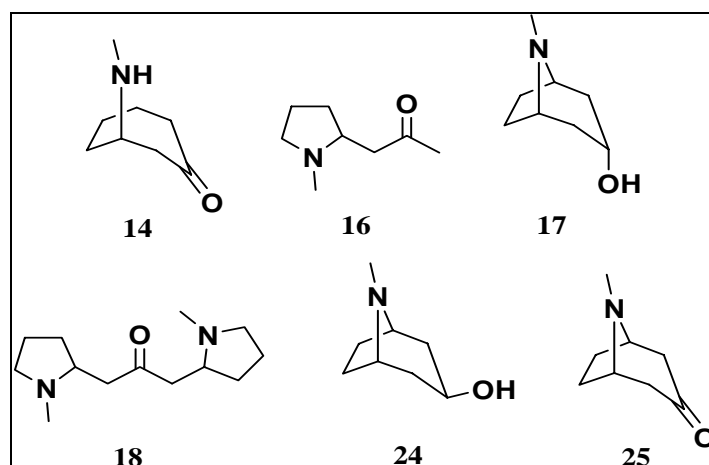


Figure 5. Physoperuvine (**14**); hygrine (**16**); tropine (**17**); cuscohygrine (**18**); pseudotropine (**24**); tropinone (**25**).

N-oxides were found in *Scopolia* spp. and *Hyoscyamus* spp. (tribe Hyoscyameae) (6) which, along with *Physochlaina* spp. and *Przewalskia* spp., contain hyoscyamine (2) as the major alkaloid with some scopolamine (3) derivatives (5, 6).

To the subfamily Cestroideae belong the tribes Anthocercideae, Nicandreae, and Salpiglossidae.

The tribe Anthocercideae includes seven genera, *Duboisia*, *Anthocercis*, *Cyphanthera*, *Anthotroche*, *Symonanthus*, *Grammosolen* and *Crenidium*.

Among *Duboisia* genus, *Duboisia myoporoides* L. Br. contains scopolamine (3) (15), the dominant alkaloid in a variety collected North of Gosford, NSW Australia, or hyoscyamine (2), the major alkaloid in a variety South of Gosford. In a region known as Acacia Plateau near Killarney, Queensland, Australia, a third variety was discovered in which nicotine (26) and anabasine (27) are the dominant alkaloids (16). *Duboisia leichhardtii* F. Muell and *D. myoporoides* R. Br. contain hyoscyamine (2) and scopolamine (3) as the most important tropane alkaloid (17), but from the first one was isolated calystegine C₂ (9), too. In *Duboisia arenitensis* Craven the alkaloid content is less than the other species and the most important are scopolamine (3) and hyoscyamine (2).

Anthocercis spp. contain either hyoscyamine (2) or scopolamine (3) as predominant alkaloids, frequently accompanied by their respective apo- and nor-derivatives. Among this genus, *Anthocercis littorea* Endl. contains mainly littorine (4) and meteloidine (28) (18) but also some mono- and ditigloyl esters of teloidine (10) (19); *Anthocercis viscosa* R. Br. and *A. fasciculata* F. Muell. contain hyoscyamine (2) (18); *Cyphanthera anthocercidea* Haegi (= *Anthocercis frondosa*) yields scopolamine (3) and hyoscyamine (2), but chiefly nicotine (26) (20).

Among *Cyphanthera* genus, *Cyphanthera albicans* Miers (= *Anthocercis albicans*) affords butyryl esters of tropine and 6 β -hydroxytropine. The alkaloids of *Cyphanthera odgersii* Haegi and *Cyphanthera tasmanica* Miers were found to be consistent with other known *Cyphanthera* species (11). Three species belonging to *Anthotroche* genus, *A. myoporoides* C. A. Gardner, *A. pannosa* Endl. and *A. walcottii* F. Muell., contain hyoscyamine (2), norhyoscyamine (29), apoatropine (11) and scopolamine (3) (20).

Among *Symonanthus* genus (two species), the roots of *Symonanthus aromaticus* Haegi contain mono- and ditigloyl esters like *Datura* species and both aerial parts and roots have scopolamine (3) and its derivative aposcopolamine (30), as the main alkaloids (11).

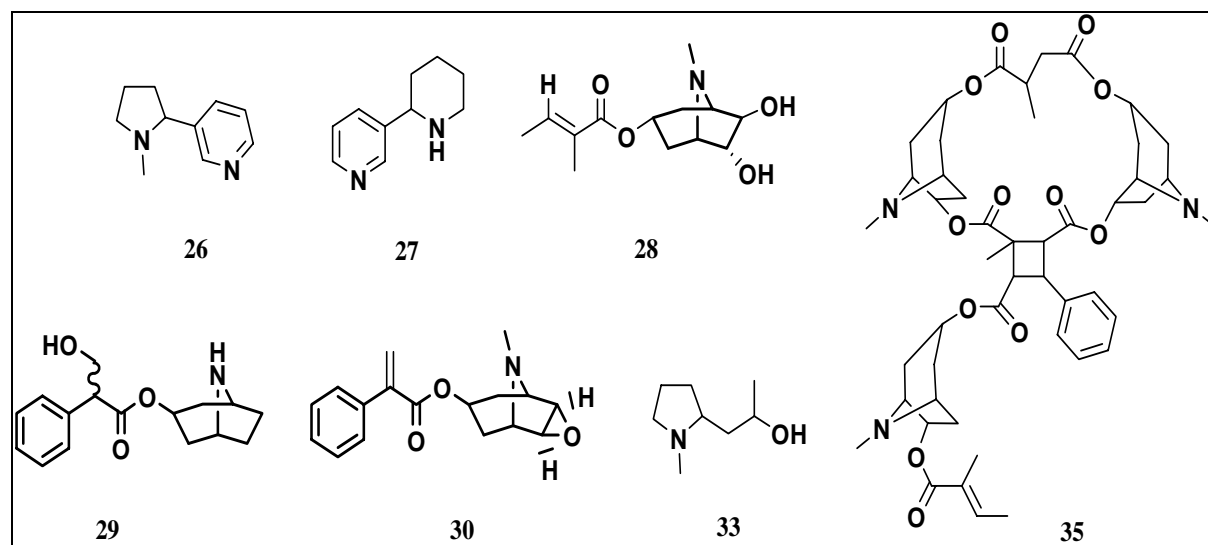


Figure 6. Nicotine (26); anabasine (27); meteloidine (28); norhyoscyamine (29); aposcopolamine (30); hygroline (33); grahamine (35).

The genus *Grammosolen* (two species) appears most closely related chemically to *Cyphanthera* and *Anthotroche* (11).

The genus *Crenidium* has a single species, *C. spinescens* Haegi, where hyoscyamine (**2**) is the predominant alkaloid, but are present also anabesine (**27**) with ursolic acid (**31**). *Anthocercis ilicifolia* Hook possesses a similar alkaloid spectrum to that of *A. littorea* Endl. *Anthocercis genistoides* Miers shows meteloidine (**28**) as the major alkaloid (11).

Esters of tropic acid (**32**) are not found in the tribe *Nicandreae*. However, tropine (**17**) was isolated from the roots of *Nicandra* species (21).

Schizanthus pinnatus Ruiz (tribe Salpiglossidae) contains tropane-derivate as schizanthines A and B which are 6 β -seneciolyloxytropan-3 β -ol esters of dibasic mesaconic acid (A is a diester of 6-seneciolyloxytropine, B is a mixed ester of 6-seneciolyloxytropine and ethanol) (22). The root alkaloids of *S. hookeri* Lodd. are tropine (**17**), a pair of diastereoisomeric hygrolines (**33**) and the alkaloids 3 α -seneciolyloxytropan-6 β -ol and 6 β -angeloyloxytropan-3 α -ol (22). *Schizanthus litoralis* Phil. contains hydroxytropane esters, hygrolines (**33**), the tropane diester of itaconic acid (**34**) and the alkaloids 6 β -seneciolyloxytropan-3 α -methylmesaconate, 6 β -cinnamoyloxytropan-3 α -methylmesaconate, 6 β -seneciolyloxytropan-3 α -ol, *cis*- and *trans*-N-(4-hydroxyphenyl)-ferulamides (23) but also 3 α -seneciolyloxytropan-6 β -ol, (-)-hygroline and (+)-pseudohygroline, which had already been reported present in *S. pinnatus* Ruiz (24). Schizanthines C, D, E, and a trimeric tropane alkaloid occur in *S. grahamii* Borrer ex Baker (25). Grahamine (**35**) contains a cyclobutane ring substituted by three carboxyl groups and is formed from cinnamoyltropine and mesaconic acid (**36**).

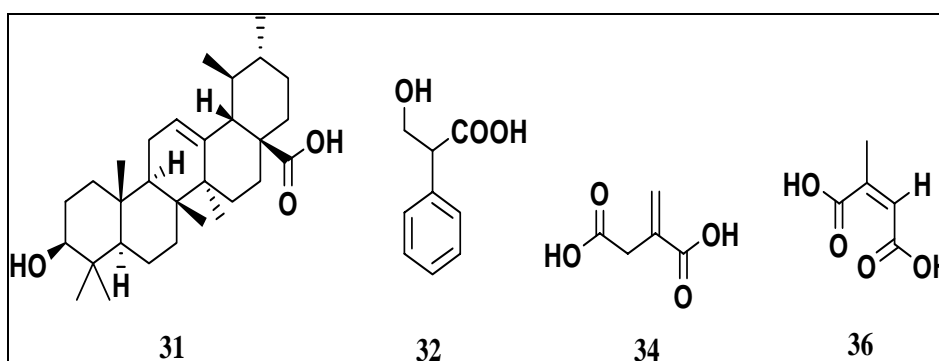


Figure 7. Ursolic acid (**31**); tropic acid (**32**); itaconic acid (**34**); mesaconic acid (**36**).

Erythroxylaceae

The genus *Erythroxylum* contains about 200 species. *Erythroxylum argentinum* O. E. Schulz, *E. cataractarum* Spruce ex Peyr, *E. cumanense* Kunth, *E. glaucum* O. E. Schulz, *E. mamacoca* Mart. and *E. shatona* Macbride contain total alkaloids in the range 0.06–0.20% compared with all species from the sections *Macrocalyx*, *Rhabdophyllum* and *Leptogramme* save one, which were lower in alkaloid content (0.002–0.04%) (11). In the six *Erythroxylum* spp. of *Archerythroxylum*, benzoic and phenylacetic acids form esters with alkamines, principally tropan-3-ols, tropan-3,6-diols and their derivatives. 3 β -Benzoyloxytropane is the most frequently occurring alkaloid, together with nortropacocaine (3 β -benzoyloxynortropane) extracted from *E. mamacoca* Mart.. Dihydrocuscohygrine and cuscohygrine (**18**) are present in some species. Cuscohygrine (**18**) as the main alkaloid of *E. cataractarum* Spruce ex Peyr is unique in the genus.

Tropacocaine (**37**) is extracted from *E. ulei* O. E. Schulz (section *Leptogramme*), *E. mamacoca* Mart. and *E. argentinum* O. E. Schulz (section *Archerythroxyllum*), but not in the sections *Marcrocalyx* and *Rhabdophyllum* (11). Dihydrocuscohygrine and cuscohygrine (**18**) are also found in the cultivated cocaine producing species, *E. coca* Lam. and *E. novogranatense* Morris, and its variety *truxillense*. Cocaine (**1**) and the *cis* and *trans* isomers of cinnamoyl cocaine (**38**) is contained in *E. coca* Lam. and *E. novogranatense* Morris var. *truxillense* (26). A series (α , β , γ , δ , ϵ) of dimeric methylecgonine (methyl ester of ecgonine, the basic moiety of cocaine) esters of truxillic acid (1,3-diphenylcyclobutane-2,4-dicarboxylic acids, dimers of cinnamic acid) was identified in Bolivian coca leaf and α -truxilline (**39**) and β -truxilline were the most abundant (27). A new alkaloid, 1-hydroxytropacocaine, was discovered in *E. novogranatense* Morris var. *novogranatense* and var. *truxillense* (28). The base 6 β -benzoyloxytropan-3 α -ol occurs in both *E. cumanense* Kunth and *E. glaucum* O. E. Schulz (both section *Archerythroxyllum*). Trimethoxybenzoyl esters are found in the roots of *E. cumanense* Kunth. *Erythroxyllum macrocarpum* O. E. Schulz and *E. sideroxyloides* Lam. of the section *Packylobus* contain a similar range of alkaloids consisting mainly of benzoyl esters of tropan-3 α -ol, tropan-3 β -ol and tropan-3 α ,6 β -diol together with their *nor*-derivatives. Three alkaloids have been found, 3 α -benzoyloxytropan-6 β -ol (from *E. sideroxyloides* Lam.), 3 α -benzoyloxynortropane and 3 β -benzoyloxynortropan-6 β -ol (both species) (29). The root barks of *E. pervillei* Baill. and *E. hypericifolium* Lam. contain the bases 3 α -(3-hydroxyphenylacetoxy)-tropane, (+)-3 α -phenylacetoxytropan-6 β -ol, 6 β -acetoxy-3 α -phenylacetoxytropane, 3 α -phenylacetoxytropan-6 β ,7 β -diol, 3 α -phenylacetoxytropane and 3 α -phenylacetoxytropane (29). Important to be noted are also the alkaloids 3 α -cinnamoyloxytropan-6 β -ol isolated as the (+) base from leaves of *Knightia* spp. (30). This is the first pseudotropine ester to be found in *Erythroxyllum hypericifolium* Lam. The benzoyl analogue is widely distributed in *Erythroxyllum* spp. The alkaloids 3 α -cinnamoyloxytropane, 3 α ,6 β -dicinnamoyloxytropane (analogous to the dibenzoyl ester found in *E. cuneatum* Kurz (31)), 3-cinnamoyloxytropan-6-ol (stereochemistry not established), 6 β -acetoxy-3 α -cinnamoyloxytropane and 6-phenylacetoxytropan-3-ol are found in *Crossostylis* spp. (32), together with cinnamate tropane dimers and esters of truxillic acid (33). These were identified as 3,3'-truxilloxyloxy-6'-acetoxyditropane, 3,3'-truxilloxyloxy-6-hydroxy-6'-acetoxyditropane and 3,3'-truxilloxyloxy-6'-hydroxyditropane. As in root bark, in the stem bark of *E. hypericifolium* Lam. predominate esters of phenylacetic acid: 3 α -phenylacetoxytropan-6 β -ol, the main alkaloid, 6 β -acetoxy-3 α -benzoyloxytropane, 3-acetoxy-6-phenylacetoxytropane and hygrine (**16**) (34). *Erythroxyllum zambesiaceum* N. Robson (section *Melanocladus*) likewise has a complex mixture of root bark alkaloids. The most important are 3 α -(3,4,5-trimethoxybenzoyloxy)-nortropane, 3 α -(3,4,5-trimethoxybenzoyloxy)-tropan-6 β -ol, 3 α -(3,4,5-trimethoxybenzoyloxy)-nortropan-6 β -ol, 6 β -benzoyloxytropan-3 α ,7 β -diol, 6 β -benzoyloxy-3 α -(3,4,5-trimethoxycinnamoyloxy)-tropan-7 β -ol, and 7 β -acetoxy-6 β -benzoyloxy-3 α -(3,4,5-trimethoxycinnamoyloxy)-tropane. Other minor bases identified include 3 α -(3,4,5-trimethoxybenzoyloxy)-tropane, 3 α -(3,4,5-trimethoxycinnamoyloxy)-tropane, 3 α -phenylacetoxytropan-6 β -ol, 3 α -(3,4,5-trimethoxybenzoyloxy)tropan-6 β ,7 β -diol, 6 β -benzoyloxytropan-3 α -ol, and 6 β -benzoyloxy-3 α -(3,4,5-trimethoxycinnamoyloxy)tropane (35). The main alkaloid of *E. zambesiaceum* N. Robson is 3 α -(3',4',5'-trimethoxybenzoyl)-oxytropane, followed by three new alkaloids characterized as 6 β -benzoyloxytropan-3-one, 6-isovaleryloxytropan-3-ol and 3-(2-methyl butyryloxy)-6,7-diol (36). Other dimeric tropane alkaloids, although in very low yield (ca 0.003%), have been isolated from *E. moonii* Hochr.: moniine A (**40**) and moniine B (**41**) (37). They are diesters of tropine and nortropine with carboxylic dibasic acids presumably resulting from tropic acid biogenesis. With the exception of tropane esters, the tropane alkaloid spectrum of the genus *Erythroxyllum* is remarkably similar to that of the Solanaceae (12, 38). *Erythroxyllum australe* F. Muell (section *Coelocarpus*) contains tigloyl esters of meteloidine (**28**), 3 α -tigloyloxytropan-6 β -ol, 3 α -tigloyloxynortropan-6 β -ol and an unknown base which was tentatively identified as 7-hydroxy-6-tigloyloxynortropan-3-yl, 2-hydroxy-3-phenylpropionate. 2-

Hydroxy-3-phenyl-propionic acid, closely related to tropic acid and a component of the solanaceous alkaloid littorine (**4**), is a further link (besides meteloidine (**28**)) with the family *Solanaceae* (39). *Erythroxyllum zeylanicum* O. E. Schulz contains three tropane alkaloids: erythrozeylanines A [1R,3R,5S,6R-6-acetoxy-3-(3',4',5'-trimethoxybenzoyloxy)-tropane], B [cis-3 β -(cinnamoyloxy)-tropane], and C [cis-6 β -acetoxy-3 α -(cinnamoyloxy)-tropane], and others already found in other *Erythroxyllum* species (40).

Calystegines were identified in the genus *Erythroxyllum*, too. *Erythroxyllum novogranatense* Morris var. *novogranatense* contained 0.2% total calystegines in dry leaves. Forty-six *Erythroxyllum* herbarium species, consisting mostly of leaf tissue, were analysed for calystegines, and 38 were found positive. Calystegines A₃ (**6**) and B₂ (**7**) were the major calystegines in most species. The simultaneous occurrence of calystegines, cocaine (**1**), other alkaloids of a 3 α -hydroxy- or 3 β -hydroxytropane structure and nicotine (**26**), supports the concept of common biosynthetic steps of these alkaloids in *Erythroxyllum* (41). Finally, recently, pervilleine A (**42**), a novel tropane alkaloid, was extracted from *Erythroxyllum pervillei* Baill. (42).

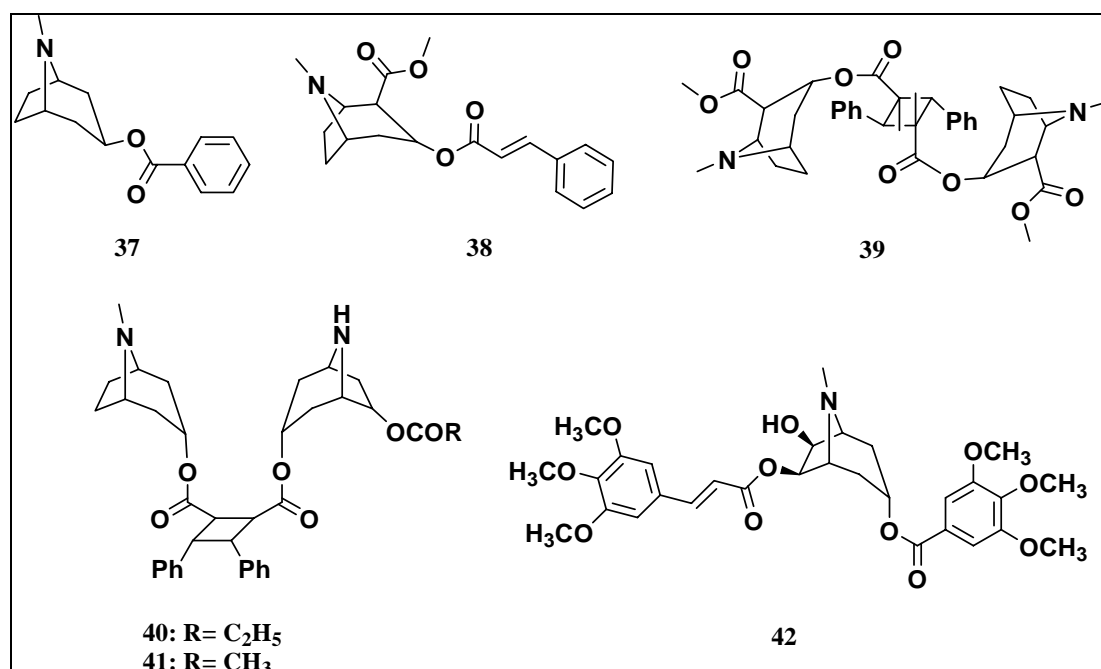


Figure 8. Tropacocaine (**37**); cinnamoyl-cocaine (**38**); α -truxilline (**39**); mooniine A (**40**); mooniine B (**41**); pervilleine A (**42**).

Proteaceae

Bellandena montana R. Br. afforded the first alkaloid from the *Proteaceae*, bellendine (**43**), an unusual tropane alkaloid (43). Similar alkaloids, such as darlingine (**44**), a pyranotropane base, were isolated from *Darlingia darlingiana* L. A. S. Johnson. (44) *D. ferruginea* J. F. Bailey contains darlingine (**44**), ferruginine (**45**), ferrugine (**46**) and 3 α -benzoyloxy-2 α -hydroxybenzyltropine (45). *Bellandena montana* R. Br. contains bellendine (**43**), isobellendine (**47**) and darlingine (**44**) (46). Alkaloids of a 2-benzyltropine type were obtained from genus *Knightia*, *K. deplanchei* Vieill ex Brongn. & Gris and *K. strobilina* R. Br. (47).

D-strobiline (**48**) is the main alkaloid of *K. strobilina* R. Br., together with 3 α -cinnamoyloxytropane-6 β -ol, 3 α -acetoxy-2 α -acetoxybenzyltropane (acetylknightinol) (**49**), 3 α -acetoxy-2 α -benzyltropane-6 β -ol (knightoline) (**50**), 6 β -benzoyloxytropane-3 α -ol (**51**), 2 α -hydroxybenzyl-3 α -acetoxytropane (knightinol) (**52**), dihydrostrobiline (**53**) (33), strobamine B (**54**), chalcostrobamine (**55**), strobolamine (**56**), knightalbinol (**57**) and knightolamine (**58**) (30). *Agastachys odorata* R. Br. contains 6 β -acetoxy-3 α -tigloyloxytropane and 3 α -(*p*-hydroxybenzoyloxy)-trop-6-ene, the first naturally occurring tropene base (45).

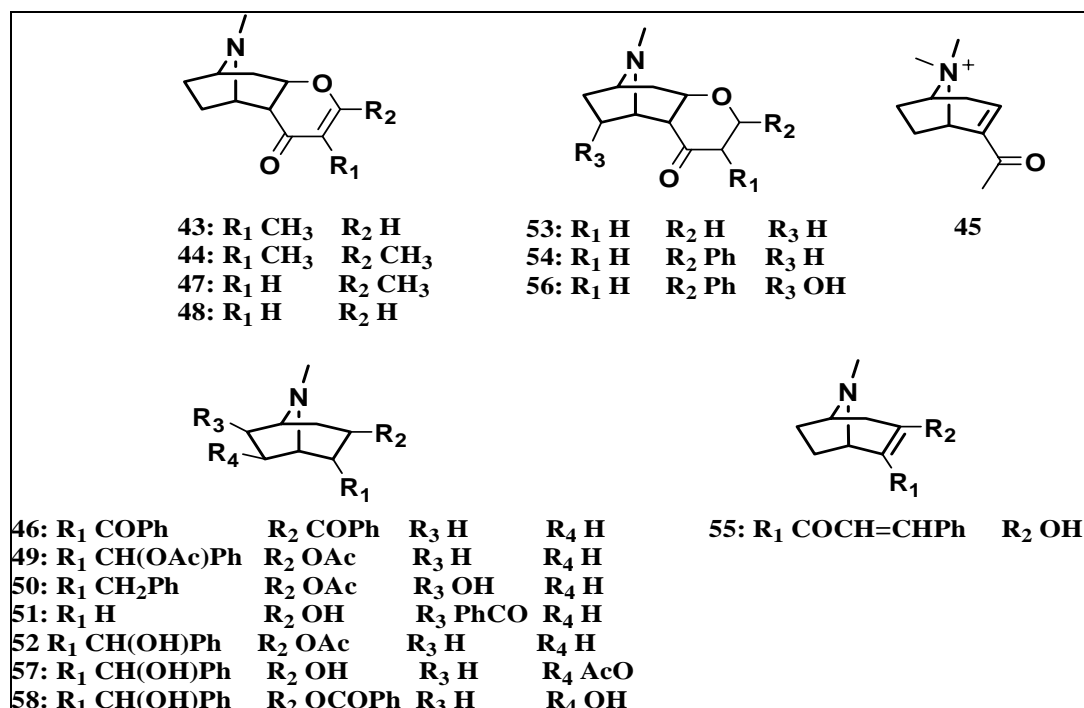


Figure 9. Bellendine (**43**); darlingine (**44**); ferruginine (**45**); ferrugine (**46**); isobellendine (**47**); strobiline (**48**); acetylknightinol (**49**); knightoline (**50**); 6 β -benzoyloxytropane-3 α -ol (**51**); knightinol (**52**); dihydrostrobiline (**53**); strobamine (**54**); chalcostrobamine (**55**); strobolamine (**56**); knightalbinol (**57**); knightolamine (**58**).

Euphorbiaceae

In this family, tropane alkaloids are isolated just from *Peripentadenia mearsii* (C.T.White) L. S. Sm. They are tropacocaine (**37**) (3 β -benzoyloxytropane), 3 α -acetoxy-6 β -hydroxytropane and 2 α -benzoyloxy-3 β -hydroxytropane (**48**).

Rhizophoraceae

This family contains some dithiolane esters: brugine (tropine 1,2-dithiolane-3-carboxylate) (**59**) was obtained from *Bruguiera sexangula* Poir., while *B. exaristata* Ding Hou contains other tropine esters of acetic, propionic, *n*-butyric, isobutyric, isovaleric and benzoic acids (**49**).

Brugine (**59**) has been also found in *Crossostylis* spp. (*C. biflora* Forst., *C. multiflora* Brongn. & Gris and *C. sebertii* Brongn. & Gris) along with tropine (**17**), 3 α -benzoyl-oxytropine and two esters of tropine, tropine cinnamate (**60**) and ferulate (**61**) (32). Of *Crossostylis* species, *C. Sebertii* (Pancher) Pierre Arbuste contains the tropane alkaloids tropan-3 α -yl cinnamate and tropan-3 α -yl benzoate; *C. multiflora* Brongn. & Gris contains tropan-3 α -yl-ferulate, 3 α -tropanol and brugine (**59**) and *C. biflora* J. R. Forst. & G. Forst. contains hygrine (**16**) and tropanone (50).

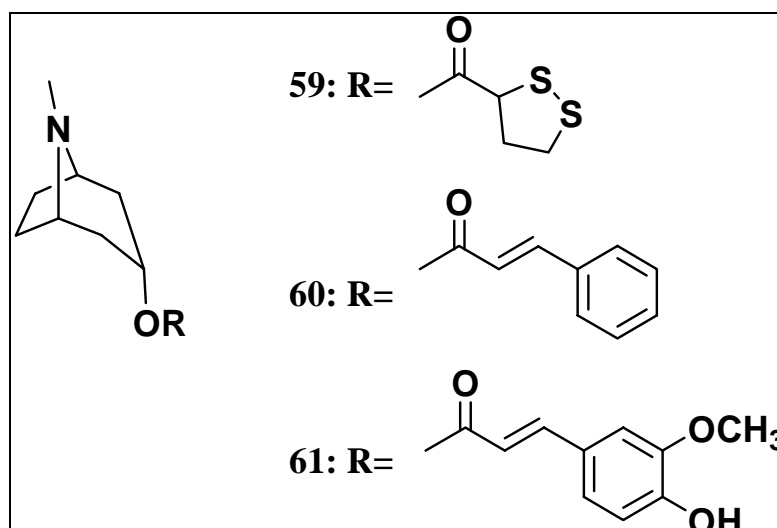


Figure 10. Brugine (**59**); tropine cinnamate (**60**); tropine ferulate (**61**).

Convolvulaceae

One-hundred twenty-nine convolvulaceous species belonging to 29 genera (all 12 tribes), revealed the occurrence of one to six polyhydroxy alkaloids of the nortropane type (calystegines) in 62 species belonging to 22 genera of all tribes except the unique parasitic *Cuscutaeae*. Specifically, the tetrahydroxylated alkaloids B₂ (**7**) and calystegine B₁ (**12**) are the most frequent compounds (90% and 68%) followed by the trihydroxynortropane A₃ (**6**) (38%) and the pentahydroxylated congener C₁ (**8**) (26%) (51). Calystegines A₅ (**19**), B₃ (**20**), and B₅ (**62**), isolated also in *Scopolia japonica* Maxim. (*Solanaceae*) (52), displayed a minor frequency, ranging from 16% to 20%. Calystegine A₆ (**63**), characterized by the lack of a hydroxy group at C-3, has been only discovered in *Stictocardia mojangensis* (51).

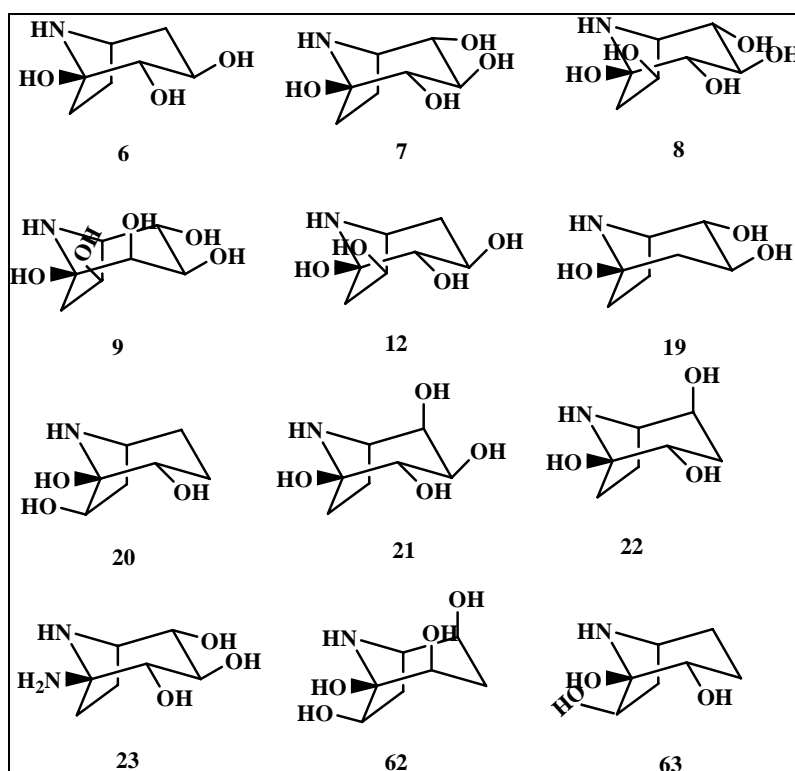


Figure 11. Calystegines: A₃ (6); B₂ (7); C₁ (8); C₂ (9); B₁ (12); A₅ (19); B₃ (20); B₄ (21); A₇ (22); N₁ (23); B₅ (62); A₆ (63).

Convolvine (64) and convolamine (65), a nortropan-3 α -yl veratrate and its N-methyl derivative, respectively, have been isolated from *Convolvulus lineatus* L. and *C. pseudocantabricus* Schrenk, together with closely related alkaloids having a veratroyl or a vanilloyl moiety as the acyl component. Convolvine (64) and convolamine (65) were extracted also from *C. krauseanus* Regel & Schmalh and *C. subhirsutus* L. Moreover, *C. krauseanus* Regel & Schmalh and *C. subhirsutus* L. contain a series of tropane esters with methoxy substituted benzoic acids of tropane alkaloids which include convolvine (3 α -veratrolyloxynortropane), convolidine (3 α -vanillolyloxynortropane) (53,54), confoline (3 α -veratroyl-*N*-formylnortropane) (55) and convolamine-*N*-oxide (56). Recently, from the aerial part of *C. subhirsutus* L. has been extracted a new tropane alkaloid, the confolidine ((\pm)3 α -vanillyl-*N*-formylnortropane), too. *Evolvulus sericeus* Sw. likewise contains convoline, convolamine (65) and convolidine. From *E. elliptilimba* Merr. & Chun and *E. hainanesis* Merr., *Erycibe* species, erycibelline (2 β ,7 β -dihydroxynortropane) (66) and baogongten C (67) have been isolated (57). Baotongten A, (2 β -hydroxy-6 β -acetoxynortropane) and baogongten B have been isolated from *Erycibe obtusifolia* Benth. The roots of *Convolvulus arvensis* L. and transformed root cultures of *Calystegia sepium* R. Br. contain calystegine (calystegin) B₁ (1 β ,2 α ,3 β ,6 α -tetrahydroxynortropane) (12), calystegine B₂ (1 β ,2 α ,3 β ,4 α -tetrahydroxynortropane) (7) and calystegine A₃ (1 β ,2 α ,3 β -trihydroxynortropane) (6) (58). From roots of *C. sepium* R. Br. also calystegines A₁, A₂ and A₄ (59) have been isolated. The roots of *C. siculus* L. contain consiculine (68), a tropan-3 α -ol esters, and merresectine B (69), while roots of *C. sabatius* Viv. ssp. *mauritanicus* contain consiculine (68) and consabatine (2'-deoxyconsiculine) (70). Finally from roots of *Convolvulus sabatius* Viv. ssp. *sabatius* are extracted 3 α -[1,4-dihydroxy-3-(3-methyl-but-2-enyl)-cyclohex-2-enecarboxy]tropane (4'-dihydroconsabatine) (71) (60).

Recently, the β -isomer of phyllalbine was found *C. cneorum* L., whose epigeal vegetative parts contain also the 3 β -vanillolyloxynortropane, named concneorine (72) (61).

From *Merremia Dissecta* Hallier F. and *M. guerichii* A. Meeuse were isolated four novel 3 α -acyloxytropanes, merresectines A–D: 3 α -(4-methoxybenzoyloxy)-nortropane (merresectine A) (73), 3 α -kurameroyloxytropane (merresectine B) (69), 3 α -nervogenoyloxytropane (merresectine C) (74) and 3 α -[4-(β -D-glucopyranosyloxy)-3-methoxy-5-(3-methyl-2-butenyl)benzoyloxy]tropane (β -D-glucoside of merresectine D (75) (76). Moreover, the novel 3 α ,6 β -di-(4-methoxybenzoyloxy)-tropane (merredissine) (77) has been isolated from *M. dissecta* Hallier F. The 4- β -D-glucoside (78) of merresectine E (79) has been identified in six *Merremia* species (60).

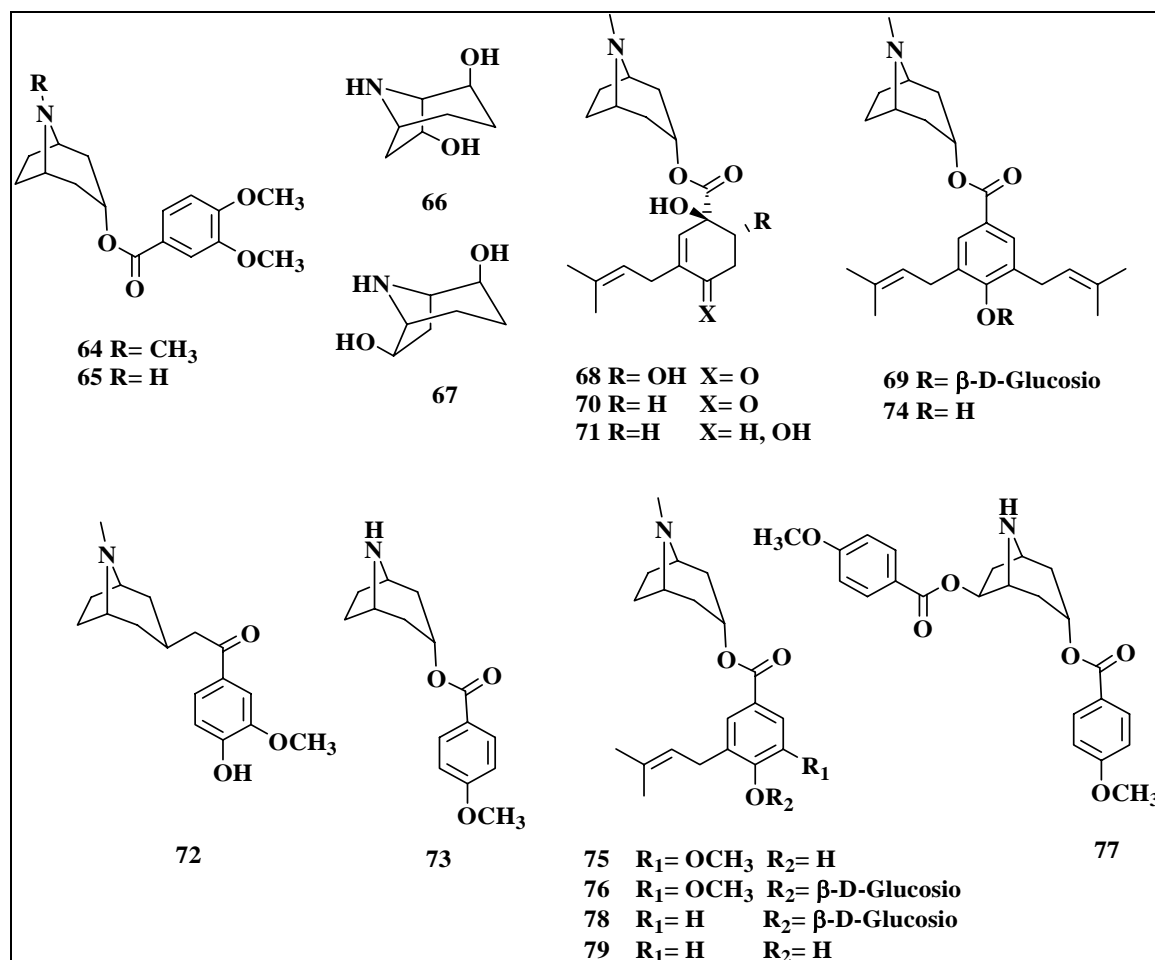


Figure 12. *Convolvine* (64); *convolamine* (65); *erycibelline* (66); *baogongten C* (67); *consiculine* (68); *merresectine B* (69); *consabatine* (70); *4'-dihydroconsabatine* (71); *conconeorine* (72); *merresectine A* (73); *merresectine C* (74); *merresectine D* (75); *merresectine D* β -D-glucoside (76); *merredissine* (77); *merresectine E* β -D-glucoside (78); *merresectine E* (79).

Cruciferae

Brassicaceae or Cruciferae contain a novel group of hydroxylated nortropane alkaloids, some calystegines, above all calystegine A and the 3-hydroxybenzoate ester of tropine and cochlearine (80). The last one is the first tropane alkaloid identified in *Cochlearia officinalis* L. and *Cochlearia arctica* Schlecht (60).

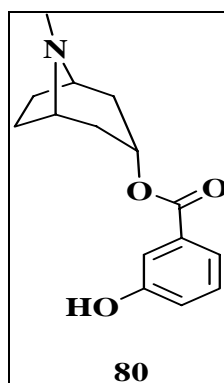


Figure 13. Cochlearine (80).

In most genus *Cochlearia* tropine (17), the alcohol moiety of cochlearine (80) and pseudotropine (24) were found in minor concentration.

Calystegines were found in high concentrations in *Cochlearia officinalis* L. tissues. However, it contains calystegine A₅ (19) as the major compound, while calystegines A₃ (6) and B₂ (7) dominate in *Solanaceae*, calystegines B₁ (12) and B₂ (7) in *Convolvulaceae* and calystegines A₃ (6) and B₂ (7) in *Erythroxylaceae* (60).

Moraceae

Among this family, *Morus alba* L. is important to be remembered for its contents of tropane alkaloids. Specifically, from its root bark the penthydroxylated nortropane calystegine C₁ (8) (61), was isolated, while fruits contain calystegines A₈ and A₉ (62).

Pharmacology of Tropane Alkaloids

Cocaine (1), first isolated from the leaves of Peruvian *Erythroxylum coca* Lam. in 1860, shows local anaesthetic properties, stimulates the Central Nervous System and improves physical endurance. In fact, it is both a stimulant of the Central Nervous System and an appetite suppressant. Specifically, it is a dopamine reuptake inhibitor. It gives a feeling to what has been described as a euphoric sense of happiness and increased energy. It is most often used recreationally for this effect. Because of the way it affects the mesolimbic reward pathway, cocaine (1) is addictive. Nevertheless, cocaine (1) is still used in medicine as a topical anesthetic, even in children, specifically in eye, nose and throat surgery (63).

Atropine (2) lowers the “rest and digest” activity of muscle and glands regulated by Parasympathetic Nervous System. For this reason, it is used topically as a cycloplegic, to temporarily paralyze the accommodation reflex, and as a midriatic, to dilate the pupils. In fact, it blocks the contraction of the circular papillary sphincter muscle which is normally stimulated by acetylcholine release, thereby allowing the radial papillary dilator muscle to contract and dilate pupil. Injections of atropine are used in the treatment of bradycardia, asystole and pulseless electrical activity (PEA) in cardiac arrest. These effects take place because the main action of the vagus nerve of the parasympathetic system on the heart is to slow it down. Atropine (2) blocks such action and therefore may speed up the heart rate.

It is also useful in treating first degree of heart block, second degree of heart block Mobitz Type 1 and third degree heart block, too, with a high Purkinje or AV-nodal escape rhythm. Moreover, this compound acting on Parasympathetic Nervous System, inhibits salivary, sweat and mucus gland. This can be useful in treating hyperhidrosis and can prevent the death rattle of dying patients. Finally, by blocking the action of acetylcholine at muscarinic receptors, atropine also serves as an antidote for poisoning by organophosphate insecticides and nerve gases (64). Finally, hyoscyamine, the laevogyrate isomer of atropine (2), is used to provide symptomatic relief to various gastrointestinal disorders including spasm, peptic ulcers, irritable bowel syndrome, pancreatitis, colic and cystitis. It is also been used to control some of the symptoms of Parkinson's disease as well as for the control of respiratory secretions in end of life care (65).

Scopolamine (3), also known as hyoscyne, is used in medicine (in form of scopolamine hydrobromide,) for its depressant activity on the Central Nervous System, though it can cause delirium in the presence of pain, miosis and cycloplegia. Combined with morphine, it produces amnesia and a tranquilized state known as twilight sleep. It is used in ophthalmology to deliberately cause cycloplegia and mydriasis for diagnostic purposes and in the treatment of iridocyclitis. In otolaryngology, it has been used to dry the upper airway prior to use medical instrumentation on the airway. Scopolamine (3) is also an antiemetic, an antivertigo and an antispasmodic and can be used in the pre-anesthetic sedation, as an antiarrhythmic during anesthesia and for the prevention of motion sickness. Some times ago it was also used in obstetrics, but now it is considered dangerous in this field (66). In October 2006 researchers at US National Institute of Mental Health found that scopolamine (3) reduces symptoms of depression within a few days and the improvement lasted for at least a week after switching to a placebo (67).

Nicotine (26) effects on the cardiovascular system are mediated by sympathetic neural stimulation associated with an increase in the levels of circulating catecholamines. Nicotine (26) causes sympathetic stimulation through central and peripheral mechanisms. Central Nervous System mediated mechanisms include activation of peripheral chemoreceptors, particularly the carotid chemoreceptor, and direct effects on the brain stem and spinal cord. Peripheral mechanisms include release of catecholamines from the adrenal glands and vascular nerve endings (68). These effects of nicotine (26) result in an acute increase in heart rate and blood pressure when it is delivered via cigarette smoking, chewing gum, nasal spray or intravenous infusion. Nicotine (26) differentially affects blood flow to different organs, causing vasoconstriction in some vascular beds (eg. skin) and vasodilatation in others (eg. skeletal muscle). Cutaneous vasoconstriction results in a decrease in the fingertip temperature (69). Nicotine (26) induces vasoconstriction in coronary arteries, as evidenced by a lack of increased blood flow in response to increased oxygen demand and by direct observation, specifically in atherosclerotic arteries. Coronary vasoconstriction appears to be mediated by catecholamines and can be abolished by the α -adrenergic blocker phentolamine (70).

Calystegines (**Fig.11**) are selective glycosidase inhibitors and could be applied as pharmaceutical compounds for diabetic patients (71).

Tigloidine (13) seems to be a substitute for atropine (2) (72), so it could be a potential anti-Parkinson agent (73); moreover, it could be used in the symptomatic treatment of spastic paraplegia (74) and in extrapyramidal syndromes (Huntington's chorea) (75).

Phyllalbine acts as central and peripheral sympathomimetic (76).

Finally, because of its structural similarity with the well known (-)-hyoscyamine and (-)-cocaine (**1**), pervilleine A (**42**) shows cholinergic and adrenergic activities (77). Moreover, recent studies attest that pervilleine A (**42**) is an inhibitor of P-glycoprotein, a protein of drug efflux associated with a poor response to cancer chemotherapy, and should be further evaluated for clinical utility (42).

Discussion

Looking at the chemical structure of the tropane alkaloids, it is possible to note that all compounds are characterized by the presence of the tropane core and it has been proposed a possible common biosynthetic pathway, starting from the amino acid L-ornitine. After several biosynthetic steps, this compound is converted to hygrine, from which α -tropine, β -tropine, ecgonine and cuscohygrine origin (Fig. 14) and act as the bases for the biosynthesis of all the tropane alkaloids known.

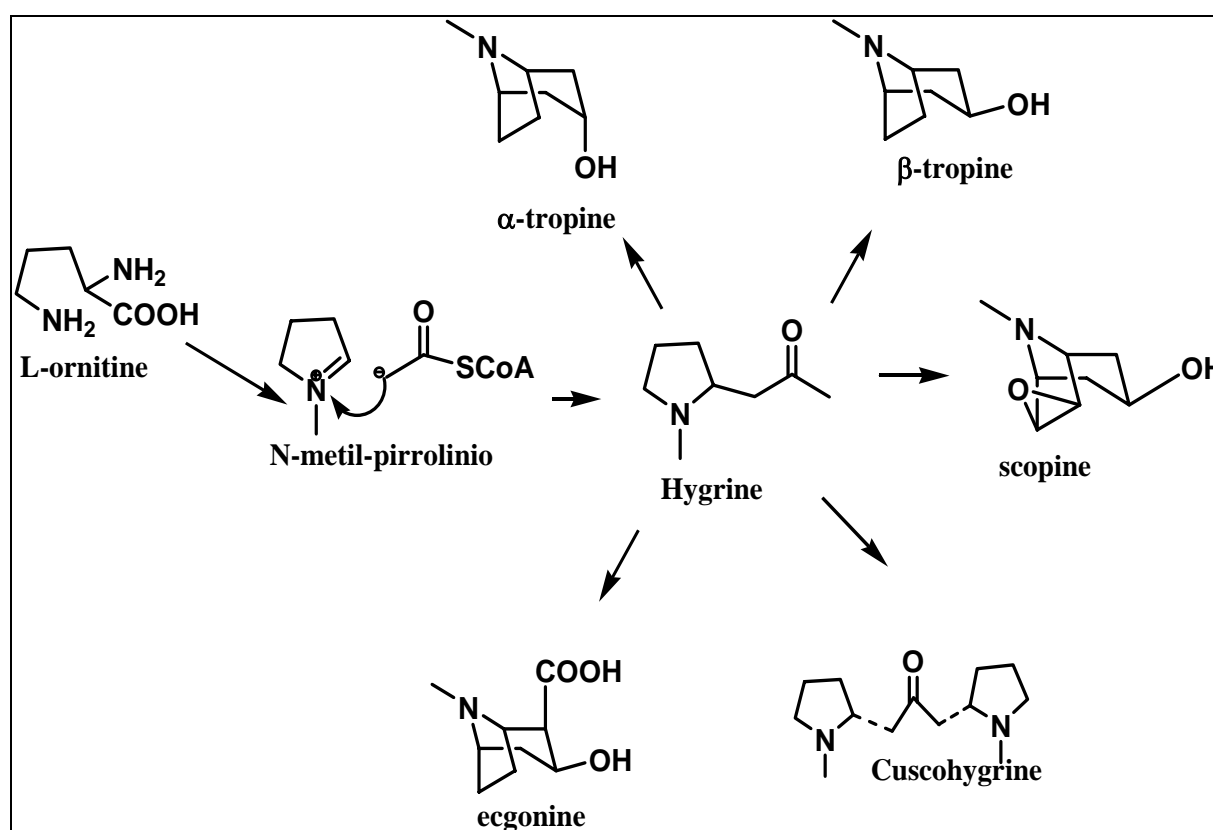


Figure 14. Biosynthesis of tropane alkaloids from L-ornitine.

Conclusion

Chemical and biological resemblances among different families of higher plants containing tropane alkaloids support the modern botanic classification of Solanaceae and Convolvulaceae in the order of Solanales and of Euphorbiaceae and Rhizophoraceae in the group of Malpighiales (**Fig.15**) (78).

Now, in fact, Solanaceae and Convolvulaceae are considered “sisters” because of their morphology and their plastidial DNA (79-84). In a similar way, Euphorbiaceae and Rhizophoraceae are now considered members of Malpighiales (83-86).

In addition, the recent botanic classification considers Brassicaceae (Brassicales), Moraceae (Rosales) and Malpighiales members of the same larger group Roside (Fig.15) (78). This is to support, again, that biosynthesis of similar secondary metabolites can be related to a common phylogenetic origin.

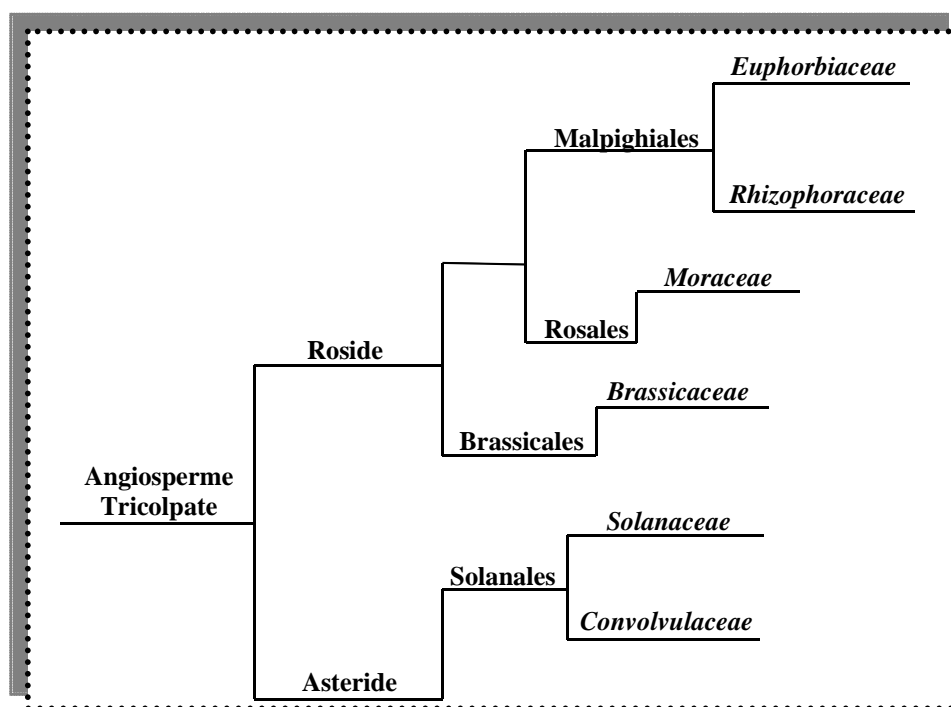


Figure 15. Simple cladogramme of the families of higher plant containing tropane alkaloids.

References

1. Griffin W.J. & Lin G.D. **Chemotaxonomy and geographical distribution of tropane alkaloids.** *Phytochemistry*. 2000; 53: 623-637.
2. De Feo, V. **The ritual use of *Brugmansia* species in traditional Andean medicine in Northern Peru.** *Economic Botany*. 2004; 58.
3. Nash R.J., Rothschild M., Porter E.A., Watson A., Waigh R. & Waterman P.G. **Calystegines in *Solanum* and *Datura* species and the death's-head hawk-moth (*Acherontia atropus*).** *Phytochemistry*. 1993; 34(5): 1281-3.
4. Evans W.C., Ghani A. & Woolley V.A. **Distribution of littorine and other alkaloids in the roots of *Datura* species.** *Phytochemistry*. 1972; 11: 470.
5. Evans, W.C. & Ramsey K.P.A. **Alkaloids of the *Solanaceae* tribe *Anthocercideae*.** *J. Pharm. Pharmacol.* 1979; 31: 9P.
6. Phillipson J.D. & Handa S.S. **Hyoscyamine N-oxide in *Atropa belladonna*.** *Phytochemistry* (Elsevier). 1976; 15(5): 605-8.

7. Phillipson J.D., Handa S.S. & El-Dabbas S.W. **N-oxides of morphine, codeine and thebaine and their occurrence in *Papaver* species.** *Phytochemistry* (Elsevier). 1976; 15(8): 1297-301.
8. Sahai M. & Ray A.B. **Secotropane alkaloids of *Physalis peruviana*.** *Journal of Organic Chemistry*. 1980; 45: 3265.
9. Kubwabo C., Rollman B. & Tillequin B. **Analysis of alkaloids from *Physalis peruviana* by capillary GC, capillary GC-MS, and GC-FTIR.** *Planta Medica*. 1993; 59: 161.
10. Asano N., Kato A., Miyauchi M., Kizu H., Tomimori T., Matsui K., Nash R.J. & Molyneux R.J. **Specific galactosidase inhibitors, N-methylcalystegines. Structure/activity relationships of calystegines from *Lycium chinense*.** *European Journal of Biochemistry*. 1997; 248: 296.
11. El-Imam Y.M.A., Evans W.C. & Plowman T. **Alkaloids of the genus *Erythroxylum*. Part 4. Alkaloids of some South American *Erythroxylum* species.** *Phytochemistry* (Elsevier). 1985; 24(10): 2285-9.
12. Evans W.C. **Tropane alkaloids of the *Solanaceae*.** *Linnean Soc. Symp.* 1979; 7: 241-54.
13. Jackson B.P. & Berry M.I. **Hydroxytropane tiglates in the roots of *Mandragora* species.** *Phytochemistry*. 1973; 12: 1165.
14. Khanna K.L., Schwarting A.E., Rother A. & Bobbitt, J.M. **Occurrence of tropine and pseudotropine in *Withania somnifera*.** *Lloydia*. 1961; 23: 179-81.
15. Hill K.L., Bottomley W. & Mortimer P.I. **Variation in the main alkaloids of *Duboisia myoporoides* and *Duboisia leichhardtii*. IV. Interspecific hybrids.** *Australian Journal of Applied Sciences*. 1954; 5: 258.
16. Mortimer P.I. & Wilkinson S. **The occurrence of nicotine, anabasine, and isopelletierine in *Duboisia myoporoides*.** *Journal of the Chemical Society*. 1957; 3967-70.
17. Hills K. Loftus, Bottomley W., & Mortimer P. I. **Variation in the main alkaloids of *Duboisia myoporoides* and *Duboisia leichhardtii*. III. *Duboisia leichhardtii*.** *Australian Journal of Applied Science*. 1954; 5: 276-82.
18. Cannon J.R., Joshi K.R., Meehan G.V. & Williams J. **Tropane alkaloids from three western Australian *Anthocercis* species.** *Australian Journal of Chemistry*. 1969; 22(1): 221-7.
19. Evans W.C. & Treagust P.G. **Alkaloids of *Datura pruinosa*.** *Phytochemistry* (Elsevier). 1973; 12(8): 2077-8.
20. Evans W.C. & Ramsey K.P.A. **Tropane alkaloids from *Anthocercis* and *Anthotroche*.** *Phytochemistry* (Elsevier). 1981; 20(3): 497-9.
21. Romeike A. **Presence of tropinone in *Nicandra* roots.** *Naturwissenschaften*. 1966; 53(3): 82.
22. San Martin A.S., Roviroso J., Gambaro V. & Castillo M. **Tropane alkaloids from *Schizanthus hookeri*.** *Phytochemistry* (Elsevier). 1980; 19(9): 2007-8.
23. Muñoz O., Piovano M., Garbarino J., Hellwing V. & Breitmaier E. **Tropane alkaloids from *Schizanthus litoralis*.** *Phytochemistry*. 1996; 43(3): 709-713.
24. Gambaro V., Labbe C. & Castillo M. **Angeloyl-, tigloyl-, and seneciolyloxytropane alkaloids from *Schizanthus hookeri*.** *Phytochemistry* (Elsevier). 1983; 22(8): 1838-9.
25. San Martin A. S., Labbe C., Muñoz O., Castillo M., Reina M., De la Fuente B. & Gonzalez A. **Tropane alkaloids from *Schizanthus grahamii*.** *Phytochemistry*. 1987; 26(3): 819-22.
26. Rivier L. **Analysis of alkaloids in leaves of cultivated *Erythroxylum* and characterization of alkaline substances used during coca chewing.** *Journal of Ethnopharmacology* 1981; 3(2-3): 313-35.
27. Moore J.M., Cooper D.A., Lurie I.S., Theodore C.M., Carr S., Harper C. & Yeh J. **Capillary gas chromatographic-electron capture detection of coca leaf-related impurities in illicit cocaine: 2,4-diphenylcyclobutane-1,3-dicarboxylic acids, 1,4-diphenylcyclobutane-2,3-dicarboxylic acids and their alkaloidal precursors, the truxillines.** *Journal of Chromatography*. 1987; 410(2): 297-318.

28. Moore J.M., Hayes P.A., Cooper D.A., Casale J.F. & Lydon J. **1-Hydroxytropacocaine: an abundant alkaloid of *Erythroxylum novogranatense* var. *novogranatense* and var. *truxillense*.** *Phytochemistry*. 1994; 36(2): 357-60.
29. Al-Said M.S., Evans W.C. & Grout R.J. **Alkaloids of the genus *Erythroxylum*. Part 5. *E. hypericifolium* Lam. root-bark.** *Journal of the Chemical Society*. 1986; 6: 957-9.
30. Lounasmaa M., Pusset J. & Sevenet T. **The New Caledonian plants. Part 57. 2-Benzyl pseudotropanes and dihydropyranotropanes, alkaloids of *Knightsia strobilina*.** *Phytochemistry* (Elsevier). 1980; 19(5): 953-5.
31. El-Imam Y.M.A., Evans W.C., Grout R.J. **Alkaloids of the genus *Erythroxylum*. Part 8. Alkaloids of *Erythroxylum cuneatum*, *E. ecarinatum* and *E. australe*.** *Phytochemistry*. (1988), 27(7), 2181-4.
32. Gnecco Medina D.H., Pusset J., Pusset M. & Husson H.P. **Tropane alkaloids from *Crossostylis* sp.** *Journal of Natural Products*. 1983; 46(3): 398-400.
33. Al-Said M.S., Evans W.C. & Grout R.J. **Alkaloids of the genus *Erythroxylum*. Part 10. Alkaloids of *Erythroxylum hypericifolium* leaves.** *Phytochemistry*. 1989; 28(11): 3211-15.
34. Al-Said M.S., Evans W.C. & Grout R.J. **Alkaloids of the genus *Erythroxylum*. Part 9. Alkaloids of *Erythroxylum hypericifolium* stem bark.** *Phytochemistry*. 1989; 28(2): 671-3.
35. El-Imam Y.M.A., Evans W.C., Grout R.J. & Ramsey K.P.A. **Alkaloids of the genus *Erythroxylum*. Part 7. Alkaloids of *Erythroxylum zambesiacum* root-bark.** *Phytochemistry*. 1987; 26(8): 2385-9.
36. Christen P., Roberts M.F., Phillipson J.D. & Evans W.C. **Alkaloids of the genus *Erythroxylum*. Part 11. Alkaloids of *Erythroxylum zambesiacum* stem-bark.** *Phytochemistry*. 1993; 34(4): 1147-51.
37. Rahman A.U., Khattak K.F., Nighat F., Shabbir M., Hemalal K.D. & Tillekeratne L.M. **Dimeric tropane alkaloids from *Erythroxylum moonii*.** *Phytochemistry*. 1998; 48(2): 377-383.
38. Evans W.C. & Ramsey K.P.A. **Alkaloids of the *Solanaceae* tribe *Anthocercideae*.** *Phytochemistry* (Elsevier). 1983; 22(10): 2219-25.
39. Griffin W.J. **A phytochemical investigation of *Erythroxylum australe* F. Muell.** *Australian Journal of Chemistry*. 1978; 31(5): 1161-5.
40. Bringmann G., Günther C., Mühlbacher J., Lalith M.D., Gunathilake P. & Wickramasinghe A. **Tropane alkaloids from *Erythroxylum zeylanicum* O.E. Schulz (*Erythroxylaceae*).** *Phytochemistry*. 2000; 53(3): 409-416.
41. Brock A., Bieri S., Christen P. & Dräger B. **Calystegines in wild and cultivated *Erythroxylum* species.** *Phytochemistry*. 2005; 66(11): 1231-40.
42. Mi Q., Cui B., Silva G.L., Lantvit D., Lim E., Chai H., You M., Hollingshead M.G., Mayo J. G., Kinghorn A.D. & Pezzuto J.M. **Pervilleine A, a novel tropane alkaloid that reverses the multidrug-resistance phenotype.** *Cancer Research*. 2001; 61(10): 4030-4037.
43. Bick I.R.C., Bremer J.B. & Gillard J.W. **Methyl(p-hydroxybenzoyl)acetate and an alkaloid, bellendine, from *Bellendena montana*.** *Phytochemistry* (Elsevier). 1971; 10(2): 475-7.
44. Ralph I., Bick C., Gillard J.W. & Leow H.M. **Alkaloids of *Darlingia darlingiana*.** *Australian Journal of Chemistry*. 1979; 32(11): 2523-36.
45. Ralph I., Bick C., Gillard J.W. & Leow H.M. **Alkaloids of *Darlingia ferruginea*.** *Australian Journal of Chemistry*. (1979), 32(11), 2537-43.
46. Bick I.R.C., Gillard J.W. & Huck-Meng L. **Alkaloids of *Bellendena montana*.** *Australian Journal of Chemistry*. 1979; 32(8): 1827-40.
47. Lounasmaa M., Wovkulich P.M. & Wenkert E. **Carbon-13 nuclear magnetic resonance spectroscopy of naturally occurring substances. XXXVI. Structures of some *Knightsia deplanchei* alkaloids.** *Journal of Organic Chemistry*. 1975; 40(25): 694-7.

48. Johns S.R., Lamberton J.A. & Sioumis A.A. **New tropane alkaloids, (+)-(3R, 6R)-3 α -acetoxy-6 β -hydroxytropine and (+)-2 α -benzoyloxy-3 β -hydroxynortropine, from *Peripentadenia mearsii* (Euphorbiaceae).** *Australian Journal of Chemistry*. 1971; 24(11): 2399-403.
49. Loder J.W. & Russel G.B. **Tropine 1,2-dithiolane-3-carboxylate, a new alkaloid from *Bruguiera sexangula*.** *Tetrahedron Letters*. 1966; 51: 6327-9.
50. Gnecco Medina D.H., Pusset M., Pusset J. & Husson H.P. **Tropine alkaloids from *Crossostylis* sp.** *Journal of Natural Products*. 1983; 46(3): 398-400.
51. Schimming T., Jenett-Siems K., Mann P., Tofern-Reblin B., Milson J., Johnson R.J., Deroin T., Austin D.F. & Eich E. **Calystegines as chemotaxonomic markers in the *Convolvulaceae*.** *Phytochemistry* (Elsevier). 2005; 66(4): 469-480.
52. Asano N., Kato A., Kizu H., Matsui K., Watson A.A. & Nash R.J. **Calystegine B₄, a novel trehalase inhibitor from *Scopolia japonica*.** *Carbohydrate Research*. 1996; 293(2): 195-204.
53. Aripova S.F., Malikov V.M. & Yunusov S. Yu. **Convolidine-new alkaloid from *Convolvulus krauseanus*.** *Khimiya Prirodnykh Soedinenii*. 1977; 2: 290-1.
54. Aripova S.F., Sharova E.G. & Yunusov S. Yu. **Convolute-a new alkaloid from *Convolvulus krauseanus*.** *Khimiya Prirodnykh Soedinenii*. 1983; 6: 749-51.
55. Sharova E.G., Aripova S.F. & Yunusov S. Yu. **Alkaloids of *Convolvulus subhirsutus*.** *Khimiya Prirodnykh Soedinenii*. 1980; 5: 672-6.
56. Aripova S.F. **N-Oxide of convolamine from *Convolvulus krauseanus*.** *Khimiya Prirodnykh Soedinenii*. 1985; 2: 275.
57. Lu Y., Yao T. & Chen Z. **Studies on the constituents of *Erycibe elliptimba*.** *Yao xue xue bao = Acta pharmaceutica Sinica*. 1986; 21(11): 829-35.
58. Goldmann A., Milat M.L., Ducrot P.H., Lallemand J., Maille A., Charpin I. & Tepfer D.T. **Tropine derivatives from *Calystegia sepium*.** *Phytochemistry*. 1990; 29(7): 2125-7.
59. Brock A., Herzfeld T., Paschke R., Koch M. & Dräger B. **Brassicaceae contain nortropine alkaloids.** *Phytochemistry* (Elsevier). 2006; 67(18): 2050-2057.
60. Jenett-Siems K., Weigl R., Böhm A., Mann P., Tofern-Reblin B., Ghomian A., Kaloga M., Siems K., Witte L., Hilker M., Müller F. & Eich E. **Chemotaxonomy of the pantropical genus *Merremia* (Convolvulaceae) based on the distribution of tropane alkaloids.** *Phytochemistry* (Elsevier). 2005; 66(12): 1448-1464.
61. Asano N., Oseki K., Tomioka E., Kizu H. & Matsui K. **N-Containing sugars from *Morus alba* and their glycosidase inhibitory activities.** *Carbohydrate Research*. 1994; 259(2): 243-55.
62. Kusano G., Orihara S., Tsukamoto D., Shibano M., Coskun M., Guvenc A. & Erdurak C.S. **Five new nortropine alkaloids and six new amino acids from the fruit of *Morus alba* Linne growing in Turkey.** *Chemical & Pharmaceutical Bulletin*. 2002; 50(2): 185-192.
63. Cabral G.A. **Drugs of abuse, immune modulation, and AIDS.** *J Neuroimmune Pharmacol*. 2006; 1(3): 280-95.
64. Kaluski E., Blatt A., Leitman M., Krakover R., Vered Z. & Cotter G. **Atropine-facilitated electrical cardioversion of persistent atrial fibrillation.** *ACC Current Journal Review*. 2004; 2: 13-55.
65. Orhan I., Naz Q., Kartal M., Tosun F., Sener B. & Choudhary M.I. **In vitro anticholinesterase activity of various alkaloids.** *Naturforsch*. 2007; 62(9-10): 684-8.
66. Furey M.L. & Drevets W.C. **Antidepressant efficacy of the antimuscarinic drug scopolamine: a randomized, placebo-controlled clinical trial.** *Archives of General Psychiatry*. 2006; 63(10): 1121-1129.
67. Blokland A. **Scopolamine-induced deficits in cognitive performance: a review of animal studies.** Faculty of Psychology, Brain and Behavior, Institute Maastricht University, the Nederland.
68. **Physical and pharmacological effects of nicotine.** Royal College of Physicians. 2007.

69. Gourlay S.G. & Benowitz N.L. **Arteriovenous differences in plasma concentration of nicotine and catecholamines and related cardiovascular effects after smoking, nicotine nasal spray, and intravenous nicotine.** *Clin. Pharmacol. Ther.* 1997; 62: 453-63.
70. Kaijser L. & Berglund B. **Effect of nicotine on coronary blood-flow in man.** *Clin. Physiol.* 1985; 5: 541-52
71. **Chemistry and biology of calystegines.** *Natural Product Reports.* 2004. Institute of Pharmaceutical Biology, Faculty of Pharmacy, Martin-Luther-University Halle-Wittenberg, Germany.
72. Trautner E.M. & Noack C.H. **Tigloidine as a substitute for atropine in the treatment of Parkinsonism.** *Med. J. Aust.* 1951; 1(21): 751-4.
73. Sanghvi I., Bindler E. & Gershon S. **Pharmacology of a potential anti-Parkinson agent: tigloidine.** *Eur. J. Pharmacol.* 1968; 4(3): 246-53.
74. O'Rourke F.J., Gershon S., Trautner E.M. & Shaw F.H. **The use of tigloidine in the symptomatic treatment of spastic paraplegia.** *Med. J. Aust.* 1960; 47(1): 73-7.
75. Trautner E.M. & Gershon S. **The effect of tigloidine on extrapyramidal syndromes (Huntington's chorea).** *Australas Ann Med.* 1958; 7(4): 286-91.
76. Quevauviller A., Foussard-Blanpin O. & Coignard D. **An alkaloid of *Phyllanthus discoides* (Euphorbiaceae) phyllalbine, a central and peripheral sympathomimetic.** *Therapie.* 1965; 20(4): 1033-41.
77. Chin Y.W., Kinghorn A.D. & Patil P.N. **Evaluation of the cholinergic and adrenergic effects of two tropane alkaloids from *Erythroxylum pervillei*.** *Phytother Res.* 2007; 21(10): 1002-5.
78. Judd W.J., Campbell C.S., Kellogg E.A. & Stevens P.F. **Plant systematics. A phylogenetic approach.** *Sinauer Associates, Inc. Publishers. Sunderland, Massachusetts U.S.A.*
79. Downie S.R. & Palmer J.D. **Restriction site mapping of chloroplast DNA inverted repeat: a molecular phylogeny of the Asteridae.** *Ann. Missouri Bot. Gard.* 1992; 79: 266-238.
80. Olmstead R.G., Michaels H.J., Scott K.M. & Palmer J.D. **Monophyly on the Asteridae and identification from DNA sequences of *rbcL*.** *Ann. Missouri Bot. Gard.* 1992; 79: 249-265.
81. Olmstead R.G., Bremer B., Scott K.M. & Palmer J.D. **A parsimony analysis of the Asteridae sensu lato based on *rbcL* sequences.** *Ann. Missouri Bot. Gard.* 1993; 80: 700-722.
82. Taktajan A. **Diversity and classification of flowering plants.** *Columbia University Press, New York.* 1997.
83. Thorne R.F. **Classification and geography of the flowering plants.** *Bot. Rev.* 1992; 58: 225-348.
84. Cronquist A. **An integrated system of classification of flowering plants.** *Columbia University Press, New York.* 1981.
85. Chase M.W., Duvall M.R., Hills H.G., Conran J.G., Cox A.V., Eguiarte L.E., Hartwell J., Fay M.F., Caddick L.R., Cameron K.M. & Hoot S. **Molecular systematics of Lilianae. In Monocotyledons: systematics and evolution.** *Royal Botanic Gardens, Kew.* 1995; 109-137.
86. Soltis D.E., Soltis P.S., Mort M.E., Chase M.W., Sarolaine, V, Hoot S.B. & Morton C.M. **Interfering complex phylogenies usins parsimony: an empirical approach using three large DNA data set for angiosperms.** *Syst. Biol.* 1998; 47: 32-42.