

Comparative Evaluation of Phenol And Flavonoid Content of Polyherbal Drugs

Divya Kumari Kajaria^{1*}, Mayank Gangwar², Amit Kumar Sharma³, Gopal Nath², Yamini Bhusan Tripathi³, J.S.Tripathi¹, S.K.Tiwari¹

1- Divya Kumari Kajaria, BAMS ,MD, Senior Resident(Ph.D persuing)

Email <divyakajaria@gmail.com> 08808652724

¹Faculty of Aurveda, Department of Kayachikitsa, Institute of Medical Sciences, Banaras Hindu University, Varanasi- 221005 (Uttar pradesh), India.

2- Mayank Gangwar, M.Pharma(pharmacology), Research scholar

Email <mayankgangwar2008@gmail.com> 09458429841, 09368190338

²Department of Microbiology, Institute of Medical Sciences, Banaras Hindu University, Varanasi- 221005 (Uttar pradesh), India.

3- Amit Kumar Sharma, Research scholar

Email <sharmaamitkumar5@gmail.com > 09451977006

³Department of Medicinal Chemistry, Institute of Medical Sciences, Banaras Hindu University, Varanasi- 221005 (Uttar pradesh), India.

4- Dr. Gopal Nath, MBBS,MD, Ph. D, Professor

Email <gopalnath@gmail.com> Fax: 05422367568, 05422309506, 05422300420, 9335058394

²Department of Microbiology, Institute of Medical Sciences, Banaras Hindu University, Varanasi- 221005 (Uttar pradesh), India.

5- Prof. Yamini Bhusan Tripathi , Professor

Email <yaminiok@yahoo.com>, <yamini [at] epatra.com>; <yamini30@sify.com> 09415694450, Phone: 0542-2366577; 307547; fax – 2317074.

³Department of Medicinal Chemistry, Institute of Medical Sciences, Banaras Hindu University, Varanasi- 221005 (Uttar pradesh), India.

6- Dr. Jyoti Shankar Tripathi , BAMS, MD, PhD, Associate Professor

Email <drjstripathi@rediffmail.com > 09838706393

¹Faculty of Aurveda, Department of Kayachikitsa, Institute of Medical Sciences, Banaras Hindu University, Varanasi- 221005 (Uttar pradesh), India.

7- Prof. Shrikant Tiwari , B.A.M.M.S, D.Ay.M, Ph.D., Professor

Email <sktkayachikitsa@yahoo.com > 9415372609, Phone: 91-542-2368926

¹Faculty of Aurveda, Department of Kayachikitsa, Institute of Medical Sciences, Banaras Hindu University, Varanasi- 221005 (Uttar pradesh), India.

Corresponding author:

Divya Kumari Kajaria * M.D. Ph.D persuing

08808652724, Phone: 91-542-2368926, Fax: 05422367568,

Institute of Medical Sciences, Faculty of Aurveda,

Department of Kayachikitsa,

Banaras Hindu University, Varanasi, 221005 (Uttar pradesh), India.

Summary

Background: In Ayurvedic system of medicine mainly polyherbal compounds are used for treatment of various infections. As several reports are on various individual phytochemical, phenolic and flavonoid content but none is having comparative study of polyherbal drugs. **Objective:** Comparative qualitative phytochemical, phenolic, and flavonoid content, TLC analysis was studied of two polyherbal drug named Bharangyadi and Shirishadi. **Methods:** Hydroalcoholic Extraction (Distilled water: Ethanol = 2:1) was done by hot percolation method through soxhlet apparatus. Qualitative phytochemical study was performed using various standard methods. Phenolic content was estimated using Follin ciocalteu reagent, flavonoid using aluminium chloride (2%) reagent as as quercetin equivalent and TLC analysis using hexane, chloroform and methanol solvent. **Result:** The phytochemical analysis showed the presence of alkaloid, saponin, tanins, terpenoids, steroids, glycosides, phenols, flavonoids. Shirishadi contain higher phenolic (112 ± 4.62 mg/g) and flavonoid contents (23.89 ± 3.24 mg/g) compare to bharangyadi polyherbal drug. Ayurveda is traditional system of Indian medicine using mainly herbal products for curing diseases. TLC analysis shows six spots using chloroform and methanol solvent in both the polyherbal drug, while three and four spots using other solvent.

Keywords: Bharangyadi Polyherbal, Shirishadi Polyherbal, Phenol, Flavonoid, TLC, Folin-Ciocalteu, Quercetin.

Introduction

Plants have been associated with the human health from time immemorial and they are the important source of medicines since the down of human civilization. In spite of tremendous developments in the field of allopathic medicines during the 20th century, plants still remain one of the major sources of drugs in modern as well as in traditional systems of medicine. The medicinal plants are rich source of secondary metabolites like alkaloids, glycosides, steroids, and flavonoids, which are potential source of drugs. Approximately, one third of all pharmaceuticals are plant origin. In Ayurvedic system of medicine mainly polyherbal compounds are used for treatment of various infections. *Clerodendrum serratum*, *Hedychium spicatum*, *Inula racemosa*, *Albizzia lebbeck*, *Solanum xanthocarpum* and *Cyprus rotundus* are extensively used in Ayurvedic system of medicine in treating various ailments. Bharangyadi polyherbal is a mixture of *Clerodendrum serratum*, *Hedychium spicatum* and *Inula racemosa*. Shirishadi polyherbal constitute *Shirisha*, *Nagarmotha*, & *Kantakari* is one such preparation.

As all these Plants are able to synthesize a multitude of organic molecules/ phytochemicals, referred to as “secondary metabolites”[1,2] . These molecules play variety of role in the life span of plants, ranging from structural ones to protection. Phenolic compounds are regarded as one such group that is synthesized by plants during development[1,3] and in response to conditions such as infection, wounding ,UV radiation[4,5] etc. Approximately 8000 naturally occurring compounds belong to the category of “phenolics”. Phenols are associated with diverse functions, including nutrient uptake, protein synthesis, enzyme activity, photosynthesis; structural components and allelopathy[6-8] . Phenolics show an array of health promoting benefits in

human health. They are of current interest due to their important biological and pharmacological properties, especially the anti-inflammatory[9] , antioxidant[10] , antimutagenic and anticarcinogenic activities[11, 12] .They are widespread in plant based foods and human consume it. The estimated range of consumption is 25mg to 1g per day, depending on diet[13].

Flavonoids are universal within the plant kingdom. More than 5000 flavonoids have been identified in nature[14]. They functions as stress protectants in plant calls by scavenging reactive oxygen species produced by the photosynthetic electron transport system[15]. Due to UV-absorbing properties, flavonoids protect plants from the UV radiation of the sun and scavenge UV-generated reactive oxygen species[16]. Flavonoids are considered as important components in the human diet, although they are generally considered as non-nutrients. Flavonoid intake can range between 50 and 800 mg per day, depending on the diet consumption. *Albizia lebbek* also known as tree of happiness is extensively used herb in various traditional medicines. Research studies had shown that it possesses anti-histaminic & mast cell stabilizing property[17] by virtue of which it is supposed to work as anti-asthmatic drug^[18]. It also has anti-inflammatory[19], antioxidant properties[20], antiallergic activity[21] and analgesic activity[22]. *Solanum xanthocarpum* known as kantakari in Ayurveda is very effective in respiratory tract disorders. It is found to have strong bronchodilator effect along with anti-inflammatory property [23, 24], hepatoprotective activity[25], antidiabetic[26], antioxidant potential[26, 27], antihyperglycemic[27], larvicidal action[28]. *Cyprus rotundus* or Mustaka reported to possess anti-inflammatory, anti-pyretic and analgesic activities[29].

Bharangi (*Clerodendrum serratum*) is found to have anti-inflammatory[30,31], antihistaminic, antiallergic, antioxidant and hepatoprotective properties[32]. In Ayurvedic system of medicine, it is mainly used in respiratory tract diseases. Sati (*Hedicium spicatum*) is found to possess hypotensive, hypoglycaemic, anti-inflammatory, vasodilator, antispasmodic, tranquillizer, anti-bacterial, anti-fungal, CNS-depressant, hypothermic, spasmolytic & analgesic effects[33,34]. Pushkarmoola (*Inula racemosa*) has been found prove beneficial for cardiovascular system, angina and dyspnoea[35,36].

Therefore this study was planned to evaluate comparative phenolic, flavonoid content and TLC analysis of hydroalcoholic extract of Shirishadi, Bharanyadi polyherbal drug with preliminary phytochemical characterization.

Material And Methods

Plant material and preparation of Extract

The plants of polyherbal drugs were collected from local market of Varanasi (India). The identification of the plants was done by Prof. A. K. Singh, Department of Dravyaguna, S.S.U., Varanasi. Air shade dried and pulverized plants parts were extracted with Hydroalcoholic solvent (Distilled water: Ethanol = 2:1) separately by hot percolation method through soxhlet apparatus. Thereafter extract was dried using rotary evaporator and dried extract was put to the process of standardization. This extract was used to investigate the total content of phenols and flavonoids.

Preliminary Phytochemical Screening

Chemical test were carried out on the aqueous and hydroalcoholic extract and on the powdered specimen using standard procedures to identify the constituents [37, 38]. The plant extract was assayed for the presence of alkaloids, proteins, free amino acids, anthraquinone, glycosides, flavonoids, tannins, phenolic compounds, carbohydrates, saponins, phytosterol and triterpenes.

Estimation of total phenolic content

Total Phenolic concentration in different fractions of alcoholic extract was measured by Folin ciocalteu assay^[39, 40]. Briefly, 5ml of distilled water, 0.5-1.0ml of sample, and 1.0ml of Folin ciocalteu reagent was added to a 25ml flask. The content was mixed and allowed to stand for 5-8min at room temperature. Next 10ml of 7% sodium carbonate solution was added followed by distilled water. Solution were mixed and allowed to stand at room temperature for 15min, and then absorbance was recorded at 750 nm. TP content was standardized against gallic acid and expressed as milligram per liter of gallic acid equivalents (GAE). The linearity range for this assay was determined as 0.5-5.0mg/l GAE ($R^2=0.999$), giving an absorbance range of 0.050-0.555 absorbance units. Experiments were performed in triplicates and results were recorded as mean \pm SEM (Standard Error Mean).

Estimation of total flavonoid content

Total flavonoid content was measured by using aluminium chloride (2%)^[41, 42] in which it is mixed with solution of test samples. Absorbance reading at 415nm (Elico SL 177) were taken after 10 min against a blank sample consisting of 5ml of sample solution and 5ml of methanol without aluminium chloride. The total Flavonoid content was determined using a standard curve of quercetin at 0-50 μ g/ml^[43]. The average of three readings were used and then expressed in μ g quercetin equivalent flavones per mg extract. Experiments were performed in triplicates and results were recorded as mean \pm SEM (Standard Error Mean).

Thin layer chromatography-(TLC): Thin layer chromatography (TLC) was used to separate the shrishadi and Bharangyadi extract into different spots on the chromatplate. The chromatograms developed on the microscope slide, were dried and observed visually for the various different part of polyherbal extract components. The developing solvent used in different extract are hexane: ethyl acetate(9:1) and chloroform:methanol(9.5:5).

The retention factor was calculated using:

$$R_f = \frac{\text{Distance move by the substance(cm)}}{\text{Distance move by the solvent(cm)}}$$

Results And Discussion

Bharangyadi polyherbal extractive value was found to be higher than shrishadi polyherbal drug. Plant extracts with high phenolic content also enclosed high flavonoid content. Estimation of total phenolic and total flavonoid content showed that shrishadi extract was having maximum phenolic content (112 \pm 4.62 mg/g) in μ g Gallic acid equivalents (GAE) followed by Bharangyadi

extract (50.98 ± 1.81 mg/g) with a very less flavonoid content in Bharangyadi extract (13.66 ± 0.54 mg/g) and Shrishadi (23.89 ± 4.62 mg/g) extracts in μg of quercetin equivalents (QE). The amount of total phenolic and flavonoids for the test samples are summarized (Table 1 and 2). The preliminary phytochemical screening of hydroalcoholic extract of polyherbal drugs are presented in Table 3, showing the presence of alkaloids, phenolic groups, flavonoids, saponins, steroids, reducing sugars, tannins and anthraquinones, cardiac glycosides, phlobatanins along with carbohydrate, amino acid & protein. The result of TLC analysis using ethyl acetate: chloroform: methanol solvent mixture as shown in table 4 which revealed three to six spots in different solvent.

Flavonoids has been considered to effect on human nutrition and health as it show antioxidant activity, and their mechanism of action are through scavenging or chelating process[44, 45]. It has been recognized that phenolic compounds are a class of antioxidant compounds which act as free radical terminators[46]. These compounds such as flavonoids, which contain hydroxyl functional groups, are responsible for antioxidant effect in the plants [47, 48]. The pharmacological activities of the drug contributed by the presence of secondary metabolites. In these classes (such as alkaloids, saponins, tannins, anthraquinones and flavonoids) of compounds are known to have activity against several pathogens and therefore could suggest their traditional use for the treatment of various illness [49].

Table 1. Extraction Yield, Total Phenol and Flavonoid (mg/g) contents of Bharangadi Polyherbal drug:

Extractive value	12%
Total Phenolic Content	50.98 ± 1.81 mg/g
Total Flavonoid Content	13.66 ± 0.54 mg/g

Table 2. Extraction Yield, Total Phenol and Flavonoid (mg/g) contents of shrishadi Polyherbal drug:

Extractive value	12%
Total Phenolic Content	112 ± 4.62 mg/g
Total Flavonoid Content	23.89 ± 4.62 mg/g

Table 3. Preliminary phytochemical screening of polyherbal drugs

Constituents	<i>Clerodendrum serratum</i>	<i>Hedychium spicatum</i>	<i>Inula racemosa</i>	<i>Albezzia lebback</i>	<i>Cyprus rotandus</i>	<i>Solanum xanthocarpum</i>
Alkaloids	+	-	+	+	+	+
Amino acids	+	+	-	+	+	+
Cardiac Glycosides	+	-	-	+	+	+
Flavones	+	-	+	+	+	+
Quinones	+	+	+	+	+	+
Saponins	+	+	-	-	-	+
Steroids	+	-	+	-	+	+
Sugars	+	-	+	-	-	+
Tannins	+	-	+	+	+	+
Triterpenes	+	+	-	+	+	+
Carbohydrates	+	+	+	+	+	+
Protein	+	-	-	+	-	+

Table 4: TLC Result of hydroalcoholic extract of polyherbal drug

Extracts	Solvent system	Number of components	Distance of spot (cm)	Solvent front (cm)	Rf value
shrishadi	Hexane:ethyl acetate (9:1)	3	11.5, 9.4, 7.3	13	0.88, 0.77, 0.56
	chloroform:methanol (9.5:5)	6	11.2, 9.2, 5.7, 3.9, 2.9, 2.4	13	0.86, 0.70, 0.43, 0.30, 0.22, 0.18
Bharangyadi	Hexane:ethyl acetate (9:1)	4	10, 5.6, 4.9, 3.3	13	0.76, 0.43, 0.37, 0.25
	chloroform:methanol (9.5:5)	6	11.5, 8.5, 6.3, 5.2, 4.3, 3.4	13	0.88, 0.65, 0.48, 0.40, 0.33, 0.26

Conclusion

This study indicates that both polyherbal hydroalcoholic extracts showed presence of high amount of phenolic and flavonoid compounds along with secondary metabolites from various medicinally important plants suggesting the use of polyherbal compounds for treatment of various infections.

Acknowledgements

Authors are thankful to the Department of Kayachikitsa, Faculty of Aurveda, Institute of Medical Sciences, Banaras Hindu University, Varanasi, India for providing the necessary laboratory facilities for the work. Authors are also thankful to University Grant Commission (UGC- Delhi) for the financial support.

References

1. Harborne JB. Introduction to Ecological Biochemistry, 2nd Ed., Academic Press, New York, NY. 1982
2. Harborne JB. Turner B. L., Plant Chemosystematics, Academic Press, London, UK, 1984.
3. Pridham JB. Phenolics in Plants in Health and Disease, Pergamon Press, New York, NY, 1960
4. Shahidi F, Naczki M. Phenolics in Food and Nutraceuticals: Sources, Applications and Health Effects, CRC Press, Boca Raton, FL., 2004
5. Beckman CH. *Physiol.Mol.Plant Pathol.*, 2000, 57,101-110
6. Wu H, Haig T, Pratley J, Lemerle D, *Journal of Chromatogr.A* 1999.864, 315-321
7. Wu H, Haig T, Pratley J, Lemerle D, An M., *Journal of Agri. Food Chem.*2000,48, 5321-5325
8. Einhellig FA. Putnam AR, Tang CS. *The Science of Allelopathy*, John Wiley and sons. New York, 1996, 171-189
9. Koshihara T, Neichi T, Murota S, Lao A, Fujimoto Y, Tatsuno T, *Biochim. Biophys. Acta*, 1984, 792, 92-97
10. Zhou J, Ashoori F, Susuki S, Nishigaki I, Yagi K, *J. Clin. Biochem. Nutr.*1993, 15,119-125
11. Tanaka T, Kojima T, Kawamori T, Yoshimi N, Mori H, *Cancer Res.* 1993,53,2775-2779.
12. Serrano A, Palacios C, Roy G, Cespon C, Villar MI, Nocito M, Porque PG, *Arch. Biochem. Biophys.* 1998, 350, 49-54
13. Clifford MN, *Journal of Sci. Food Agri.* 1999, 79,362-372
14. Prior RL, Wu H, Gu L. Flavonoid metabolism and challenges to understanding mechanisms of health effects. *J. Sci. Food Agric.* **2006**, 86, 2487-2491.
15. Harborne JB. *Flavonoids: Advances in Research since 1986*, Chapman and Hall, London 1994, 589-618
16. Shirley BW. *Trends Plant Sci.*1996, 31,377-382
17. Shashidhara S, Bhandarkar AV, Deepak M. Comparative evaluation of successive extracts of leaf and stem bark of *Albizia lebeck* for mast cell stabilization activity. *Fitoterapia* 2008 ;**79**(4):301-2.

18. Pratibha N, Saxena VS, Amit A, D'Souza P, Bagchi M, Bagchi D. Antiinflammatory activities of Aller-7, a novel polyherbal formulation for allergic rhinitis. *Int. J. Tissue. React* 2004; **26**: 43-51.
19. Babu NP, Pandikumar P, Ignacimuthu S. Anti-inflammatory activity of *Albizia lebbeck* Benth., an ethnomedicinal plant, in acute and chronic animal models of inflammation. *J Ethnopharmacol* 2009; **125**(2):356-60.
20. Resmi CR, Venukumar MR, Latha MS. Antioxidant activity of *Albizia lebbeck* (Linn.) Benth. in alloxan diabetic rats. *Indian J Physiol Pharmacol* 2006; **50**(3):297-302.
21. Venkatesh P, Mukherjee PK, Kumar NS, Bandyopadhyay A, Fukui H, Mizuguchi, Islam N. Anti-allergic activity of standardized extract of *Albizia lebbeck* with reference to catechin as a phytomarker. *Immunopharmacol Immunotoxicol* 2010; **32**(2):272-6.
22. Saha A, Ahmed M. The analgesic and anti-inflammatory activities of the extract of *Albizia lebbeck* in animal model. *Pak J Pharm Sci* 2009; **22**(1):74-7.
23. Anwikar S, Bhitre M. Study of the synergistic anti-inflammatory activity of *Solanum xanthocarpum* Schrad and Wendl and *Cassia fistula* Linn. *Int J Ayurveda Res* 2010; **1**(3):167-71.
24. Bhitre J, Bhakti P, Milind. Study of the synergistic antiinflammatory activity of *solanum xanthocarpum* schrad and wendl and *piper nigrum* linn; *International Journal of Ayurvedic and Herbal Medicine* 2011; **1**: 42 –53.
25. Gupta RK, Hussain T, Panigrahi G, Das A, Singh GN, Sweety K, Faiyazuddin M, Rao CV. Hepatoprotective effect of *Solanum xanthocarpum* fruit extract against CCl₄ induced acute liver toxicity in experimental animals. *Asian Pac J Trop Med* 2011; **4**(12):964-8.
26. Gupta R, Sharma AK, Sharma MC, Dobhal MP, Gupta RS. Evaluation of antidiabetic and antioxidant potential of lupeol in experimental hyperglycaemia. *Nat Prod Res* 2011. DOI:10.1080/14786419.2011.560845.
27. Pongothai K, Ponmurugan P, Ahmed KS, Kumar BS, Sheriff SA. Antihyperglycemic and antioxidant effects of *Solanum xanthocarpum* leaves (field grown & in vitro raised) extracts on alloxan induced diabetic rats. *Asian Pac J Trop Med* 2011; **4** (10):778-85.
28. Mohan L, Sharma P, Srivastava CN. Combination larvicidal action of *Solanum xanthocarpum* extract and certain synthetic insecticides against filarial vector, *Culex quinquefasciatus* (SAY). *Southeast Asian J Trop Med Public Health* 2010; **41**(2):311-9.
29. Nagulendran K, Velavan S, Mahesh R. In Vitro Antioxidant Activity and Total Polyphenolic Content of *Cyperus rotundus* Rhizomes, *E-Journal of Chemistry* 2007; **4**: 440-449.
30. Das S, Haldar PK, Pramanik G, Suresh RB, Evaluation of Anti-Inflammatory Activity of *Clerodendron infortunatum* Linn. Extract in Rats; *Global Journal of Pharmacology*, 4, 2010, 48-50.
31. Narayanan N, Thirugnanasambantham P, Viswanathan S, Vijayasekaran V, Sukumar E. Evaluation of antinociceptive, antiinflammatory and antipyretic activities of ethanolic extract of roots of *Clerodendron serratum* on experimental animal models, *J Ethnopharmacol*, 65,1999, 237-241.
32. Vidya SM, Krishna V, Manjunatha BK, Mankani, MAnzoor Ahmed KL, Singh J, S.D. Evaluation of hepatoprotective activity of *Clerodendrum serratum* L, *Indian Journal of Experimental Biology*, 45, 2007, 538-542.

33. Gupta SS, Rai M, Gupta NK, Histamine releasing effects of a few Indian medicinal plants used in bronchial asthma, *Curr. Sci*, 36,1967, 42.
34. Modh PR, Gupta SS, Effect of a plant saponin on Histamine release in relation to their anticholinesterase activity, *Indian J.Physioal, Pharmacol*, 13, 1969, 57.
35. Srimal RC, Sharma SC, Tandon JS, Antinflammatory and other pharmacological effects of *Hedychium spicatum* (Buch-Hem), *Indian Journal of Pharmacology*, 16, 1984, 143-147.
36. Bhatt ID, Prasad K, Rawat S, Rawal RS, Evaluation of antioxidant phytochemical diversity in *Hedychium spicatum*: A high value medicinal plant of Himalaya. *PHOG MAG*, 4, 2008, 202-205.
37. Ganga Rao Battu, Sambasivarao Ethadi, Prayaga Murthy.P, V.S.Praneeth.D, Mallikarjuna Rao.T. In-Vitro Antibacterial Activity And Preliminary Phytochemical Screening Of Three Algae From Visakhapatnam Coast, Andhra Pradesh, India. *Int J Pharm Pharm Sci* 2011; **3(4)** : 399-401.
38. Harborne JB, *Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis*, (3rd edition). Chapman and Hall Co., New York, 1998, 1-302.
39. Malik EP, Singh MB. *Plant Enzymology and Hittoenzymology*, (First Edn.) Kalyani Publishers, New Delhi, 1980, 286.
40. Rasika CT, Lavate SM , Jadhav RB, Kamble GS and Deshpande NR. Evaluation Of Phenol And Flavonoid Content From Aerial Parts Of *Tecoma Stans*. *Int J Pharm Pharm Sci* 2011; **3(4)** 126-127.
41. Chang C. Yang M, Wen H, Chern J Estimation of total flavonoid content in propolis by two complementary calorimetric methods. *J. Food Drug Analysis* 2002, 10:178-182.
42. Meda A, Lamien CE, Romito M, Millogo J, Nacoulma OG, Determination of the total phenolic, flavonoid and proline contents in Burkina Fasan honey, as well as their radical scavenging activity. *Food Chem* 2005; **91**: 571-577.
43. Zhishen J, Mengcheng T, Jianming W. The determination of flavanoid contents in mulberry and their scavenging effects on superoxid radicals. *Food Chem.* **1999**, *64*, 555–559.
44. Kessler M, Ubeaud G, Jung L. Anti- and pro-oxidant activity of rutin and quercetin derivatives. *J. Pharm. Pharmacol.* 2003;**55**: 131-142.
45. Cook NC, Samman S (1996). Flavonoids- chemistry, metabolism, cardioprotective effects, and dietary sources. *Nutr. Biochem.* 7: 66- 76.
46. Shahidi F, Wanasundara PK. Phenolic antioxidants. *Crit Rev. Food Sci. Nutr.* 1992. **32**: 67-103.
47. Das NP, Pereira TA Effects of flavonoids on thermal autooxidation of Palm oil: structure-activity relationship. *J. Am. Oil Chem. Soc.* 1990; **67**: 255- 258.
48. Younes M. Inhibitory action of some flavonoids on enhanced spontaneous lipid peroxidation following glutathione depletion. *Plant Med* ,1981;**43**: 240-245.
49. Hassan MM, Oyewale AO, Amupitan JO, Abdullahi MS, Okonkwo EM, Preliminary Phytochemical and antibacterial investigation of crude extracts of the root bark of *Detarium microcarpum*, *J.Chem. Soc. Nigeria* 2004; **29**: 26-29.