

Doxorubicin-Induced Dilated Cardiomyopathy: Physiopathological Mechanisms and Therapeutic Alternatives.

Yanet Hernández-Matos¹, Livan Delgado Roche¹

1. Center of Studies for Research and Biological Evaluations (Department of Biochemistry). Pharmacy and Food Sciences College, University of Havana. Havana, Cuba.

* Author for correspondence: Ave. 23 # 21425 e/ 214 y 222, La Coronela, La Lisa. Ciudad de la Habana. Cuba. E-mail: ymatos@ifal.uh.cu

Summary

The doxorubicin (DOX) is one of the most effective chemotherapeutic agents against a wide variety of cancers. However, its use is seriously limited by the development in the heart of acute and chronic toxic effects. Oxidative stress is the major etiopathological factor in DOX-induced cardiotoxicity. Mechanisms of action and toxicity of DOX are revised in this review, as also the potential therapeutic alternatives. The mechanism of doxorubicin-induced dilated cardiomyopathy is characterized by left ventricular dilation that is associated with systolic dysfunction also the strategies to attenuate DOX toxicity are dosage optimisation, synthesis and use of analogues or combined therapy with antioxidants. The most promising results come from the combination of the drug delivery together with an antioxidant in order to reduce oxidative stress.

Key Words: doxorubicin, dilated cardiomyopathy, oxidative stress, antioxidant defense.

Introduction

The doxorubicin (DOX) is an excellent drug for the treatment of a wide variety of human tumors. However, the development of irreversible cardiotoxicity has limited its use. Due to the successful action of DOX as a chemotherapeutic agent, several strategies have been tried to prevent/attenuate the side effects of DOX (1). DOX-induced cardiotoxicity is thought to be a complex multifactorial process, which includes oxidative stress. DOX localizes to the mitochondria and is highly susceptible to enzymatic reduction to generate superoxide radicals and reactive oxygen species (ROS), which can cause mitochondrial swelling and ultrastructural changes and alter mitochondrial function (2). ROS play an important role in the pathogenesis of the chronic cardiotoxicity and are in part generated by redox-cycling of DOX. DOX also enters mitochondria where it either directly inhibits the respiratory chain by binding to cardiolipin, or interacts with mitochondrial DNA (mtDNA). Somatic acquired and accumulating respiratory chain defects which perpetuate ROS formation and contributing quantitative and qualitative lesions in mtDNA (mtDNA depletion, deletions, and point mutations) represent a plausible mechanism for the delayed onset of the cardiomyopathy and the molecular basis of the dose memory for the cumulative doxorubicin exposure (3).

DOX-induced oxidative stress in cardiomyocyte mitochondria is associated with cell death and responsible for cardiac damage (2). DOX increases cardiomyocyte susceptibility by reducing SOD activity and therefore reducing the ability of cardiac cells to inactivate ROS. Oxidative stress also has been shown to cause depolarization of the mitochondrial membrane, leading to programmed cell death (4). The production of free radicals and oxidative stress is closely involved with DOX action, regarding both anti-tumour and toxic effects (5).

Myocardial mitochondria also play an important role in the physiological regulation of the intracellular homeostasis of calcium ions by either sequestering or releasing Ca²⁺. It is not known whether the alterations in myocellular Ca²⁺ homeostasis are primary or secondary to the respiratory chain dysfunction in chronic DOX cardiomyopathy. In vitro models suggest that the DOX-induced Ca²⁺ uptake by cardiomyocytes and their mitochondria inhibits Complexes I and IV of the respiratory chain and enhances ROS generation, effects which may subsequently limit ATP production. These data are supported by the observation that decreased cellular ATP levels and thus mitochondrial dysfunctions were preceded by increases in mitochondrial Ca²⁺ (3).

DOX-induced cardiomyopathy: Dilated cardiomyopathy

DCM, the most common of the cardiomyopathies, is characterized by an increase in both myocardial mass and volume. The walls become thin and stretched, compromising cardiac contractility and ultimately resulting in poor left ventricular function. DCM may occur at any age, but is common in males between the ages of 20 and 50 years (6). Genetically inherited (familial) forms of DCM have been identified in 25–35% of patients presenting with this disease, but many other acquired conditions may result in an identical clinical presentation and pathological function. These conditions include alcohol-induced cardiomyopathy, peripartum cardiomyopathy, haemochromatosis, chronic anaemia, non-compaction cardiomyopathy, DOX toxicity, sarcoidosis and viral myocarditis. DCM may also occur secondary to ischaemic heart disease, valvular heart disease, hypertension and congenital heart disease (7). In cases where an underlying pathology cannot be identified, the patient is diagnosed with an idiopathic dilated cardiomyopathy (iDCM). The 5 year survival after diagnosis is 50%, as patients often develop progressive congestive heart failure (CHF) and complications such as thromboembolic conditions and arrhythmias. Mutations in lamin A/C are often associated with conduction system disease; desmin and dystrophin with a skeletal myopathy; desmoplakin and plakoglobin are often associated with woolly hair and keratoderma. Mutations in d-sarcoglycan have been found to cause loss of skeletal muscle and a form of muscular dystrophy (8).

DOX is very effective anticancer drug that is prescribed worldwide. However, many patients treated with this drug, irrespective of age, develop insidious DCM and heart failure. The scope of the problem remains to be adequately defined. Causative mechanism of the disease is multifactorial, but seems to be associated with generation of ROS, disruption of mitochondria, and uncoupling of the electron-transport chain (9, 10). Use of dexrazoxane might be cardioprotective by attenuation of the formation of free radicals. Early detection of disease by both non-invasive and serological techniques shows promise (11).

Left ventricular non-compaction

Left ventricular non-compaction can be difficult to diagnose unless the physician has a high level of suspicion during echocardiographic assessment. With careful review of echocardiograms and other clinical data, this disorder seems to be common in children, and is also reported in adults (12). About 9% of all cases of cardiomyopathy are diagnosed as left ventricular noncompaction, with only DCM and hypertrophic cardiomyopathy being more common. In the most recent American College of Cardiology/American Heart Association (AHA/ACC) cardiomyopathy classification, left ventricular non-compaction was recognised for the first time as a formal form of cardiomyopathy (13). A substantial percentage of these patients have a dilated left ventricle with systolic dysfunction, mimicking DCM. Signs, symptoms, and outcomes of these patients mirror those of patients with pure DCM, but in young children, outcomes are worse than for those with DCM.

Doxorubicin: molecular mechanisms and toxicity

Adriamycin, also called DOX, is an antibiotic anthracycline that was isolated from a pigment of *Streptomyces peacetius* in the early 1960s but which is now chemically synthesised. Having been employed for more than 30 years in the battle against cancer, DOX is essential in treating breast and oesophageal carcinomas, solid tumours in childhood, osteosarcomas, Kaposi's sarcoma, soft tissue sarcomas, and Hodgkin and non-Hodgkin lymphomas (14, 15). Anthracyclines (doxorubicin, daunorubicin, epirubicin, and idarubicin) are the most effective anti-neoplastic family in current clinical practice.

Specifically, DOX is a key chemotherapeutic drug for cancer treatment; although its use is limited by the chronic and acute toxic side effects it produces (15). Acute side effects related to intravenous injection of DOX appear within minutes after infusion, including nausea, vomiting, myelosuppression and arrhythmia. On the other hand, chronic effects may develop several weeks or even months after the recurrent administration of the drug, provoking heart, liver, brain or kidney injury. Since cardiomyocytes, as well as neurons, are post-mitotic cells, the vast majority of the damage is irreversible, and unalterably affects cardiac and brain functions. Furthermore, heart repercussions are even more prevalent because of this organ's greater sensitivity to damage induced by free radicals, given the high oxidative metabolism of the heart and its lower level of antioxidant defences.

DOX-related cardiotoxicity may cause dose-dependent cardiomyopathy and congestive heart failure; moreover, persistent changes of cognitive function (memory and concentration loss and difficulty of performing multiple tasks) may occur (15, 16). Dosage-dependent chronic cardiomyopathy associated with DOX administration generates marked hypotension, tachycardia, cardiac dilatation and ventricular failure. At the serum level, increased glutamate oxalacetic transaminase, lactate dehydrogenase, and creatinine phosphokinase enzyme activities have been noted.

At the ultrastructural level, myofibril loss, mitochondrial swelling, cytoplasmic vacuolization, and an increased number of lysosomes have been reported (17, 18). Toxicity associated with DOX therapy in the brain is due to the indirect action of the drug, because it is not able to cross the blood-brain barrier. DOX raises circulating levels of tumour necrosis factor (TNF α), which can cross this barrier, reaching and activating glial cells to initiate the local production of TNF α and raise its circulating

levels. This TNF α induces the local generation of reactive oxygen species (ROS) through nitric oxide synthase induction, and therefore intensifies the oxidative stress responsible for brain injury (16). Molecular mechanisms of DOX account for both its anti-cancer and its toxic effects (in the heart, brain, kidney, etc.). DOX acts at two fundamental levels: altering DNA and producing free radicals; in this respect, too, other mechanisms have been studied (5).

Generation of free radicals during metabolism of doxorubicin

The production of free radicals and oxidative stress is closely involved with DOX action, regarding both anti-tumour and toxic effects. There are four different modes of free radical production by DOX:

(1) Production of semiquinone: DOX is transformed into a semiquinone free radical through electron reduction by various NAD (P)H-dependent reductases in the complex I of the electron transport chain (cytochrome P-450 reductase). This semiquinone reacts with molecular oxygen to produce the superoxide radical ($O_2^{\cdot-}$) and it converts DOX into quinone. This quinone–semiquinone cycle generates large amounts of $O_2^{\cdot-}$, which subsequently give rise to ROS and RNS species such as hydrogen peroxide (H_2O_2), hydroxyl radical (HO^{\cdot}) or peroxynitrite ($ONOO^{\cdot}$) (15, 16).

(2) Activation of NAD(P)H oxidases (NOXs): DOX activates NOXs, which give rise to free radicals that participate in activating the apoptotic pathway in cardiac cells (19). NOX activation may generate $ONOO^{\cdot}$ through the mitochondrial production of ROS as $O_2^{\cdot-}$ and the reaction with nitric oxide (20). $ONOO^{\cdot}$ also activates matrix metalloproteinases (MMPs); these proteases have been implicated as a major mechanism of the $ONOO^{\cdot}$ dependent cardiotoxicity of DOX (21).

(3) DOX free radicals come from a non-enzymatic mechanism that involves reactions with iron. For example, Fe^{3+} reacts with DOX in a redox reaction after which the iron atom accepts an electron and a Fe^{2+} -DOX free radical complex is produced. This iron-adriamycin complex can reduce oxygen to H_2O_2 and other ROS (22, 23).

(4) Non-enzymatic mechanism: DOX interferes with non-enzymatic metabolic reactions in which iron is involved, and leads to ROS production. Thus, the DOX semiquinone, $O_2^{\cdot-}$, and H_2O_2 , can promote the release of iron from ferritin and cytoplasmic aconitase, thus altering iron metabolism. Subsequently, iron can react with DOX and subsequently produce HO^{\cdot} (14, 16).

(5) Products from the metabolism of DOX: this metabolism leads to ROS production. On the one hand, the side chain carbonyl group of the carbon 13 in DOX is converted into a hydroxyl group by aldoketo reductases, giving rise to a secondary alcohol (doxorubicinol), which can release iron from cytoplasmic aconitase, disturbing the iron metabolism and, therefore, causing oxidative stress. On the other hand, it can be metabolised into a lipophilic aglycone capable of diffusing through the mitochondrial membrane and accumulating within it. This aglycone is the starting point for several reactions that release electrons, producing ROS and disturbing the functional integrity of the respiratory chain (16). All the oxidative mechanisms described above are triggered by DOX to induce cancer cell death and toxic effects in cardiac myocytes. Moreover, it is important to take into account that the heart is very rich in mitochondria. DOX has the ability to modify the chemical composition, structure and function of biological membranes, mainly at the mitochondrial level, fundamentally due to the

peroxidation generated by DOX (24-26). It has been reported that phospholipid peroxidation induced by DOX can cause an exchange of mitochondrial and microsomal cholesterol with exogenous pools (24). The mitochondria contain a phospholipid, cardiolipin, in their inner membrane, and DOX has a high affinity for this cardiolipin, which results in the accumulation of DOX inside cardiac cells (15, 17). This effect may be enhanced by a highly unsaturated diet, producing a cardiolipin that is very rich in highly peroxidisable unsaturated fatty acids (26). In 2007, berthiaume and Wallace reported a gene expression profile in the heart of male rats treated with 6 weekly injections of 2 mg/kg DOX followed by a 5-week drug free period. Several pathways are closely related to mitochondria, including glycolysis and fatty acid metabolism, which supports the hypothesis that these organelles are key targets in DOX toxicity.

The same authors (27) also reviewed the importance of the mitochondrion in DOX cardiotoxicity, this being an important target of this chemotherapeutic drug, which induces ROS production due to the DOX redox cycling at complex I of the electron transport chain. This oxidative damage can impair both short and long term mitochondrial functioning, causing reduced energy production, oxidation of the mitochondrial DNA and loss of mitochondrial membrane potential by generating mitochondrial permeability transition (MPT) pores. It has been suggested that the adenine nucleotide translocator may be the principal component of those MPT pores, and the concentration of this protein has been shown to diminish after DOX administration, thus aggravating mitochondrial dysfunction (28). In addition to these events, a decrease in other mitochondrial components such as coenzyme Q (CoQ) has been reported, as a consequence of the oxidative stress associated with the administration of DOX (25).

Mechanisms related to alterations of DNA

Thus, it has been reported that DOX leads to direct oxidative injure to DNA (29, 30) and generates lipid peroxidation (25, 26, 31).

Topoisomerase II is likely to be one of the primary targets for the activity of anthracycline antibiotics (32). The induction by DOX of strand breaks in the DNA of L1210 leukemic cells was described more than 20 years ago (33). These strand breaks were protein-associated and slowly and incompletely repaired after removal of cells from the presence of the drug. Tewey et al. (1984) (34) have shown that topoisomerase II as the target enzyme for DOX and demonstrated that the subunits of the homodimeric enzyme remain locked onto the 5' end of the DNA molecule after completing the cleavage reaction. The capacity of DOX to inhibit DNA synthesis has been proposed as a mechanism of action of DOX (32). This mechanism may be related to DNA intercalation or inhibition of DNA polymerase activity (35). It is possible that this effect may be related to signalling events of growth arrest and p53 function (36). Another way of action of DOX through alterations in DNA is the induction of enzymatic or chemically activated DNA adducts (37), and DNA cross-linking (38).

Interference with DNA strand separation and helicase activity has also been postulated as mechanisms of action for DOX. It has been demonstrated that low concentrations of DOX interfere with DNA unwinding in MCF-7 breast tumour cells. This finding could potentially be related to the induction of DNA cross-links or to drug effects at the level of helicases (32).

Such genetic disturbances, together with the activation of tumour suppressor p53, and other additional mechanisms, are responsible for the apoptosis induced by DOX. Among those additional mechanisms, DOX can trigger apoptosis by producing ceramide (which prompts apoptosis by activating p53 or other downstream pathways such as JNK), the degradation of Akt by serine threonine proteases, the mitochondrial release of cytochrome c, increased FasL (death receptor Fas/CD95 ligand) mRNA production (14, 15, 17, 39), and a greater production of free radicals (16).

Tumor suppressor protein p53 occupies a pivotal position in maintaining genomic integrity (40). In response to cellular stresses that lead to DNA damage, wild-type p53 orchestrates the transcription of numerous genes and directs cells to cell cycle arrest, senescence, or apoptosis via differential activation of target genes (41), preventing the propagation of damaged DNA (42). One of the most important questions in the study of p53 is how p53 determines a specific cellular outcome (e.g., selecting cell cycle arrest between senescence and apoptosis) via selectively regulating certain groups of target genes. Current knowledge shows that various effectors, including proteins and even noncoding RNAs (such as Myc (43), hCAS/CSE1L (44), Hzf (45), and miR-34 (46)), can play a role in selective transactivation of p53 target genes that leads to different cellular outcomes.

Given that both ROS and p53 participate in multiple cellular processes, there should be interactions between ROS and p53 and intersections between their signaling pathways. A microarray analysis of H₂O₂-treated human cells identified one-third of the 48 highly H₂O₂-reponsive genes as targets of p53 (47). Though it is generally recognized that oxidative stress is associated with p53-dependent cell cycle arrest, DNA repair, and apoptosis, a clear understanding of the mechanisms of the interactions between ROS and p53 is still elusive.

Metabolism of calcium and induced-DOX apoptosis

Many studies have reported alterations in calcium metabolism (mainly calcium overload) after DOX treatment (48). These alterations include increase in intracellular Ca²⁺, accumulation of calcium in ventricular myocardium, and Ca²⁺ inclusions in mitochondria, Ca²⁺ transport abnormalities in cardiac tissue and alterations in the Ca²⁺ release function of the sarcoplasmic reticulum by effects on the Ca²⁺-ATPase and the Ca²⁺ release channel.

Myocardial mitochondria also play an important role in the physiological regulation of the intracellular homeostasis of calcium ions by either sequestering or releasing Ca²⁺ (49). For this purpose, mitochondria and cardiomyocyte membranes are equipped with a complex array of Ca²⁺ transporters. Ca²⁺ is a key regulator of muscle contraction and of important mitochondrial enzymes such as pyruvate dehydrogenase, isocitrate dehydrogenase, α -ketoglutarate dehydrogenase, and ATP synthase (50, 51). Doxorubicin interferes at various levels with the mitochondrial membrane and furthermore induces an influx of Ca²⁺ from the extracellular compartment into the cardiomyocytes (52). Alterations in mitochondrial Ca²⁺ transport in heart have been reported to range from inhibition of uptake by mitochondria upon exposure to doxorubicin in vitro to stimulation of uptake in rabbits after repeated DOX exposure. It is not known whether the alterations in myocellular Ca²⁺ homeostasis are primary or secondary to the respiratory chain dysfunction in chronic DOX cardiomyopathy. In vitro models suggest that the DOX-induced Ca²⁺ uptake by cardiomyocytes and their mitochondria inhibits

Complexes I and IV of the respiratory chain and enhances ROS generation, effects which may subsequently limit ATP production (53). These data are supported by the observation that decreased cellular ATP levels and thus mitochondrial dysfunctions were preceded by increases in mitochondrial Ca^{2+} . These studies, however, were carried out in vitro and therefore more closely reflect acute rather than chronic doxorubicin cardiotoxicity. We therefore aimed to investigate the effects of DOX on the mitochondrial respiratory chain and Ca^{2+} homeostasis in a model which closely mimics the chronic course of the patients (3). Heart failure may lead to the accumulation of calcium inside cardiac cells, making the described increase in Ca^{2+} levels a consequence of DOX action more than a cause. The fact that the multi-drug resistance pump frequently mediates resistance to the anthracyclines suggests the concept that DOX must enter the cell to express its toxicity. In fact, cell membrane and mitochondrial membrane have been related to DOX action (32).

There is extensive evidence that treatment with DOX induces apoptosis (38). However, apoptosis appears only at low concentrations of the drug, which supports the hypothesis that elevated concentrations of anthracyclines may kill the tumour cell through nonphysiological mechanisms (32). Another proposed mechanism is the formation of DOX metabolites.

Use of antioxidant in the prevention of DOX-induced cardiotoxicity

DOX increases cardiomyocyte susceptibility by reducing SOD activity and therefore reducing the ability of cardiac cells to inactivate ROS (54). Oxidative stress also has been shown to cause depolarization of the mitochondrial membrane, leading to programmed cell death (55).

Due to the great importance of DOX in chemotherapy for the treatment of many types of cancer, researchers have expended great efforts trying to prevent or attenuate the side effects of DOX administration. In this sense several strategies have been followed as dosage optimisation, synthesis and use of analogues or combined therapy. No promising results have been found with different routes and schedules of drug delivery (56, 57) and the use of several DOX analogues available clinically did not show stronger antitumor efficacy or lower toxic effects than DOX (58). In relation to combined therapy, although antitumor action of DOX may be mediated by a wide number of mechanisms, free radical production is among the main causes of cardiotoxicity mediated by this drug. This fact allows the assay of strategies trying to reduce the toxic effects of DOX without interference with its antitumor properties. The most immediate approach has been the combination of the drug delivery together with an antioxidant in order to reduce oxidative stress (59).

Vitamin E (α -tocopherol) has a high antioxidant capacity and plays a fundamental biologic role, especially in protecting cells and tissues from oxidative damage, and membrane lipid and lipoprotein peroxidation (15, 17). In general, preclinical studies in rodents have shown that oral vitamin E tends to increase antitumour actions and protects against the toxic effects of DOX (15); nevertheless, it has been reported that both vitamins E and C can increase the expression of P-glycoprotein and of hypoxia inducible factor-1 α in Nox-1 overexpressing prostate tumour cells (60). Therefore, these vitamins might fortify resistance to chemotherapy, although this proposal needs to be more extensively studied. As a beneficial agent, vitamin E has been found to lengthen the life span of laboratory animals and to diminish the weight loss provoked by

chemotherapy. Moreover, this antioxidant can protect from both acute and chronic cardiotoxicity caused by DOX, and it increases antioxidant capacity in the heart (15).

Vitamin C (ascorbic acid) is an effective water soluble antioxidant against lipid peroxidation, scavenging ROS in the aqueous fraction before these molecules can give rise to lipid oxidation (61). An intervention review by van Dalen et al. (2008) showed that none of the individual studies carried out demonstrated a cardioprotective effect by vitamin C combined with vitamin E, mainly administered by oral supplementation (62).

Carotenoids such as β -carotene can reduce the lipid peroxidation associated with DOX and augment the anti-tumour effect of this drug (63). Oral supplementation with lycopene (5 mg/kg/day for 7 weeks), the carotenoid presenting the most powerful antioxidant activity, has demonstrated a cardioprotective effect at the myocyte level in rats treated with DOX (4 mg/kg) intraperitoneally by weeks 3, 4, 5 and 6, but it fails to prevent DOX-induced cardiac dysfunction (1). Moreover, it has been reported that a tomato oleoresin supplement containing lycopene (95%), all-trans- β -carotene (5%), and 13-cis- β -carotene (1%), reduces cardiomyocyte oxidative DNA damage caused by DOX in rats (64). The same authors reported that DOX maintains levels of lycopene in the myocardial tissue of rats, and at the same time it raises the antioxidant capacity of this tissue. This suggests DOX has an antioxidant more than a pro-oxidant effect (65). Vitamin A also has a protective dose-dependent effect against the chromosomal aberrations induced by DOX in rat bone marrow cells, 15 μ g/kg being the most effective dose, whereas 30 μ g/kg was found to be clastogenic (66).

Coenzyme Q (CoQ), or ubiquinone, plays a critical role in the mitochondrial respiratory chain, acting as a redox link between flavoproteins and cytochromes, being an essential component in extramitochondrial redox chains. Its concentration in blood and tissues depends on biologic requirements, endogenous biosynthesis, and of course the dietary intake (17). Due to the high presence of CoQ in the mitochondria, its concentration reflects the cellular content of mitochondria among different tissues, being greater in the heart than in the liver (fivefold more), kidney, pancreas, spleen (10-fold more), and skeletal muscle (5% more) (67). Adriamycin induces cardiotoxicity by lipid peroxidation in cardiac myocytes, reduces the content of CoQ10 in mitochondrial membranes, and inhibits the mitochondrial biosynthesis of CoQ10 as well as respiratory chain CoQ10-dependent enzymes (62). Similar effects have been found in rats where the plasma and mitochondria levels of CoQ10 and CoQ9, respectively, were sharply decreased by the oxidative stress generated by DOX (25).

Flavonoids are characterized by high antioxidant power, and have been considered potential protectors against the chronic cardiotoxicity associated with DOX (17). This protective effect of flavonoids is closely related to their antioxidant, iron chelating (68) and carbonyl reductase 1 (CBR1)-inhibitory properties (69). The proposed mechanism involving iron chelating and antioxidant activities involves two steps: (1) iron is chelated by the flavonoid; (2) ROS production is quickly scavenged by flavonoids at the place of generation; such a concept has been termed site-specific scavenging (68). The semisynthetic flavonoid 7-monohydroxyethylrutinose has been extensively studied as a good cardioprotective compound, both in preclinical (intraperitoneally administered in mice) (70) and in clinical trials after intravenous injection. This flavonoid inhibits negative cardiac effects in a dose-dependent manner, in accordance with the essential properties of all flavonoids, i.e. their iron chelating and antioxidant characteristics. Other flavonoids, such as catechins, have cardioprotective properties at low doses,

exhibiting an iron chelating activity. These catechins also have beneficial properties for the liver, and Kalender et al. (2005) reported that catechin (200 mg/kg/week for 6 weeks by intraperitoneal injection) depressed malondialdehyde, glutathione peroxidase and catalase activities in rats against DOX- induced hepatotoxicity (71). Two fundamental biochemical mechanisms of flavonoids can increase the anti-tumour capacity of DOX when they are administered jointly: (1) the inhibition of the intracellular metabolism of the drug; (2) the blocking of intracellular drug eliminating mechanisms. In previous in vitro studies, it has been shown that green tea polyphenols such as caffeine and catechins such as epigallocatechin gallate or epigallocatechin can enhance DOX-induced anti-tumour activity and increase DOX concentration in tumours by inhibiting its efflux (17).

Conclusions

Cellular damage induced by DOX is mediated by the formation of an iron-DOX complex that generates free radical, which, in turn, causes severe damage to the plasma membrane and interferences with the cytoskeletal structure. Due to the low levels of antioxidant defenses, heart is particularly vulnerable to injury by DOX-induced ROS. DOX can induce structural changes and death in the myocardium of treated patients, and the extent of structural damage has been correlated with early cardiac dysfunction.

Therapeutic strategies, designed to augment cellular endogenous antioxidant systems has been identified as a promising approach to combat oxidative stress-associated disease condition.

References

1. Anjos Ferreira AL, Yeum K-J, Matsubara LS, Matsubara BB, Correa CR, Percira EJ, et al. Doxorubicin as an antioxidant: Maintenance of myocardial levels of lycopene under doxorubicin treatment. *Free Rad Biol Med* 2007; 43:740-51.
2. Danz ED, Skramsted J, Henry N, Bennett JA, Keller RS. Resveratrol prevents doxorubicin cardiotoxicity through mitochondrial stabilization and the sirt1 pathway. *Free Rad Biol Med* 2009; 46: 1589-97.
3. Lebrecht D, Kirschner J, Geist A, Haberstroh J, Walker UA. Respiratory chain deficiency precedes the disrupted calcium homeostasis in chronic doxorubicin cardiomyopathy. *Cardiov Pathol* 2009; doi:10.1016/j.carpath.2009.06.006.
4. Xu M, Asharf M. Melatonin protection against lethal myocyte injury induced by doxorubicin as reflected by effects on mitochondrial membrane potential. *J Mol Cell Cardiol* 2002; 34: 75-9.
5. Granados-Principal S, Quiles JL, Ramírez-Tortosa CL, Sánchez-Rovira P, Ramírez-Tortosa MC. New advances in molecular mechanisms and the prevention of adriamicin toxicity by antioxidant nutrients. *Food Chem Toxicol* 2010; doi:10.1016/j.fct.2010.04.007.
6. Schoen F. The heart. In: Kumar V, Fausto N, Abbas AK, eds. *Robbins and Cotran pathological basis of disease*. Philadelphia: Saunders Elsevier, 2005:555–618.
7. Lester WM, Gotlieb AI. Cardiovascular effects of systemic diseases and conditions. In: Silver MD, Gotlieb AI, Schoen FJ, eds. *Cardiov Pathol*. New York: Churchill Livingstone, 2001:493–540.
8. Luk A, Ahn E, Soor GS, et al., Dilated cardiomyopathy: a review. *J Clin Pathol* 2009, 62: 219-25.

9. Wallace KB. Adriamycin-induced interference with cardiac mitochondrial calcium homeostasis. *Cardiov Toxicol* 2007; 7: 101-7.
10. Lebrecht D, Walker UA. Role of mtDNA lesions in anthracycline cardiotoxicity. *Cardiov Toxicol* 2007; 7: 108-13.
11. Gianni I, Herman EH, Lipshultz SE, Minotti G, Sarvazyan N, Sawyer DB. Anthracycline cardiotoxicity: from bench to bedside. *J Clin Oncol* 2008; 26: 3777-84.
12. Pantazis AA, Elliot PM. Left ventricular noncompaction. *Curr Opin Cardiol* 2009; 24: 209-13.
13. Towbin JA, Lowe AM, Colan SD, et al. Incidence, causes and outcomes of dilated cardiomyopathy in children. *JAMA* 2006; 296: 1867-76.
14. Minotti, G., Menna, P., Salvatorelli, E., Cairo, G., Gianni, L. Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. *Pharmacol Rev* 2004; 56: 185–229.
15. Quiles JL, Ochoa JJ, Huertas JR, Lopez-Frias M, Mataix J. Olive oil and mitochondrial oxidative stress: studies on adriamycin toxicity, physical exercise and ageing. In: Quiles JL, Ramirez-Tortosa MC, Yaqoob P (Eds.), *Olive Oil and Health*. CABI Publishing, Oxford, 2006: 119–51.
16. Chen Y, Jungsuwadee P, Vore M, Butterfield D.A, St Clair DK. Collateral damage in cancer chemotherapy: oxidative stress in nontargeted tissues. *Mol Interv* 2007; 7: 147–56.
17. Quiles JL, Huertas JR, Battino M, Mataix J, Ramirez-Tortosa MC. Antioxidant nutrients and adriamycin toxicity. *Toxicol* 2002; 180: 79–95.
18. Bertinchant JP, Polge A, Juan JM, Oliva-Lauraire MC, Giuliani I, Marty-Double C, et al. Evaluation of cardiac troponin I and T levels as markers of myocardial damage in doxorubicin-induced cardiomyopathy rats, and their relationship with echocardiographic and histological findings. *Clin Chim Acta* 2003; 329: 39–51.
19. Gilleron M, Marechal X, Montaigne D, Franczak J, Neviere R, Lancel S. NADPH oxidases participate to doxorubicin-induced cardiac myocyte apoptosis. *Biochem Biophys Res Commun*. 2009; 388: 727–31.
20. Kimura S, Zhang GX, Nishiyama A, Shokoji T, Yao L, Fan YY, et al. Role of NAD (P)H oxidase- and mitochondriaderived reactive oxygen species in cardioprotection of ischemic reperfusion injury by angiotensin II. *Hypertension* 2005; 45: 860–6.
21. Bai P, Mabley JG, Liaudet L, Virag L, Szabo C, Pacher P. Matrix metalloproteinase activation is an early event in doxorubicin-induced cardiotoxicity. *Oncol Rep* 2004; 11: 505–8.
22. Sinha BK, Polliti PM. Anthracyclines. *Cancer Chemother* 1990; 11: 45- 57.
23. Gianni L, Zweier JL, Levy A, Myers CE. Characterization of the cycle of iron-mediated electron transfer from adriamycin to molecular oxygen. *J Biol Chem* 1985; 260: 6820-6.
24. Huertas JR, Battino, M, Barzanti V, Maranesi M, Parenti-Castelli G, Littarru GP, Turchetto E, Mataix FJ, Lenaz G. Mitochondrial and microsomal cholesterol mobilization after oxidative stress induced by adriamycin in rats fed with dietary olive and corn oil. *Life Sci* 1992; 50: 2111–8.
25. Huertas JR, Battino M, Lenaz G, Mataix FJ. Changes in mitochondrial and microsomal rat liver coenzyme Q9 and Q10 content induced by dietary fat and endogenous lipid peroxidation. *FEBS Lett* 1991a; 287: 89–92.

26. Huertas JR, Battino M, Mataix FJ, Lenaz G. Cytochrome oxidase induction after oxidative stress induced by adriamycin in liver of rats fed with dietary olive oil. *Biochem Biophys Res Commun* 1991b; 181: 375–82.
27. Berthiaume JM, Wallace KB. Adriamycin-induced oxidative mitochondrial cardiotoxicity. *Cell Biol Toxicol* 2007; 23: 15–25.
28. Oliveira PJ, Wallace KB. Depletion of adenine nucleotide translocator protein in heart mitochondria from doxorubicin-treated rats—relevance for mitochondrial dysfunction. *Toxicology* 2006; 220: 160–8.
29. Gutteridge JMC, Quinlan GJ. Free radical damage to deoxyribose by anthracycline, aureolic acid and aminoquinone antitumour antibiotics. An essential requirement for iron, semiquinones and hydrogen peroxide. *Biochem Pharmacol* 1985; 34: 4099-103.
30. Feinstein E, Canaani E, Weiner LM. Dependence of nucleic acid degradation on in situ free-radical production by adriamycin. *Biochemistry* 1993; 32: 13156-161.
31. Mataix J, Mañas M, Quiles JL, Battino M, Cassinello M, López- Frías M, Huertas JR. Coenzyme Q content depends upon oxidative stress and dietary fat unsaturation. *Mol Asp Med* 1997; 18: 129-35.
32. Gewirtz DA. A critical evaluation of the mechanisms of action proposed for the antitumor effects of the anthracycline antibiotics adriamycin and daunorubicin. *Biochem Pharmacol* 1999; 57: 727-41.
33. Ross WR, Glaubiger DL, Kohn KW. Protein-associated DNA breaks in cells treated with adriamycin on ellipticine. *Biochim Biophys Acta* 1978; 519: 23-30.
34. Tewey KM, Rowe TC, Yang L, Halligam BD, Liu LF. Adriamycin-induced DNA damage mediated by mammalian topoisomerase II. *Science* 1984; 226: 466-8.
35. Tanaka M, Yoshida S. Mechanism of the inhibition of calf thymus DNA polymerase a and b by daunomycin and adriamycin. *J Biochem* 1980; 87: 911-8.
36. Kastan MB, Onyckwere O, Sidransky D, Vogelstein B, Craig RW. Participation of p53 in the cellular response to DNA damage. *Cancer Res* 1991; 51: 6304-11.
37. Cullinane C, Cutts SM, van Rosmalen A, Phillips DR. Formation of adriamycin-DNA adducts in vitro. *Nucleic Acids Res* 1994; 22: 2296-303.
38. Skladanowski A, Konopa J. Interstrand DNA cross-linking induced by anthracyclines in tumour cells. *Biochem Pharmacol* 1994; 47: 2269-78.
39. Ozben T. Oxidative stress and apoptosis: impact on cancer the therapy. *J Pharm Sci* 2007; 96: 2181-96.
40. Vousden KH, Lu X. Live or let die: the cell's response to p53. *Nat Rev Cancer* 2002; 2: 594-604.
41. Gang Liu XC. Regulation of the p53 transcriptional activity. *J Cell Biochem* 2006; 97: 448-58.
42. Lim YP, Lim TT, Chan YL, Song ACM, Yeo BH, Vojtesek B, et al. The p53 knowledge base: an integrated information resource for p53 research. *Oncogen* 2006; 26: 1517-21.
43. Seone J, H-V Le, Massague J. Myc suppression of the p21 cip1 Cdk inhibitor influences the outcome of the p53 response to DNA damage. *Nature* 2002; 419: 729-34.
44. Tanaka T, Ohkabo S, Tatsuro I, Prives C. Hcas/Csell associates with chromatin and regulate expression of select p53 target genes. *Cell* 2007; 130: 638-50.

45. Das S, Raj I, Zhao B, Kimura Y, Berntein A, Aaronson SA, et al. H₂f determines cell survival upon genotoxic stress by modulating p53 transactivation. *Cell* 2007; 130: 624-37.
46. He L, He X, Lowe SW, Hannon GJ. MicroRNAs join the p53 network-another piece in the tumor-suppression puzzle. *Nat Rev Cancer* 2007; 7: 819-22.
47. Desaint S, Lurian S, Aude JC, Rousselet G, Toledano MB. Mammalian antioxidant defenses are not inducible by H₂O₂. *J Biol Chem* 2004; 279: 31157-63.
48. Arai M, Yoguchi A, Takizama T, Yokoyama T, Kanda T, Kurabayashi M, et al. Mechanism of doxorubicin-induced inhibition of sarcoplasmic reticulum Ca-ATPase gene transcription. *Circ Res* 2000; 86: 8-14.
49. Crompton M, Costi A. Kinetic evidence for a heart mitochondrial pore activated by Ca²⁺, inorganic phosphate and oxidative stress. A potential mechanism for mitochondrial dysfunction during cellular Ca²⁺ overload. *Ever J Biochem* 1988; 178: 489-501.
50. Das AM, Harris DA. Control of mitochondrial ATP synthase in heart cells: inactive to active transitions caused by beating or positive inotropic agents. *Cardiovasc Res* 1990; 24: 411-7.
51. McCormack JG, Denton RM. Mitochondrial Ca²⁺ transport and the role of intramitochondrial Ca²⁺ in the regulation of energy metabolism. *Dev Neurosci* 1993; 15: 165-73.
52. Lehninger AL. Ca²⁺ transport by mitochondria and its possible role in the cardiac contraction cycle. *Circ Res* 1974; 35: 83-90.
53. Zhou S, Starkou A, Froberg MK, Leino RL, Wallace KB. Cumulative and irreversible cardiac mitochondrial dysfunction induced by doxorubicin. *Cancer res* 2001; 61: 771-7.
54. Doroshov JH, Locker GY, Myers CE. Enzymatic defenses of the mouse heart against reactive oxygen metabolites: alterations produced by doxorubicin. *J Clin Invest* 1980; 65: 128-35.
55. Kim DS, Woo ER, Chae SW, Ha KC, Lee GH, Hong ST, et al. Plantainoside D protect adriamycin-induced apoptosis in H9c₂ cardiac muscle cells via the inhibition of ROS generation and NF-kappa B activation. *Life Sci* 2007; 80: 314-23.
56. Steinherz LS, Steinherz PG, Tan CTC, Heller G, Merphy L. Cardiac toxicity 4 to 20 years after completing anthracycline therapy. *J Am Med Assoc* 1991; 266: 1672-7.
57. Ewer MS, Jaffe N, Reid H, Zietz HA, Benjamin RS. Doxorubicin cardiotoxicity in children: comparison of consecutive divided dose administration schedule with single dose (rapid) infusion administration. *Med Pediatr Oncol* 1998; 32: 512-5.
58. Weiss RB. The anthracyclines: will we ever find a better doxorubicin? *Semin Oncol* 1992; 19: 670-86.
59. Singal PK, Li T, Kumar D, Danelisen I, Iliskovic N. Adriamycin-induced heart-failure: mechanism and modulation. *Mol Cell Biochem* 2000; 207: 77-85.
60. Wartenberg M, Hoffmann E, Schwindt H, Grünheck F, Petros J, Arnold JR, et al. Reactive oxygen species-linked regulation of the multidrug resistance transporter p-glycoprotein in Nax-1 overexpressing prostate tumour spheroids. *FEBS* 2005; 579: 4541-49.

61. Conklin KA. Dietary antioxidants during cancer chemotherapy: impact on chemotherapeutic effectiveness and development of side effects. *Nutr Cancer* 2000; 37: 1-18.
62. Van Dalen EC, Caron HN, Dickinson HO, Kremer LC. Cardioprotective interventions for cancer patients receiving anthracyclines. *Cochrane Database. Syst Rev* 2008; 2, CD003917.
63. Conklin KA. Chemotherapy-associated oxidative stress: impact on chemotherapeutic effectiveness. *Integr Cancer Ther* 2004; 3: 294-300.
64. Ferreira AL, Salvadori DM, Nascimento MC, Rocha NS, Correa CR, Pereira EJ, et al. Tomato-oleoresin supplement prevents doxorubicin-induced cardiac myocyte oxidative DNA damage in rats. *Mutat Res* 2007; 631: 26-35.
65. Ferreira AL, Yeum KS, Matsubara LS, Matsubara BB, Correa CR, Pereira ES, et al. Doxorubicin as an antioxidant: maintenance of myocardial levels of lycopene under doxorubicin treatment. *Free Rad Biol Med* 2007; 43: 740-51.
66. Gülçac MD, Akpınar G, Ustün H, Özön Kanlı A. Effect of vitamin A on doxorubicin-induced chromosomal aberrations in bone marrow cells of rats. *Mutagenesis* 2004; 19: 231-6.
67. Conklin KA. Coenzyme q10 for prevention of anthracycline-induced cardiotoxicity. *Integr Cancer Ther* 2005; 4: 110-30.
68. Kaiserová H, Simunek T, van der Vijgh WJ, Bast A, Kvasnicková E. Flavonoids as protectors against doxorubicin cardiotoxicity: role of iron chelation, antioxidant activity and inhibition of carbonyl reductase. *Biochim Biophys Acta* 2007; 1772: 1065-74.
69. Carlquist M, Frejd T, Gorna-Grauslund MF. Flavonoids as inhibitors of human carbonyl reductase 1. *Chem Biol Interact* 2008; 174: 98-108.
70. Bast A, Kaiserová H, den Hartoy GS, Haenen GR, van der Vijgh WJ. Protectors against doxorubicin-induced cardiotoxicity: flavonoids. *Cell Biol* 2007; 23: 39-47.
71. Kalender Y, Yel M, Kalender S. Doxorubicin hepatotoxicity and hepatic free radical metabolism in rats. The effects of vitamin E and catechin. *Toxicology* 2005; 209: 39-45.