Antifungal Activity of the Ethanolic Extract of Nitella hyalina

Pokharkar Raghunath D, *Funde Prasad E, Pingale Shirish S, Pawar Kiran.S, Kalhapure Vaibhav M, Gavshete Sunil L.

Department of Chemistry, Arts, Commerce & Science College, Narayangaon, Pune, Maharashtra, India P.G.S.P. Charitable Foundation Funde Takali Ahmednagar Maharashtra India. Ravikiran and Tribal Woman Development Institution Maharashtra India

Summary

The Antifungal activity of the ethanol extract of *Nitella hyalina* was evaluated at two different concentrations by the diffusion method. The ethanol extract of the *Nitella hyalina* shows antifungal activity at varied levels in Yeasts (*Candida albicans, C. parapsilosis*) and Dermatophytes (*Trichophyton rubru, T. mentagrophytes*). The yeast *Candida albicans* was found to be more active and *C. parapsilosis* was found to be less active in inhibition zone. **Dermatophytes** *Trichophyton rubrum and T. mentagrophytes* also shows Antifungal activity. *T. mentagrophytes* was found to be more active than reference inhibition zone.

Keywords Nitella hyalina, Antifungal activity, extract.

Correspondence to: Funde Prasad, E "P.G.S.P. Charitable Foundation," Ahmednagar, Maharashtra At/Po: Funde Takali ,Tal: Pathardi, Di:A-Nagar, 414102 E-mail:-pef@rediffmail.com

Introduction

Marine medicine represents one of the most important fields of marine medicine all over the world. To promote the proper use of marine medicine and to determine their potential as sources for new drugs, it is essential to study medicinal bacteria. Over the past 20 years, there has been an increased interest in the investigation of natural materials as sources of new antifungal agents. Different extracts from marine medicinal bacteria have been tested to identify the source of the therapeutic effects. As a result some natural products have been approved as new antifungal drugs, but there is still an urgent need to identify novel substances that are active towards pathogens with high resistance. Recently, multiple drug resistance has developed due to indiscriminate use of commercial antifungal drugs commonly used in the treatment of fungal infectious diseases making it a global growing-problem. Isolation of antifungal agents less susceptible to regular antifungal and recovery of increasing resistant isolates during antifungal therapy is rising throughout the world which highlights the need for new principles. Natural products, a new source of antifungal agents shows possibly novel mechanisms of action.

Pharmacologyonline 3: 34-39 (2008) Newsletter Pokharkar et al.

Contrary to the synthetic drugs, antifungal of marine bacteria origin are not associated with many side effects and have an enormous therapeutic potential to heal many infectious diseases. For example, vincristine (an antitumor drug), digitalis (a heart regulator), and ephedrine (a bronchodilator used to decrease respiratory congestion) were all originally discovered through research on marine bacteria. This paper reports to study the antifungal activity of Indian marine bacteria, *Nitella hyalina* against an array of human pathogens. The ethanol extract of the *Nitella hyalina* was evaluated for the potential antifungal property. The selection of this plant for evaluation was based on its traditional usage. The marine bacteria posseses high content of active compound by HPLC methods and the Antifungal activity of the crude ethanol extract of *Nitella hyalina* was evaluated at two different concentrations by the diffusion method.

Materials and Methods

Marine bacteria

Nitella hyalina marine bacteria were collected in marine water from Konkan, Maharashtra, India. The dried marine bacteria were homogenized to fine powder and further subjected to extraction.

Crude Extraction

The crude ethanol extract was obtained by extracting 15gm of dried powder in 150 ml ethanol and was kept on a rotary shaker for 24hrs. The extract was filtered, centrifuged at 5000rpm for 15 min and was dried under reduced pressure. The yield obtained for ethanol extract was 40.10% with respect to the initial dry material. The extract was stored at 5°C in airtight bottles.

HPLC spectrum of ethanol extract of Nitella hyalina

HPLC was applied for testing the presence of number of organic compounds available of ethanol extract of *Nitella hyalina* One of the major organic component with 90.274% and 7.074 retention time may have antifungal activity. Method

Column : YMC ODS 150 mm × 4.6 mm × 5 μ , ID: E-AC-1/06/COL/013 Mobile Phase: A: 0.05 % TFA in Water /B: 0.05 % TFA in Acetonitrile Inj. Vol: 10 μL , Col. Temp.: 30°C, Flow rate: 1.4 mL/min Gradient: 5% to 95 % B in 8 min, Hold for 1.5 min ,9.51-12 min 05 % B



PDA Ch1 254nm 4nm	PDA	Ch1	254nm	4nm	
-------------------	-----	-----	-------	-----	--

Peak#	Ret. Time	Area	Area %	Peak Purity Index
1	5.758	896	0.239	Cannot be calculated
2	6.221	1788	0.476	0.99990
3	6.956	30592	8.143	0.56194
4	7.074	339130	90.274	0.99926
5	7.968	3260	0.868	0.99904
Total		375668	100.000	

Antifungal investigation

Antifungal activity of Nitella hyalina extract was tested by the agar diffusion method described in European Pharmacopoeia. Minimum inhibitory concentration (MIC) was determined using the serial two-fold broth dilution method described by Pepeljnjak et al. . For the agar diffusion method, 25 ± 2 mL of sterile and melted SGA at 45–50°C was inoculated with 1 mL of approximately 1to5 x 10^{6} CFU mL⁻¹ of yeast cells or conidia and mycelial structures of dermatophytes in sterile physiological saline in Petri dishes (9cm). Inoculum density was measured with McFarland's standard solution of freshly prepared barium sulfate in sterile water; density of 0.01% BaCl₂ in 1% H₂SO₄ solution equals approximately 3 x 10^8 cells mL⁻¹. After drying SGA at room temperature for a maximum of 30 minutes, holes of 6 mm in diameter were made with stainless steel cylinders and filled (60 μ L) with the ethanol extract. One experiment was done in a Petri dish for one fungal strain. Plates were then incubated at $+4^{\circ}$ C for 1 hour and at $25 \pm 2^{\circ}$ C for 48hrs for veasts and 7 days for dermatophytes. After the incubation period, inhibition zones were measured and expressed in mm. Minimum inhibitory concentartion (MIC) and minimum fungicidal concentration (MFC) were determined using the serial broth dilution method described by Pepeljnjak et al. . MIC was determined by the broth two-fold macro dilution method in Sabouraud 2% glucose broth starting with 50% (V/V) of the fluid extract or essential oil. MIC is defined as the lowest concentration of extract or essential oil that allows no more than 20% growth of the fungi, visualized as a reduced number of colonies after removing the loop with approx. 10 μ L of each dilution on SGA and incubation at 25 $\pm 2^{\circ}$ C for 72 hrs for yeasts and 7 days for dermatophytes. *MFC* is defined as the lowest concentration of the extract that completely inhibited the growth of fungi.

Newsletter

Results and Conclusions

In the Nitella hyalina sample studied, HPLC was applied for testing the presence of number of organic compounds available of ethanol extract Nitella hyalina. Antifungal activity is shown in Tables I and II. It shows antifungal activity against all the species of dermatophytes investigated, with inhibition zones from 20 to 27 mm. The largest inhibition zone was observed against T. mentagrophytes (27 mm). The ethanol extract Nitella hyalina of extract had a well-balanced antifungal effect on the fungi species Candida albicans, C. parapsilosis with an inhibition zone of 14 to 16 mm. The ethanol extract Nitella hyalina of extract shows antifungal activity against all species of dermatophytes studied, with inhibition zones of 22 mm Trichophyton rubrum and 27 mm (Trichophyton mentagrophytes), All the fungi investigated are sensitive to the ethanol extract Nitella hyalina. inhibition of the growth of fungi was observed both on the surface and in the depth of the agar during the experiment when the ethanol extract Nitella hyalina was dropped into the hole (60 µL) of the Petri dish: concentration of the ethanol extract Nitella hyalina aerosol in the Petri dish was 0.9 µL cm⁻³, with an incubation temperature of 25±2°C. Data on minimum inhibitory concentrations and mimimum fungicidal concentrations for the ethanol extract Nitella hyaline

The antifungal activity of the ethanol extract *Nitella hyalina* was testes invitro. Yeasts *Candida albicans, C. parapsilosis* and Dermatocytes *Trichophyton rubrum, T. mentagrophytes* showed more potent antifungal activity. The extract inhibited the growth of Yeasts and dermatophytes in lower concentrations than the extract itself.

	Fungi	Inhibition Zo Reference substance	ne (mm)* ethanol extract
Yeasts	Candida albicans	18 mm	16 mm
	C. parapsilosis	18 mm	14 mm
Dermatophytes	Trichophyton rubrum	21 mm	22 mm
	T. mentagrophytes	24 mm	27 mm

Table I. Antifungal activity of anise ethanol extract *Nitella hyalina* by the diffusion method

a: Nistatin as reference substance (0.5 mg mL^{-1}) for yeasts and ketoconazole for dermatophytes.

b: GPA – growth promotion activity.

c: AE – fungicidal influence of aerosol (0.7 µL cm–3).

Pharmacologyonline 3: 34-39 (2008) Newsletter Pokharkar *et al.*

		Inhibition Zone (mm)*			
	Fungi	Reference substance $(\mu g m L^{-1})$		ethanol extract (%, v/v)	
		MICb	MFCc	MIC	MFC
Yeasts	Candida albicans	0.5	1	18	26
	C. parapsilosis	1	2	17	25
Dermatophytes	Trichophyton rubrum	0.25	0.5	1.4	4
	T. mentagrophytes	0.50	1	8.0	18

Table II. Antifungal activity of anise ethanol extract Nitella hyalina by the dilution method

a Nistatin as reference antimycotic against yeasts tested and ketoconazole against dermatophytes tested.

b *MIC* – minimum inhibitory concentration.

c MFC – minimum fungicidal concentration.

d Significantly different from fluid extract (p < 0.01).

nt – not tested.

References

- 1. P. Eggimann, J. Garbino and D. Pittet, Epidemiology of *Candida* species infections in critically ill non-immunosuppressed patients, *Lancet Infect. Dis.* **3** (2003) 685–702.
- 2. V. Krcmery and A. J. Barnesz, Non-albicans *Candida* spp. causing fungaemia: pathogenicity and antifungal resistance, *J. Hosp. Infect.* **50** (2002) 243–260.
- 3. J.-P. Latgé and R. Calderone, Host-microbe interactions: fungi, Invasive human fungal opportunistic infections, *Curr. Opin. Microbiol.* **5** (2002) 355–358.
- 4. R. Hänsel, O. Sticher and E. Steinegger, *Pharmakognosie-Phytopharmazie*, 6th ed., Springer-Verlag, Berlin 1999, pp. 692–695.
- 5. V. E. Tyler, L. R. Brady and J. E. Robbers, *Pharmacognosy*, 9th ed., Lea and Fabiger, Philadelphia 1988, p. 125.
- 6. J. Lawless, *The Illustrated Encyclopedia of Essential Oils*, The Bridgewater Book Company Ltd., Shaftesbury 1999, pp. 44–45.
- 7. K. Hostettmann, O. Potterat and J.-L. Wolfender, The potential of higher plants as a source of new drugs, *Chimia* **52** (1998) 10–17.
- 8. M. F. Balandrin, J. A. Klocke, E. S. Wurtele and W. H. Bollinger, Natural plant chemicals: sources of industrial and medicinal materials, *Science* **228** (1985) 1154–1160.

Pharmacologyonline 3: 34-39 (2008) Newsletter Pokharkar et al.

- 9. S. Pepeljnjak, I. Kosalec, Z. Kalo|era and D. Ku{trak, *Natural Antimycotics from Croatian Plants*, in *Plant-derived Antimycotics* (Eds. M. Rai and D. Mares), The Haworth Press, Binghampton 2003, pp. 49–81.
- 10. E. O. Lima, O. F. Gompertz, A. M. Giesbrecht and M. Q. Paulo, *In vitro* antifungal activity of essential oils obtained from officinal plants against dermatophytes, *Mycoses* **36** (1993) 333–336.
- 11. S. A. Zacchino, R. A. Yunes, V. C. Filho, R. D. Enriz, V. Kouznetsov and J. C. Ribas, The Need for New Antifungal Drugs: Screening for Antifungal Compounds with a Selective Mode of Action with Emphasison the Inhibitors of the Fungal Cell Wall, in Plant-derived Antimycotics (Eds. M. Rai and D. Mares), The Haworth Press, Binghampton 2003, pp. 1–47.
- 12. F. C. Czygan, Anis (Anisi fructus DAB 10) Pimpinella anisum, Z. Phytother. 13 (1992) 101–106.
- 13. *European Pharmacopoeia*, 4th ed, Council of Europe, Strasbourg 2001, pp. 93–99.