## **ROLE OF OXIDATIVE STRESS IN PATHOGENESIS OF DIABETES AND ITS COMPLICATIONS**

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#### **Summary**

This review article is comprised of pathogenesis and role of free radicals and oxidative stress in progression of diabetes and diabetic complications. Reactive oxygen species are formed disproportionately in diabetes by glucose autooxidation, impaired polyol pathway, nonenzymatic glycation of proteins, and the subsequent oxidative degradation of glycated proteins. Abnormally high levels of free radicals and the simultaneous decline of antioxidant defense mechanisms can lead to damage cell structures, including lipids and membranes, proteins, DNA, increased lipid peroxidation, and development of insulin resistance. It has been suggested that enhanced production of free radicals and oxidative stress is central event to the development of diabetic complications. Changes in oxidative stress biomarkers including superoxide dismutase, catalase, glutathione reductase, glutathione peroxidase, glutathione levels, vitamins, lipid peroxidation, nitrite concentration, nonenzymatic glycosylated proteins, and hyperglycemia play major role in the pathogenesis of both types of diabetes mellitus as well as diabetic complications including diabetic neuropathy, diabetic nephropathy, diabetic retinopathy, dyslipidemia, coronary heart disease, diabetic foot ulcer, diabetic ketoacidosis and peripheral vascular diseases. Antioxidants, capable of neutralizing free radicals or their actions, act at different stages. They act at the levels of prevention, interception and repair caused by free radical induced cell damage.

Keywords: Reactive oxygen species; hyperglycemia; free radicals; Antioxidants; cell damage

### Abbreviations

AGEs: advanced glycation end products; ALA: alpha lipoic acid; ATP: adenosine triphosphate; CHD: Coronary heart disease; DHLA: dihydrolipoic acid; ECD: endothelial cell dysfunction; GST: glutathione S-transferase; GPx: glutathione peroxidase; GSH: glutathione; GFAP: glial fibrillary acidic protein; H2O2: hydrogen peroxide; HDL: high density lipoprotein; IL: interleukin; IRS: insulin receptor substrate; JNK: Jun N terminal kinase; LDL: low density lipoprotein; MAPK: mitogen-activated protein kinase; NO: nitrous oxide; NF: necrosis factor; 'OH: hydroxyl radical; PUFA: polyunsaturated fatty acids; PGI: prostaglandin I; PDGF: platelet derived growth factor; PKC: Protein kinase C; PKB: protein kinase B; ROS: reactive oxygen species; RAGE: receptor for advanced glycation end products; RNS: reactive nitrogen species; SOCS: suppressor of cytokine signaling proteins; SAPK: stress activated protein kinase; SOD: sulphoxide dismutase; TNFa: tumour necrosis factor alpha; TGF: transformine growth factor; VEGF: vascular endothelial growth factor; VLDL: vary low density lipoprotein; LOO': lipid peroxyl radicals.

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#### Introduction

Oxygen is an element indispensable for life. When cells use oxygen to generate energy, free radicals are created as a consequence of ATP production by the mitochondria. These by products are generally reactive oxygen species (ROS) as well as reactive nitrogen species (RNS) that result from the cellular redox process. These species play a dual role as both toxic and beneficial compounds. The delicate balance between their two antagonistic effects is clearly an important aspect of life. At low or moderate levels, ROS and RNS exert beneficial effects on cellular responses and immune function. At high concentrations, they generate oxidative stress, a deleterious process that can damage all cell structures [1-10]. Oxidative stress plays a major part in the development of chronic and degenerative ailments such as cancer, arthritis, aging, autoimmune disorders, cardiovascular and neurodegenerative diseases. The human body has several mechanisms to counteract oxidative stress by producing antioxidants, which are either naturally produced in situ, or externally supplied through foods and or supplements. Endogenous and exogenous antioxidants act as 'free radical scavengers' by preventing and repairing damages caused by ROS and RNS, and therefore can enhance the immune defense and lower the risk of cancer and degenerative diseases [11-15].

#### **Generation of free radicals**

Free radicals can be produced by several different biochemical processes within the body including: reduction of molecular oxygen during aerobic respiration yielding superoxide and hydroxyl radicals; by products of chemistry such as oxidation of catecholamines and activation of the arachidonic acid cascade product electrons, which can reduce molecular oxygen to superoxide; production of superoxide hypochlorous acid (HOCl), a powerful oxidant, by activated phagocytes; nitric oxide production by vascular endothelium and other cells. In addition, free radicals can be produced in response to external electromagnetic radiation, such as gamma rays, which can split water to produce hydroxyl radicals [16].

## Increased oxidative stress: Promoter of diabetes

There are many possible pathways of oxidative stress that can be associated with hyperglycemia [17-20].

#### Type I diabetes: Pancreatic β cell dysfunction

The common findings of prolong hyperglycemia and elevated lipid levels in the blood of diabetic patients led to the hypotheses of glucose toxicity [21], lipotoxicity [22], and oxidative stress [23-25]. These include glycolysis and oxidative phosphorylation; methyl glyoxal formation and glycation, enediol and acetaldehyde formation (glucoxidation); diacylglycerol formation and protein kinase C activation, glucosamine formation; and hexosamine metablolism and sorbitol metabolism. Conceptually, as  $\beta$  cells are exposed to high glucose concentrations for increasingly prolonged periods of time, glucose saturates the normal route of glycolysis and increasingly is shunted to alternate pathways, such that ROS are generated from distinct metabolic processes within and outside the mitochondria. It has been proved that excessive levels of palmitate are associated with abnormal islet function (especially in the presence of high glucose concentrations), which leads to excessive lipid esterification that, in turn, can generate ceramide, thereby increasing oxidative stress [22, 26,27]. It seems unlikely; however, that circulating lipid itself, such as triglyceride or cholesterol, would be responsible for damaging islet tissue. It seems more likely that excessive circulating glucose levels lead to accelerated de novo synthesis of islet lipid. One mechanism by which glucose might contribute to liptoxicity is by virtue of its ability to drive

synthesis of malonyl CoA, which inhibits  $\beta$ -oxidation of free fatty acids. This in turn shunts free fatty acids towards esterification pathways, thereby forming triglyceride, ceramide and other esterification products [27, 28].

Oxidative stress is responsible for the decreased protein expression and levels of Pdx-1 and Maf-A. Both proteins are critical for normal insulin gene expression, as their absence or mutation of their DNA binding sites on the insulin promoter leads to decreased insulin mRNA levels, insulin content and insulin secretion. Chronic hyperglycemia leads to worsening of  $\beta$  cell function [29-34]. It was demonstrated that pancreatic islets contain relatively small amounts of the antioxidant enzymes CuZn-SOD, Mn-SOD, catalase, and glutathione peroxidase (GPx) [35]. Due to the low level of antioxidant enzyme expression and activity, the  $\beta$  cells are at greater risk of oxidative damage than tissues with higher levels of antioxidant protection [36].

### Type II diabetes: Insulin resistance

Insulin rapidly interacts with its receptor at target tissues. The insulin receptor (IR, composed of two extracellular subunits and two transmembrane subunits linked by -S-Sbonds) possess an intrinsic tyrosine kinase activity. Tyrosine autophosphorylation of the IR subunit is induced following binding of insulin to the subunit [37]. The activated IR phosphorylates the insulin receptor substrate (IRS) proteins and other substrates. The process of phosphorylation leads to activation of different signaling pathways. While the ERK pathway is mainly involved in growth, the activation of phosphatidylinositol 3-kinase (PI 3kinase), mainly through insulin receptor substrates 1 and 2 (IRS1, IRS2), is involved in the metabolic actions of insulin. IRS1 belongs to the IRS family and plays a key role in insulin signaling. While the phosphorylation of IRS1 on tyrosine residue is critical for insulinstimulated responses, the phosphorylation of IRS1 on serine residues has a dual role: either to enhance or to terminate the insulin effects. The imbalance between the positive IRS1 tyrosine phosphorylation and the negative IRS1 serine phosphorylation is strongly stimulated by 'diabetogenic' factors including free fatty acids, TNFα and oxidative stress. Insulin activated protein kinase B (PKB) propagates insulin signaling and promotes the phosphorylation of IRS1 on serine residue, which in turn generates a positive-feedback loop for insulin action [38]. (Refer fig 1)

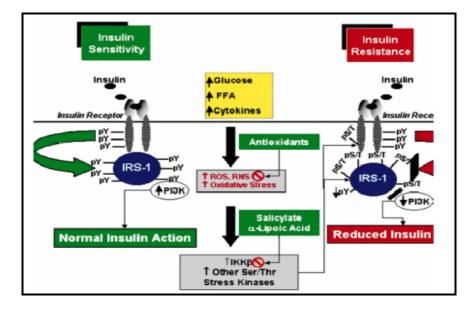


Fig 1: The role of serine kinase activation in oxidative stress-induced insulin resistance.

Various insulin resistance inducing agents such as angiotensin II, cytokines, free fatty acids, endothelin-1, cellular oxidative stress and hyperinsulinemia lead to both activation of several serine/threonine kinases and phosphorylation of IRS1 [39]. These agents are known to negatively regulate the IRS1 functions by phosphorylation, however also via other molecular mechanisms such as suppressor of cytokine signaling proteins (SOCS) expression, IRS degradation, and O-linked glycosylation. Understanding the mechanisms of IRS1 inhibition and identication of kinases involved in these processes may reveal novel targets for development of strategies to prevent insulin resistance [4].

## Mechanisms for increased oxidative stress in diabetes

### Autoxidative glycosylation

Autoxidative glycosylation is initiated by the oxidation of an aldose or ketose to a more reactive dicarbonyl sugar, which would then react with a protein. The reduced oxygen products formed in the autoxidation reaction include superoxide, the hydrogen radical and hydrogen peroxide, which, in the presence of metal ions, would cause oxidative damage to neighboring molecules. Therefore, autoxidative glycosylation is a possible mechanism for the production of free radicals, leading to fragmentation of proteins and oxidation of associated lipids during glycosylation reactions [17,40,41]. Autoxidative glycosylation is increased in the presence of hyperglycemia.

Sequential glycation, followed by free radical-mediated oxidation, will generate early glycosylation products. Some of these products undergo further chemical rearrangements to form irreversible, advanced glycosylation end products AGEs, which accumulate with aging and duration of diabetes in long-lived structural proteins, such as collagens. The formation of AGEs can induce covalent cross linking, resulting in hardening of the blood vessel walls with loss of their elasticity and increased vascular permeability [42]. Protein glycosylation is increased in diabetes and used as an index of long-term glucose control HbA1c. It correlates with diabetic complications. AGEs bind to a cell surface receptor known as receptor for AGE (RAGE), a multiligand member of the Ig super family. This binding initiates a cascade of signal transduction events involving p44/p42 MAPKs, nuclear factor-B, p21Ras, and other intermediates [43,44]. Interaction of AGEs with RAGE induces the production of ROS through a mechanism that involves localization of prooxidant molecules at the cell surface [45] and a key role for activated NADPH oxidase [46]. In neuronal cell lines, application of AGEs depletes GSH, but this is prevented in the presence of antioxidants [47]. Antioxidants or antibodies against RAGE prevent both oxidative stress and the downstream signaling pathways that can be activated by ligation of RAGE. AGE-mediated ROS production is particularly implicated in blood vessel endothelial activation and diabetic vascular complications [48,49].

# Lipid peroxidation

Oxygen radicals catalyse the oxidative modification of lipids. This peroxidation chain reaction is illustrated in Fig 2. Lipid peroxy radicals react with other lipids, proteins, and nucleic acids; propagating thereby the transfer of electrons and bringing about the oxidation of substrates. Cell membranes, which are structurally made up of large amounts of PUFA, are highly susceptible to oxidative attack and, consequently, changes in membrane fluidity, permeability, and cellular metabolic functions result.



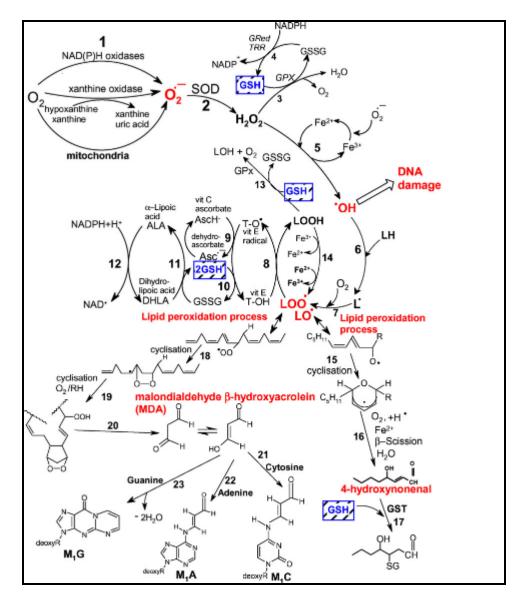


Fig 2: Pathways of ROS formation, the lipid peroxidation process and the role of glutathione (GSH) and other antioxidants (Vitamin E, Vitamin C, lipoic acid) in the management of oxidative stress. (Valko et al., 2007)

Reaction 1: The superoxide anion radical is formed by the process of reduction of molecular oxygen mediated by NAD(P)H oxidases and xanthine oxidase or non-enzymatically by redox reactive compounds such as the semi-ubiquinone compound of the mitochondrial electron transport chain. Reaction 2: Superoxide radical is dismutated by the superoxide dismutase (SOD) to hydrogen peroxide. Reaction 3: Hydrogen peroxide is most efficiently scavenged by the enzyme glutathione peroxidase (GPx) which requires GSH as the electron donor. Reaction 4: The oxidised glutathione is reduced back to GSH by the enzyme glutathione reductase (Gred) which uses NADPH as the electron donor. Reaction 5: Some transition metals (e.g.  $Fe^{2+}$ ,  $Cu^+$  and others) can breakdown hydrogen peroxide to the reactive hydroxyl radical (Fenton reaction). Reaction 6: The hydroxyl radical can abstract an electron from polyunsaturated fatty acid (LH) to give rise to a carbon-centred lipid radical (L<sup>•</sup>). Reaction 7: The lipid radical (L<sup>•</sup>) can further interact with molecular oxygen to give a lipid peroxyl radical (LOO<sup>•</sup>). If the resulting lipid peroxyl radical LOO<sup>•</sup> is not reduced by antioxidants, the lipid peroxidation process occurs (reactions 18-23 and 15-17). Reaction 8: The lipid peroxyl

radical (LOO<sup>•</sup>) is reduced within the membrane by the reduced form of Vitamin E (T-OH) resulting in the formation of a lipid hydroperoxide and a radical of Vitamin E (T-O<sup>•</sup>). Reaction 9: The regeneration of Vitamin E by Vitamin C: the Vitamin E radical (T-O<sup>•</sup>) is reduced back to Vitamin E (T-OH) by ascorbic acid (the physiological form of ascorbate is ascorbate monoanion, AscH<sup>–</sup>) leaving behind the ascorbyl radical (Asc<sup>–</sup>). Reaction 10: The regeneration of Vitamin E by GSH: the oxidised Vitamin E radical (T-O<sup>•</sup>) is reduced back to GSH and ascorbate monoanion, AscH<sup>–</sup>, respectively, by the dihydrolipoic acid (DHLA) which is itself converted to  $\alpha$ -lipoic acid (ALA). Reaction 12: The regeneration of DHLA from ALA using NADPH. Reaction 13: Lipid hydroperoxides are reduced to alcohols and dioxygen by GPx using GSH as the electron donor.

*Lipid peroxidation process*: Reaction 14: Lipid hydroperoxides can react fast with  $Fe^{2+}$  to form lipid alkoxyl radicals (LO•), or much slower with  $Fe^{3+}$  to form lipid peroxyl radicals (LOO•). Reaction 15: Lipid alkoxyl radical (LO•) derived for example from arachidonic acid undergoes cyclisation reaction to form a six-membered ring hydroperoxide. Reaction 16: Six-membered ring hydroperoxide udergoes further reactions to from 4-hydroxy-nonenal. Reaction 17: 4-hydroxynonenal is rendered into an innocuous glutathiyl adduct (GST, glutathione S-transferase). Reaction 18: A peroxyl radical located in the internal position of the fatty acid can react by cyclisation to produce cyclic peroxide adjacent to a carbon-centred radical. Reaction 19: This radical can then either be reduced to form a hydroperoxide (reaction not shown) or it can undergo a second cyclisation to form bicyclic peroxide which after coupling to dioxygen and reduction yields a molecule structurally analogous to the endoperoxide. Reactions 21, 22, 23: Malondialdehyde can react with DNA bases Cytosine, Adenine, and Guanine to form adducts M1C, M1A and M1G, respectively [4].

### The polyol pathway

The enzyme aldose reductase converts toxic aldehydes to inactive alcohols [50]. Glucose is a poor substrate for aldose reductase, but at high concentrations this enzyme converts glucose to sorbitol, initiating the polyol pathway of glucose conversion to fructose. Similar to GSH reductase, the enzyme aldose reductase is dependent upon NADPH as a cofactor. Therefore, excessive activation of the polyol pathway depletes cytosolic NADPH and subsequently depletes GSH, leaving the cell vulnerable to free radicals produced during normal cellular functions such as electron transfer. In addition, accumulation of sorbitol produces a cellular osmotic stress that also generates oxidative stress [51]. This pathway has been a target for therapies against diabetes complications including neuropathy. Recent human genetic and biochemical data link polymorphisms of the aldose reductase gene to increased risk of diabetic complications, with the principal allele associated with increased disease risk causing a 2- to 3-fold increase in aldose reductase gene expression [52].

# Protein kinase C (PKC) activation

Prooxidants react with the regulatory domain to stimulate PKC activity, but antioxidants react with the catalytic domain of PKC and inhibit its activity [53]. Activity of PKC is increased in the retina, kidney, and microvasculature of diabetic rats, but there is no evidence for altered activity of any of the PKC isoforms in the peripheral neurons [54,55]. This suggests that the lipolytic pathway and production of diacylglycerol are the main causes of PKC activation in nonneuronal cell types [56]. Once activated, PKC activates the MAPKs that phosphorylate transcription factors and thus alter the balance of gene expression [57]. Specifically, it is the stress genes such as heat shock proteins and c-Jun kinases that increase

after PKC activation and can lead to apoptosis or vascular atherosclerosis. A role for PKC in inducing neuronal degeneration possibly at the level of the endothelial cell is implicated by three studies. Inhibition of PKCß reduces oxidative stress and normalizes blood flow and nerve conduction deficits in diabetic rats [56,58].High glucose causes nuclear factor-B activation in endothelial cells, leading to ROS formation, and cellular activation, an effect that is prevented in the presence of a PKC inhibitor [59].

### **Reduced anti-oxidant defense**

In addition to the increased generation of free radicals in diabetes, impaired generation of naturally occurring antioxidants also result in increased oxidative injury by failure of protective mechanisms [60]. Antioxidant defense system appears to be compromised in diabetic patients. It has been demonstrated that reduced scavenging of free radicals by SOD [61] and lake of GSH [62] and ascorbic acid [63] are associated with diabetic vascular pathology. Reduced other antioxidants, such as vitamin A and E, uric acid, reduced activity of catalase and GPx are also found in diabetes [64-66]. The mechanism by which the antioxidant reserve is reduced is not clear. Protein damage due to the protein glycosylation may be a mechanism that lowers the activities of primary antioxidant enzymes [61]. In addition, GSH deficiency may result from depletion of NADH in polyol pathway [67].

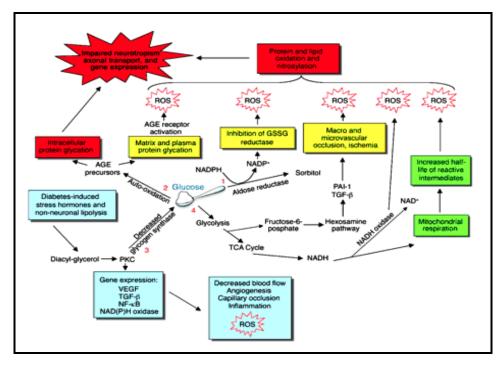
### Oxidative stress and diabetic complications Diabetic nephropathy

Hyperglycemia leads to increased accumulation of AGEs in the kidney, which in part, via enhanced free radical mechanisms, induces cross linking of proteins and membrane peroxidation. This results in increased membrane permeability and loss of integrity. Hyperglycemia-induced secondary mediator's activation such as protein kinase-C (PKC), mitogen-activated protein (MAP) kinases and cytokine lead to the production of predominantly vasodilatory prostaglandins. The increased glomerular permeability and glomerular blood flow initiate and maintain the early proteinuria and glomerular hyperfiltration which are also responsible for oxidative stress-induced renal injury in the diabetic condition [68]. In the late stage of diabetes, prostaglandines (F2 isoprostanes), formed by free radical catalysed lipid peroxidation, cause progressive vasoconstriction and systemic hypertension. The continued deposition of extracellular matrix proteins in the glomeruli and the reduced blood flow eventually lead to glomerulosclerosis and renal failure [69].

A causal relationship between oxidative stress, ECD and diabetic nephropathy has been established [70] by observations that: a) High glucose can directly cause endothelial cell dysfunction (ECD) and increases oxidative stress in glomerular mesangial cells, a target cell of diabetic nephropathy. b) Lipid peroxides and 8-hydroxydeoyguanosine, indices of oxidative tissue injury, were increased in the kidneys of diabetic patient with albuminuria. The oxidized lipids are toxic to tissues especially the vascular endothelium, glomerular mesangial cells, smooth muscle cells and renal tubular epithelium. c) Oxidative stress induces mRNA expression of NFkB genes which in turn promotes production of proinflammatory proteins-TGF-B, fibronection, laminin, elastin, IL-1, IL-6, and PDGF, and d) Inhibition of oxidative stress ameliorates all the manifestation associated with ECD and diabetic nephropathy [71-74].

#### **Diabetic neuropathy**

One of the mechanisms by which hyperglycemia causes neural degeneration is via the increased oxidative stress that accompanies diabetes. Metabolic and oxidative insults often cause rapid changes in glial cells. Key indicators of this response are increased synthesis of glial fibrillary acidic protein (GFAP) and S100B, both astrocytic markers [75]. Hyperglycemia activates many signaling mechanisms in cells. Four major pathways that can lead to cell injury downstream of hyperglycemia are illustrated in fig 3.



### Fig 3: Four major pathways that can lead to diabetic neuropathy.

1) Excess glucose shunts to the polyol pathway that depletes cytosolic NADPH and subsequently GSH. In the development of neuropathy, the hyperglycemic state leads to an increase in action of the enzymes aldose reductase and sorbitol dehydrogenase. This results in the conversion of intracellular glucose to sorbitol and fructose.

The accumulation of these sugar products results in a decrease in the synthesis of nerve cell myoinositol, required for normal neuron conduction. Additionally, the chemical conversion of glucose results in a depletion of nicotinamide adenine dinucleotide phosphate stores, which are necessary for the detoxification of reactive oxygen species and for the synthesis of the vasodilator nitric oxide. There is a resultant increase in oxidative stress on the nerve cell and an increase in vasoconstriction leading to ischemia, which will promote nerve cell injury and death. Hyperglycemia and oxidative stress also contribute to the abnormal glycation of nerve cell proteins and the inappropriate activation of protein kinase C, resulting in further nerve dysfunction and ischemia [76-79]. 2) Excess glucose also undergoes autooxidation to produce AGEs that impair protein function and also activate RAGEs that use ROS as second messengers. 3) PKC activation both further increases hyperglycemia and also exacerbates tissue hypoxia. 4) Overload and slowing of the electron transfer chain leads to escape of reactive intermediates to produce  $O_2^{-}$  as well as activation of NADH oxidase that also produces  $O_2^{-}$ . A unifying mechanism of injury in each case is the production of ROS that impair protein and gene function [80].

## **Diabetic retinopathy**

Diabetic retinopathy develops in patients with both type 1 and type 2 diabetes and is the major cause of vision loss and blindness in the working population. The main risk factor of diabetic retinopathy is hyperglycemia accompanied by enhanced mitochondrial production of reactive oxygen species and oxidative stress, formation of advanced glycation end products (AGE) and hexosamines, increased polyol metabolism of glucose. The severity of vascular injury depends on the individual genetic background and is modified by other metabolic and haemodynamic factors influencing numbers of intracellular signalling molecules such as PKC, MAPK or NF-kappaB. In diabetes, damage to the retina occurs in the vasculature (endothelial cells and pericytes), neurons and glia, pigment epithelial cells and infiltrating immunocompetent cells: monocytes, granulocytes, lymphocytes [81]. The retina has high content of polyunsaturated fatty acids and has the highest oxygen uptake and glucose oxidation relative to any other tissue. This phenomenon renders retina more susceptible to oxidative stress [82]. It has been suggested that the correlation between hyperglycemia, changes in the redox homeostasis, and oxidative stress are the key events in the pathogenesis of diabetic retinopathy. Animal studies have demonstrated that oxidative stress contributes not only to the development of diabetic retinopathy but also to the resistance of retinopathy to reverse after good glycemic control is reinstituted the metabolic memory phenomenon [83]. Resistance of diabetic retinopathy to reverse is probably attributed to accumulation of damaged molecules and ROS that are not easily removed even after good glycemic control is reestablished. Superoxide and hydrogen peroxide levels are elevated in the retina of diabetic rats and in retinal cells incubated in high glucose media [83-86]. Membrane lipid peroxidation and oxidative damage to DNA (indicated by 8-hydroxy-2'-deoxyguanosine, 8-OHdG), the consequences of ROS induced injury, are elevated in the retina in diabetes [87-90]. In diabetes, the activities of antioxidant defense enzymes responsible for scavenging free radicals and maintaining redox homeostasis such as SOD, glutathione reductase, glutathione peroxidase, and catalase are diminished in the retina [91]. The levels of GSH intracellular antioxidant are decreased in the retina in diabetes [92], and the enzymes responsible for its metabolism are compromised [93,94]. Apart from the antioxidant defense enzymes, non- enzymatic antioxidants such as vitamin C, vitamin E, and β-carotene that exist biologically for the regulation of redox homeostasis are also depressed during hyperglycemia induced oxidative stress [95].

Diabetic cataract is a major complication of diabetes mellitus, and is primarily caused by polyol accumulation and glycation within lens fibers and the epithelium. Blood sugar can passively diffuse into lens tissue insulin independently and then be converted by aldose reductase to polyols, which can not diffuse passively out of the lens, and thus they accumulate. This accumulation of polyols causes osmotic changes, which lead to lens hydration and swelling that are followed by biochemical and physiological damage to cell membranes [18].

# 5.4 Coronary heart disease (CHD)

Hyperglycaemia could aggravate CHD by several mechanisms. Hyperglycaemia alters vascular reactivity, platelet aggregation, clot formation, lysis and foam cell formation. All these factors enhance atherosclerosis. Hyperglycaemia predisposes to formation of advanced glycation end products. Increased uptake of AGE in vascular wall has shown to increase vascular permeability and decrease its vasodilatory response to nitroglycerin and acetylcholine [96]. AGE products also activate leucocytes or vessel wall cells to enhance production of oxygen metabolites that can promote lipoprotein oxidation.

Glycation of matrix proteins also increases their stability, thus promoting their accumulation, which is characteristic of hyperplastic diseases [97]. Normal endothelium synthesises substances that contribute to maintenance of vascular tone such as PGI2 and endothelium derived relaxation factor (now identified as Nitric Oxide - NO). PGI2 also inhibits platelet aggregation. Synthesis of NO is decreased and its removal increased by advanced glycation end products. Its inactivation by oxygen free radicals is also enhanced and this leads to increased vascular resistance and platelet adhesion to endothelium [97]. Decreased synthesis of PGI2 also adds to vasoconstriction and vessel wall reactivity. Enhanced activity of protein kinase C in endothelial cells might contribute to synthesis of vasoconstrictor prostanoids [97]. Impaired endothelium mediated vasodilatation could result in hypertension and shear induced platelet aggregation. Reactivity of platelets and adhesion to vessel wall is enhanced by interaction with glycated LDL and increased levels of von Willebrand factor [98]. Altered metabolism due to decreased platelet polyphosphoinositide content causes enhanced release of thromboxane B2 which leads to increased platelet aggregation. Hyperglycaemia leads to glucose autooxidation and release of free radicals. Free radicals activate platelets and moreover, oxidative stress decreases the activity of antithrombin III [99]. This coupled with increased fibrinogen levels results in enhanced fibrin clot formation. Lipoprotein (a) is known to inhibit acitivity of plasminogen. Decreased fibrinolytic activity further increases the stability of the clot formed. Lipoprotein levels are also increased in diabetics [99]. The macrophage is the precursor of the cholesterol engorged foam cell, characteristic of atherosclerotic lesions. LDL and VLDL isolated from diabetic patients are taken up by the macrophages and they induce cholesterol ester synthesis and accumulation giving rise to foam cells [100]. Hyperglycaemia and oxidative stress can increase monocyte binding to endothelial cells and thus result in accelerated atherosclerosis [101].

Insulin resistance and hyperinsulinaemia correlate strongly with VLDL-TG secretion rate and plasma TG concentration. Once the pool of VLDL-TG increases, HDL levels are decreased. The risk for Coronary artery disease (CAD) is further accentuated with increase in small dense LDL levels, which in turn is associated with insulin resistance. Another associated finding is that of postprandial lipaemia [102]. Hyperinsulinaemia inhibits the binding of HDL to its specific binding sites on human skin fibroblasts and thus retards the transfer of intracellular sterol to cell membrane and its transfer to HDL [103]. These actions may contribute to cholesterol accumulation and formation of foam cells. Plasminogen activator inhibitor-1 (PAI-1) levels vary directly with insulin resistance and are associated with recurrent myocardial infarction in younger men [99]. Hyperinsulinaemia and insulin resistance are associated with enhanced thrombus formation in response to endothelium injury: This coupled with low fibrinolytic activity accentuates the athero-thrombotic process.

#### **Dyslipidemia**

The impairment of insulin secretion results in enhanced metabolism of lipids from the adipose tissue to the plasma [104]. Increased production of very low density lipoprotein (LDL) by the liver results from increased delivery of fatty acids because of decreased utilization by muscle and increased delivery of fatty acids from visceral abdominal fat to the liver via the portal circulation. Decreased catabolism of postprandial triglyceride rich lipoprotein particles because of reduced lipoprotein lipase activity accentuates diabetic dyslipidemia [105]. The levels of serum lipids are usually elevated in diabetes mellitus and such an elevation represents the risk factor for coronary heart diseases [106].

### **Diabetic foot ulcers**

Neuropathy in diabetic patients is manifested in the motor, autonomic, and sensory components of the nervous system. Damage to the innervations of the intrinsic foot muscles leads to an imbalance between flexion and extension of the affected foot. This produces anatomic foot deformities that create abnormal bony prominences and pressure points, which gradually cause skin breakdown and ulceration. Autonomic neuropathy leads to a diminution in sweat and oil gland functionality. As a result, the foot loses its natural ability to moisturize the overlying skin and becomes dry and increasingly susceptible to tears and the subsequent development of infection. The loss of sensation as a part of peripheral neuropathy exacerbates the development of ulcerations. As trauma occurs at the affected site, patients are often unable to detect the insult to their lower extremities. As a result, many wounds go unnoticed and progressively worsen as the affected area is continuously subjected to repetitive pressure and shear forces from ambulation and weight bearing [107,108]. Neuropathy, mechanical stress, and macrovascular disease are involved in the pathogenesis of diabetic foot ulceration. Implicit in the development of gangrene and ulceration is the recognition that these factors interact with the microcirculation, resulting in the failure of skin capillary flow to meet nutritive requirements. These late functional abnormalities include loss of autoregulation and reduced hyperaemic responses which interact with loss of neurogenic flow regulation, disturbed endothelial function, and abnormal rheology to produce the familiar clinical picture of the diabetic foot [109].

## Peripheral vascular disease

Peripheral vascular disease is a contributing factor to the development of foot ulcers in up to 50% of cases [110,111]. It commonly affects the tibial and peroneal arteries of the extremities. Endothelial cell dysfunction and smooth cell abnormalities develop in peripheral arteries as a consequence of the persistent hyperglycemic state [76]. There is a resultant decrease in endothelium-derived vasodilators leading to constriction. Further, the hyperglycemia in diabetes is associated with an increase in thromboxane A2, a vasoconstrictor and platelet aggregation agonist, which leads to an increased risk for plasma hypercoagulability [112]. Platelet function is abnormal in diabetes as well. Expression of both glycoprotein Ib and IIb/IIIa is increased, augmenting both platelet von Willebrand factor and platelet-fibrin interaction [113]. The intracellular platelet glucose concentration mirrors the extracellular environment and is associated with increased superoxide anion formation and PKC activity and decreased platelet-derived NO [113,114]. Hyperglycemia further changes platelet function by impairing calcium homeostasis and thereby alters aspects of platelet activation and aggregation, including platelet conformation and release of mediators [115]. In diabetes, plasma coagulation factors (eg, factor VII and thrombin) and lesion-based coagulants (eg, tissue factor) are increased, and endogenous anticoagulants (eg, thrombomodulin and protein C) are decreased [116-118]. Also, the production of plasminogen activator inhibitor-1, a fibrinolysis inhibitor, is increased [113, 116, 119-125]. Thus, a propensity for platelet activation and aggregation, coupled with a tendency for coaulation, is relevant to a risk of thrombosis. There is also the potential for alterations in the vascular extracellular matrix leading to stenosis of the arterial lumen [112]. Moreover, smoking, hypertension, and hyperlipidemia are other factors that are common in diabetic patients and contribute to the development of Peripheral arterial disease [126]. Cumulatively, this leads to occlusive arterial diseases that result in ischemia in the lower extremity and an increased risk of ulceration in diabetic patients.

#### **Diabetic ketoacidosis**

During the normal fed state, an increase in glucose concentration stimulates the pancreatic beta cells to secrete insulin and inhibits the pancreatic alpha cell production of glucagon. This increase in insulin and decrease in glucagon stimulates the liver to undergo glycolysis and glycogenesis of the ingested carbohydrates. The body is able to maintain the blood glucose levels within a very narrow range during both the fasting and fed state by increasing and decreasing the concentration of insulin. In diabetic ketoacidosis there is a decrease in the ratio of insulin to glucagon as well as an increase in the concentrations of the counter regulatory hormones. This decreased insulin and increased glucagons levels impairs entry of glucose into cells, which results in gluconeogenesis, glycogenolysis, and the breakdown of triglycerides (lipolysis) causing a complex metabolic disturbance of carbohydrate, protein, and lipid metabolism. As the glucose concentration and osmolality of extra cellular fluid increases, an osmolar gradient is created that draws water out of the cells leading to osmotic diuresis. With continued osmotic diuresis, hypovolemia eventually occurs which leads to a progressive decline in glomerular filtration rate, and hyperglycemia becomes more severe [127].

The counter regulatory hormones play a key role by triggering pathways that are active when inadequate insulin is in the liver and peripheral tissues. This results in excessive lipolysis leading to the accumulation of free fatty acids. These free fatty acids are used for the formation of large amounts of acetyl Co A. This excess of acetyl Co A cannot enter the citric acid cycle, but is used for ketone body formation instead. Most of the acid produced in normal metabolism is in the form of carbon dioxide which is readily excreted from the lungs. In contrast ketone bodies cannot be excreted through the pulmonary system, which leads to metabolic acidosis [128,129].

#### Antioxidants

'Antioxidants' are substances that neutralize free radicals or their actions [130].Nature has endowed each cell with adequate protective mechanisms against any harmful effects of free radicals: superoxide dismutase (SOD), glutathione peroxidase, glutathione reductase, thioredoxin, thiols and disulfide bonding are buffering systems in every cell.  $\alpha$ -Tocopherol (vitamin E) is an essential nutrient which functions as a chain-breaking antioxidant which prevents the propagation of free radical reactions in all cell membranes in the human body. Ascorbic acid (vitamin C) is also part of the normal protecting mechanism. Other non-enzymatic antioxidants include carotenoids, flavonoids and related polyphenols,  $\alpha$ -lipoic acid, glutathione etc.

Antioxidants, capable of neutralizing free radicals or their actions, act at different stages. They act at the levels of prevention, interception and repair. Preventive antioxidants attempt to stop the formation of ROS. These include superoxide dismutase that catalyses the dismutation of superoxide to H<sub>2</sub>O<sub>2</sub> and catalase that breaks it down to water [130,131]. Interception of free radicals is mainly by radical scavenging, while at the secondary level scavenging of peroxyl radicals are effected. The effectors include various antioxidants like vitamins C and E, glutathione, other thiol compounds, carotenoids, flavonoids, etc. At the repair and reconstitution level, mainly repair enzymes are involved [132, 130, 131,133].

#### Conclusion

The molecular mechanisms whereby oxidative stress causes insulin resistance are undefined. In a variety of tissues, hyperglycaemia and elevated free fatty acids result in the generation of ROS and RNS, leading to increased oxidative stress. In the absence of an appropriate compensatory response from the endogenous antioxidant network, the system becomes overwhelmed (redox imbalance), leading to the activation of stress-sensitive signaling pathways, such as NF-kB, MAPK, JNK/SAPK, PKC, AGE/ RAGE, sorbitol, and others. The consequence is the production of gene products such as VEGF and others that cause cellular damage, and are ultimately responsible for the long-term complications of diabetes. In addition, activation of the same or similar pathways appears to mediate insulin resistance and impaired insulin secretion. It is our view that there appears to be a common biochemical basis that involves oxidative-stress induced activation of stress sensitive signaling pathways.

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#### References

- 1) Halliwell B, Gutteridge JMC. Free radicals in biology and medicine. 4th ed. UK: Oxford Clarendon Press; 2007.
- 2) Bahorun T, Soobrattee MA, Luximon-Ramma, V Aruoma OI. Free radicals and antioxidants in cardiovascular health and disease. Inter J Med Upd 2006; 11-17.
- 3) Valko M, Izakovic M, Mazur M, Rhodesm CJ, Telser J. Role of oxygen radicals in DNA damage and cancer incidence. Mol Cell Biochem J 2004; 266:37-56.
- 4) Valko M, Leibfritz D, Moncola J, Cronin MD et al. Free radicals and antioxidants in normal physiological functions and human disease Review. Int J Biochem Cell Bio 2007; 39:44-84.
- 5) Droge W. Free radicals in the physiological control of cell function Review. Physiolog Rev 2002; 82: 47-95.
- 6) Willcox JK, Ash SL, Catignani GL. Antioxidants and prevention of chronic disease Review. Crit Rev Food Sci Nutr J 2004; 44:275-295.
- 7) Pacher P, Beckman JS, Liaudet L. Nitric oxide and peroxynitrite in health and disease. Physiol Rev 2007; 87:315-424.
- 8) Genestra M. Oxyl radicals redox-sensitive signalling cascades and antioxidants Review. Cell Signal 2007; 19:1807-1819.
- 9) Halliwell B. Biochemistry of oxidative stress. Biochem Soc Trans 2007; 35:1147-1150.
- 10) Young I, Woodside J. Antioxidants in health and disease. J Clin Path 2001; 54:176-186.
- 11) Valko M, Morris H, Cronin MTD. Metals toxicity and oxidative stress. Curr Med Chem 2005; 12:1161-1208.
- 12) Valko M, Rhodes CJ, Moncol J, Izakovic M et al. Free radicals metals and antioxidants in oxidative stress-induced cancer Mini-review. Chem Bio Inter 2006; 160: 1-40.
- 13) Parthasarathy S, Santanam N, Ramachandran S, and Meilhac O. Oxidants and antioxidants in atherogenesis: An appraisal. J Lip Res 1999; 40:2143-2157.

- 14) Frei B. Reactive oxygen species and antioxidant vitamins Linus Pauling Institute Oregon State University. 1997; http://lpioregonstateedu/ f-w97/reactivehtml.
- 15) Chatterjee M, Saluja R, Kanneganti S et al. Biochemical and molecular evaluation of neutrophil NOS in spontaneously hypertensive rats. J Cell Mol Bio 2007; 53:84-93.
- 16) Betteridge DJ. What is oxidative stress? Metabolism 2000; 492(1):3-8.
- 17) Baynes JW. Role of oxidative stress in development of complications indiabetes. Diabetes 1991; 40: 405-412.
- 18) Abdollahi M, Rahimi R, Nikfar S, Larijani B. A review on the role of antioxidants in the management of diabetes and its complications Dossier: Antioxidants in the prevention of human diseases. Biomed Pharmacoth 2005; 59:365-373.
- 19) Penckofer S, Schwertz D, Florczak K. Oxidative stress and cardiovascular disease in type 2 diabetes: the role of antioxidants and prooxidants. J Cardiovasc Nur 2002; 16(2):68-85.
- 20) Baynes JW, Thorpe SR. Role of oxidative stress in diabetic complications: a new perspective on an old paradigm. Diabetes 1999; 48:1-9.
- 21) Unger RH, Grundy S. Hyperglycaemia as an inducer as well as a consequence of impaired islet cell function and insulin resistance: implications for the management of diabetes. Diabetologia 1985; 28:119-121.
- 22) Unger RH. Lipotoxicity In Diabetes Mellitus: A fundamental and Clinical Text. 3rd ed. ited by LeRoith D OJ Taylor S: Lippincott Williams & Wilkins. 2004;
- 23) Vlassara H, Bucala R Striker L. Pathogenetic effects of advanced glycosylation: Biochemical biologic and clinical implications of diabetes and ageing Lab Investigation 1994; 70:138-151
- 24) Brownlee M. Advanced protein glycosylation in diabetes and aging. Ann Rev Med 1996; 46:223-234.
- 25) Robertson RP. Chronic oxidative stress as a central mechanism for glucose toxicity in pancreatic islet beta cells in diabetes. J Bio Chem 2004; 279:42351-42354.
- 26) Briaud I, Harmon JS, Kelpe CL, Segu VB, Poitout V. Lipotoxicity of the pancreatic betacell is associated with glucosedependent esterification of fatty acids into neutral lipids. Diabetes 2001; 50:315-321.
- 27) Prentki M, Corkey BE. Are the beta-cell signaling molecules malonyl-CoA and cystolic long-chain acyl-CoA implicated in multiple tissue defects of obesity and NIDDM? Diabetes 1996; 45:273-283.
- 28) Poitout V, Robertson RP. Minireview: secondary beta-cell failure in type 2 diabetes–a convergence of glucotoxicity and lipotoxicity. Endocrinol 2002; 143:339-342.
- 29) Olson LK, Sharma A, Peshavaria M, Wright CV, Towle HC, Robertson RP et al. Reduction of insulin gene transcription in HIT-T15 beta cells chronically exposed to a supraphysiologic glucose concentration is associated with loss of STF-1 transcription factor expression. Proc Nat Aca Sci USA 1995; 92:9127-9131.
- 30) Poitout V, Olson LK, Robertson RP. Chronic exposure of betaTC-6 cells to supraphysiologic concentrations of glucose decreases binding of the RIPE3b1 insulin gene transcription activator. J Clin Invest 1996; 97:1041-1046.
- 31) Harmon JS, Stein R, Robertson RP. Oxidative stress-mediated post-translational loss of MafA protein as a contributing mechanism to loss of insulin gene expression in glucotoxic beta cells. Bio Chem 2005; 280:11107-11113.
- 32) Tanaka Y, Gleason CE, Tran PO, Harmon JS, Robertson RP. Prevention of glucose toxicity in HIT-T15 cells and Zucker diabetic fatty rats by antioxidants. Proc Nat Acad Sci USA 1999; 96:10857-10862.
- 33) Kaneto H, Kajimoto Y, Miyagawa J, Matsuoka T, Fujitani Y, Umayahara Y et al. Beneficial effects of antioxidants in diabetes: possible protection of pancreatic beta-cells against glucose toxicity. Diabetes 1999; 48:2398-2406

- 34) Kajimoto Y, Kaneto H. Role of Oxidative Stress in Pancreatic B-Cell Dysfunction. Ann New York Acad Sci 2004; 1011:168-176.
- 35) Grankvist K, Marklund SL, Taljedal IB. CuZn superoxide dismutase Mn-superoxide dismutase catalase and glutathione-peroxidase in pancreatic-islets and other tissues in the mouse. Biochem 1981; 393-398.
- 36) Malaisse WJ, Malaisse-Lagae F, Sener A, Pipeleers DG. Determinants of the selective toxicity of alloxan to the pancreatic β-cells. Proc Nat Acad Sci USA 1982; 79: 927-930.
- 37) White MF. The IRS-signalling system: Anetwork of docking proteins that mediate insulin action. Mol Cell Biochem 1998;182:3-11.
- 38) Lawlor MA, Alessi DR. PKB/Akt: A key mediator of cell proliferation survival and insulin responses? J Cell Sci 2001; 114:2903-2910.
- 39) Vicent D, Ilany J, Kondo T, Naruse K, Fisher SJ, Kisanuki YY et al. The role of endothelial insulin signaling in the regulation of vascular tone and insulin resistance. J Clin Invest 2003: 111:1373-1380.
- 40) Hunt JV, Smith CCT, Wolff SP. Autoxidative glycosylation and possible involvement of peroxides and free radicals in LDL modification by glucose. Diabetes 1990; 39:1420-1424.
- 41) Ceriello A, Quatraro A, Giugliano D. New insights on non-enzymatic glycosylation may lead to therapeutic approaches for prevention of diabetic complications. Diab Med 1992; 9:297-299.
- 42) Vlassara H. Recent progress in advanced glycation end products and diabetic complications. Diabetes 1997; 46(2): S19-S25.
- 43) Lander HM, Tauras JM, Ogiste JS, Hori O, Moss RA, Schmidt AM. Activation of the receptor for advanced glycation end products triggers a p21ras;-dependent mitogenactivated protein kinase pathway regulated by oxidant stress. J Bio Chem 1997; 272:17810-17814.
- 44) Wautier JL, Wautier MP, Schmidt AM, Anderson GM, Hori O, Zoukourian C et al. Advanced glycation end products AGEs; on the surface of diabetic erythrocytes bind to the vessel wall via a specific receptor inducing oxidant stress in the vasculature: a link between surface-associated AGEs and diabetic complications. Proc Nat Acad Sci USA 1994; 91:7742-7746.
- 45) Yan SD, Schmidt AM, Anderson GM, Zhang J, Brett J, Zou YS et al. Enhanced cellular oxidant stress by the interaction of advanced glycation endproducts with their receptors/binding proteins. J Bio Chem 1994; 269:9889-9897.
- 46) Wautier MP, Chappey O, Corda S, Stern DM, Schmidt AM, Wautier JL. Activation of NADPH oxidase by AGE links oxidant stress to altered gene expression via RAGE. Am J Physiol 2001; 280:E685-E694.
- 47) Deuther-Conrad W, Loske C, Schinzel R, Dringen R, Riederer P, Munch G. Advanced glycation endproducts change glutathione redox status in SH-SY5Y human neuroblastoma cells by a hydrogen peroxide dependent mechanism. Neurosci Lett 2001; 312: 29-32.
- 48) Cameron NE, Cotter MA. Effects of antioxidants on nerve and vascular dysfunction in experimental diabetes. Diab Res Clin Pract 1999; 45:137-146.
- 49) Mullarkey CJ, Edelstein D, Brownlee M. Free radical generation by early glycation products: a mechanism for accelerated atherogenesis in diabetes. Biochem Biophys Res Comm 1990; 173:932-939.
- 50) Greene DA, Sima AA, Stevens MJ, Feldman EL, Lattimer SA. Complications: neuropathy pathogenetic considerations. Diabetes Care 1992; 15:1902-1925.

- 51) Stevens MJ, Lattimer SA, Kamijo M, VanHuysen C, Sima AAF, Greene DA. Osmotically-induced nerve taurine depletion and the compatible osmolyte hypothesis in experimental diabetic neuropathy in the rat. Diabetologia 1993; 36:608-614.
- 52) Oates PJ, Mylari BL. Aldose reductase inhibitors: therapeutic implications for diabetic complications. Ex Opi Invest Dr 1999; 8:2095-2119.
- 53) Gopalakrishna R, Jaken S. Protein kinase C signaling and oxidative stress. Free Rad Bio Med 2000; 28:1349-1361.
- 54) Craven PA, DeRubertis FR. Protein kinase C is activated in glomeruli from streptozotocin diabetic rats Possible mediation by glucose. J Clin Invest 1989; 83:1667-1675.
- 55) Lee TS, Saltsman KA, Ohashi H, King GL. Activation of protein kinase C by elevation of glucose concentration: proposal for a mechanism in the development of diabetic vascular complications. Proc Nati Acad Sci USA 1989; 86:5141-5145.
- 56) Ishii H, Koya D, King GL. Protein kinase C activation and its role in the development of vascular complication in diabetes mellitus. Mol Med 1998; 76:21-31.
- 57) Tomlinson DR. Mitogen-activated protein kinases as glucose transducers for diabetic complications. Diabetologia 1999; 42:1271-1281.
- 58) Cameron NE, Cotter MA. Effects of protein kinase CB inhibition on neurovascular dysfunction in diabetic rats: interaction with oxidative stress and essential fatty acid dysmetabolism. Diab Meta Res Rev 2002;18:315-323.
- 59) Srivastava AK. High glucose-induced activation of protein kinase signaling pathways in vascular smooth muscle cells: a potential role in the pathogenesis of vascular dysfunction in diabetes review. Int J Mol Med 2002; 9:85-89.
- 60) Glugliano D, Eriello A, Paolosso G. Oxidative stress and diabetic vascular complications. Diabetes Care 1996; 19:257-267.
- 61) Arai K, Lizuka S, Tada Y, Oikawa K, Taniguchi N. Increase in the glycosylated form of erythrocyte CuZnSOD in diabetes and close association of non-enzymatic glycosylation with enzyme activity. Biochim Biophys Acta 1987; 924:292-296.
- 62) Chari SW, Nath N, Rathi AB. Glutathione and its redox system in diabetic polymorphonuclear leukocytes. Am J Med Sci 1984; 287:14-15
- 63) Jennings PE, Chirico S, Jones AF. Vitamin C metabolites and microangiopathy in diabetes mellitus. Diab Res 1987; 6:151-154.
- 64) Sinclair AJ, Lunec J, Girling AJ, Barnett AH. Modulators of free radical activity in diabetes mellitus: role of ascorbic acid. EXS 1992; 62:342-352.
- 65) Maxwell SRJ, Thomason H, Sandler D et al. Antioxidant status in patients with uncomplicated insulin-dependent and non-insulin dependent diabetes mellitus. Eur J Clin Invest 1997; 27:484-490.
- 66) Tsai EC, Hirsch IB, Brunzell JD, Chait A. Reduced plasma peroxyl radical trapping capacity and increased susceptibility of LDL to oxidation in poorly controlled IDDM. Diabetes 1994; 43:1010-1014.
- 67) Giugliano D, Ceriello A, Paolisso G. Diabetes Mellitus hypertension and cardiovascular disease: which role for oxidative stress. Metabolism 1995: 44:363-368.
- 68) Anjanevulu M, Chopra K. Nordihydroguairetic acid a lignin prevents oxidative stress and the development of diabetic nephropathy in rats. Pharmacol 2004; 72: 42-50.
- 69) Salahudeen AK, Kanji V, Reckelhoff JF, Schmidt AM. athogenesis of diabetic nephropathy: a radical approach. Nephrol Dial Transpl 1997; 12:664-668.
- 70) Ha H, Kin KH. Pathogenesis of diabetic nephropathy: the role of oxidative stress and protein kinase C. Diab Res Clin Pract 1999; 5(2):147-151.
- 71) Nuhad I, Bryan B, Piotr S, Eberhard R. Perspective in renal medicine: renal disease and hypertension in non-insulin dependent diabetes mellitus. Kid Int 1999; 55:1-28.

- 72) Wardle EN. How does hyperglycaemia predispose to diabetes nephropathy? QJM<u>Int J</u> <u>Med</u> 1996; 89:943-951.
- 73) Hiragushi K, Sugimoto H, Shikata K et al. Nitric oxide system is involved in glomerular hyperfiltration in Japanese normo- and micro-albuminuric patients with type 2 diabetes mellitus. Diab Res Clin Pract 2001; 53:149-159.
- 74) Dunlop M. Aldose reductase and the role of the polyol pathway in diabetic nephropathy. Kidney Int 2000; 77:S3-S12.
- 75) Baydas G, Reiter RJ, Yaser A, Tuzcu M, Akdemir I, Nedzvetskii V. Melatonin produces glial reactivity in the hippocampus cortex and cerebellum of streptozocin-induced diabetic rats. Free Rad Bio Med 2003; 35(7):797-804.
- 76) Zochodone DW. Diabetic polyneuropathy: an update. Curr Opi Neur 2008; 21:527-533.
- 77) Feldman EL, Russell JW, Sullivan KA, Golovoy D. New insights into the pathogenesis of diabetic neuropathy. Curr Opin Neurolog 1999; 5:553-563.
- 78) Simmons Z, Feldman E. Update on diabetic neuropathy Current Opinion in Neurology 2002; 15:95-603
- 79) Boulton AJ, Armstrong DG, Albert SF, Frykberg RG, Hellman R, Kirkman MS et al. Comprehensive foot examination and risk assessment. Diabetes Care 2008; 31:1679-1685.
- 80) Feldman EL. Oxidative stress and diabetic neuropathy: a new understanding of an old problem. J Clin Invest 2003; 111:431-433.
- 81) Pelikánová T. Pathogenesis of diabetic retinopathy. Vnitr Lek 2007; 53(5):498-505.
- 82) Anderson RE, Rapp LM, Wiegand RD. Lipid peroxidation and retinal degeneration. Cur Eye Res 1984; 31:223-227.
- 83) Kowluru RA, Abbas SN. Diabetes-induced mitochondrial dysfunction in the retina. Invest Ophthalmol Visu Sci 2003; 44(12): 5327-5334.
- 84) Du Y, Miller CM, Kern TS. Hyperglycemia increases mitochondrial superoxide in retina and retinal cells. Free Rad Bio and Med 2003; 35(11):1491-1499.
- 85) Cui Y, Xu X, Bi H et al. Expression modification of uncoupling proteins and Mn SOD in retinal endothelial cells and pericytes induced by high glucose: the role of reactive oxygen species in diabetic retinopathy. Exp Eye Res 2006; 83(4): 807-816.
- 86) Ellis EA, Guberski DL, Somogyi-Mann M, Grant MB. Increased H2O2 vascular endothelial growth factor and receptors in the retina of the BBZ/WOR diabetic rat. Free Rad Bio Med 2000; 28 (1):91-101.
- 87) Kowluru RA. Effect of reinstitution of good glycemic control on retinal oxidative stress and nitrative stress in diabetic rats. Diabetes 2003; 52(3): 818-823.
- 88) Kowluru RA, Koppolu P Termination of experimental galactosemia in rats and progression of retinal metabolic abnormalities. Invest Ophthalmol Visu Sci 2002; 43(10): 3287-3291.
- 89) Kowluru RA, Tang J, and Kern TS. Abnormalities of retinal metabolism in diabetes and experimental galactosemia VII Effect of long-term administration of antioxidants on the development of retinopathy. Diabetes 2001; 50(8): 1938-1942.
- 90) Kowluru RA. Diabetes-induced elevations in retinal oxidative stress protein kinase C and nitric oxide are interrelated. Acta Diabetolog 2001; 38(4): 179-185.
- 91) Haskins K, Bradley B, Powers K et al. Oxidative stress in type 1 diabetes. Ann New York Acad Sci 2003; 1005:43-54.
- 92) Kern TS, Kowluru RA, Engerman RL. Abnormalities of retinal metabolism in diabetes or galactosemia: ATPases and glutathione. Invest Ophthalmol Visu Sci 1994; 35(7):2962-2967.

- 93) Kowluru RA, Kern TS, Engerman RL. Abnormalities of retinal metabolism in diabetes or galactosemia II Comparison of γ-glutamyl transpeptidase in retina and cerebral cortex and effects of antioxidant therapy. Curr Eye Res 1994; 13(12): 891-896.
- 94) Kowluru RA, Kern TS, Engerman RL. Abnormalities of retinal metabolism in diabetes or experimental galactosemia IV Antioxidant defense system. Free Rad Bio Med 1996; 22(4):587-592.
- 95) Ford ES, Mokdad AH, Giles WH, Brown DW. The metabolic syndrome and antioxidant concentrations: findings from the Third National Health and Nutrition Examination Survey. Diabetes 2003; 52(9):2346-2352.
- 96) Vlassara H, Fuh H, Makita Z et al 1992; Exogenous advanced glycosylation end products induce complex vascular dysfunction in normal animals: a model for diabetic and aging complications. Proc Nat Acad Sci USA 89:12043.
- 97) Libby P, Rabbani LE, Brogi E, Creager MA. The challenge of diabetic vascular disease In: Year book of Endocrinology. Chicago: Mosby Year Book; 1993.
- 98) Winocour PD, Lopes-Virella M, Colwell JA. Effect of insulin treatment in streptozotocin induced diabetic rats on in vitro platelet function and plasma von Willebrand factor activity and Factor VIII related antigen. J Lab Clin Med 1985; 106:319.
- 99) Ceriello A. Coagulation activation in diabetes mellitus: the role of hyperglycaemia and therapeutic prospects. Diabetologia 1993; 36:1119.
- 100) Ross R, Agius L. The process of atherogenesis cellular and molecular interaction: from experimental animal models to humans. Diabetologia 1992; 35(2):S34-40.
- 101) Kim JA, Berliner JA, Natrajan RD, Nadler JL. Evidence that glucose increases monocyte binding to human aortic endothelial cells. Diabetes 1994; 43:1103.
- 102) Reaven GM, Laws A. Insulin resistance compensatory hyperinsulinaemia and coronary heart disease. Diabetologia 1994; 37:948.
- 103) Brazg RL, Bierman EL. Insulin excess counteracts the effects of HDL on intracellular sterol accumulation in cultured human skin fibroblasts. Diabetologia 1993; 36: 942.
- 104) Punitha R, Manoharan S. Antihyperglycemic and antilipidperoxidative effects of Pongamia pinnata Linn; Pierre flowers in alloxan induced diabetic rats. Ethnopharmacol 2006; 105:39-46.
- 105) Braunwald E, Zipes DP, Libby P. Heart disease: Textbook of cardiovascular medicine. 7<sup>th</sup> ed. Saunders; 2008.
- 106) Davidson MB. Diabetes Mellitus Diagnosis and Treatment. 4th ed. Philadelphia: Saunders; 1998.
- 107) Bowering CK. Diabetic foot ulcers: pathophysiology assessment and therapy. Can Family Physician 2001; 47:1007-1016.
- 108) Clayton W, Elasy TA. A Review of the Pathophysiology Classification and Treatment of Foot Ulcers in Diabetic Patients. Clin Diab Spring 2009; 27(2): 52-58.
- 109) Flynn MD, Tooke JE. Aetiology of Diabetic Foot Ulceration: A Role for the Microcirculation? Issue. Diab Med 1992; 9(4): 320-329.
- 110) Boulton AJ, Armstrong DG, Albert SF, Frykberg RG, Hellman R, Kirkman MS et al. Comprehensive foot examination and risk assessment. Diabetes Care 2008; 31:1679-1685.
- 111) Huijberts MS, Schaper NC, Schalkwijk CG 2008; Advanced glycation end products and diabetic foot disease. Diab Meta Res Rev 24 (1):S19-S24.
- 112) Paraskevas KI, Baker DM, Pompella A, Mikhailidis DP. Does diabetes mellitus play a role in restenosis and patency rates following lower extremity peripheral arterial revascularization? A critical overview. Ann Vasc Sur 2008; 22:481-491.
- 113) Vinik AI, Erbas T, Park TS et al. Platelet dysfunction in type 2 diabetes. Diabetes Care 2001; 24:1476-1485.

- 114) Assert R, Scherk G, Bumbure A et al. Regulation of protein kinase C by short term hyperglycaemia in human platelets in vivo and in vitro. Diabetologia 2001; 44: 188-195.
- 115) Li Y, Woo V, and Bose R. Platelet hyperactivity and abnormal Ca<sup>2+</sup> homeostasis in diabetes mellitus. Am J Physiol-Heart Circul Physiol 2001; 280:H1480-H1489.
- 116) Hafer-Macko CE, Ivey FM, Gyure KA et al. Thrombomodulin deficiency in human diabetic nerve microvasculature. Diabetes 2002; 51:1957-1963.
- 117) Ceriello A, Giacomello R, Stel G et al. Hyperglycemia-induced thrombin formation in diabetes: the possible role of oxidative stress. Diabetes 1995; 44:924–928.
- 118) Ceriello A, Giugliano D, Quatraro A et al. Evidence for a hyperglycaemia-dependent decrease of antithrombin III-thrombin complex formation in humans. Diabetologia 1990; 33:163-167
- 119) Ren S, Lee H, Hu L et al. Impact of diabetes-associated lipoproteins on generation of fibrinolytic regulators from vascular endothelial cells. J Clin Endocrinol Metab 2002; 87:286-291.
- 120) Kario K, Matsuo T, Kobayashi H et al. Activation of tissue factor induced coagulation and endothelial cell dysfunction in non-insulin dependent diabetic patients with microalbuminuria. Arterioscl Thromb Vasc Bio 1995; 15:1114-1120.
- 121) Pandolfi A, Cetrullo D, Polishuck R et al. Plasminogen activator inhibitor type 1 is increased in the arterial wall of type II diabetic subjects. Arterioscl Thromb Vasc Bio 2001; 21:1378-1382.
- 122) McDaid EA, Monaghan B, Parker AI et al. Peripheral autonomic impairment in patients newly diagnosed with type II diabetes. Diabetes Care 1994; 17:1422-1427.
- 123) Hattori Y, Hattori S, Sato N et al. High-glucose-induced nuclear factor kappaB activation in vascular smooth muscle cells. Cardiovasc Res 2000; 46:188-197.
- 124) Suzuki LA, Poot M, Gerrity RG et al. Diabetes accelerates smooth muscle accumulation in lesions of atherosclerosis: lack of direct growth-promoting effects of high glucose levels. Diabetes 2001; 50:851-860.
- 125) Fukumoto H, Naito Z, Asano G et al. Immunohistochemical and morphometric evaluations of coronary atherosclerotic plaques associated with myocardial infarction and diabetes mellitus. J Atheroscl Throm 1998; 5:29-35.
- 126) Armstrong DG, Lavery LA. Diabetic foot ulcers: prevention diagnosis and classification. Am Family Physician 1998; 57(6):1325-1332, 1337-1338.
- 127) Kitabachi, A.E., Wall, B.M., 1995. Diabetic ketoacidosis. Med. Clin. N. Am.79,9-37.
- 128) Smith, C. P., 2006. Diabetic ketoacidosis. Curr. Paed. 16, 111-116.
- 129) Shapero, C., Exley, S. H., Fox, I. M., Rajput, V. J., 2000. Review and case report: Diabetic ketoacidosis. The Foot 10, 105-108.
- 130) Sies H. Antioxidants in Disease, Mechanisms and Therapy. New York: Academic Press; 1996.
- 131) Cadenas E, and Packer L. Hand Book of Antioxidants. New York: Plenum Publishers; 1996.
- 132) Devasagayam TPA, Tilak JC, Boloor KK, Sane KS, Ghaskadbi SS, Lele RD. Free Radicals and Antioxidants in Human Health: Current Status and Future Prospects. J Asso phys Ind 2004; 52:794-804.
- 133) Halliwell B, Aruoma OI. DNA and Free Radicals. Boca Raton Press; 1993.