

REVISITING ANTHROPOZOONOSES IN THE MEDITERRANEAN BASIN. A SINGLE-CENTRE PERSPECTIVE. A SOUTHERN ITALIAN EXPERIENCE

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Abstract

Zoonoses, often better defined with the term anthroponoses, are diseases that can be transmitted to humans either by direct contact with animals or through arthropod vector intervention. Microbial interaction between humans and animals constitutes an important public health challenge, particularly in the Mediterranean basin. The main reasons this challenge is still relevant today include the phenomenon of migration, of both humans and animals, and climate change, which tend to alter the geographical distribution of zoonosis or the zoonotic agent, as well as the distribution range of potential vectors. The Mediterranean area seems to be affected by plentiful and widely distributed zoonoses, the main diseases being rickettsiosis, leishmaniasis, brucellosis, hydatid disease and viral zoonoses. The aim of this study is to revisit the prevalence and main clinical features of anthroponoses observed at the Department of Sciences for Health Promotion and Mother & Child Care, University of Palermo, Sicily, Southern Italy.

Keywords: Anthroponoses, Mediterranean basin, Sicily

Introduction

The Mediterranean area seems to be affected by plentiful and widely distributed zoonoses. The reasons for this concentration are to be referred to the great biodiversity and the close co-existence between people and animals [1-3]. Zoonoses, an all encompassing term, better often defined with the term “anthropozoonoses” (diseases transmitted to man from other vertebrates), defines a large group of infectious diseases that still represent a public health challenge due to the complex microbial interaction between humans and animals [4-13]. Many zoonotic diseases have disappeared or been eradicated: historical examples are the plague (absent for more than three centuries), canine rabies (or urban, last reported case in 1974), sylvatic rabies and American myiasis (*Cochliomyia hominivorax*, the killer fly, introduced in the 1990s and promptly eradicated thanks to the release into the environment of sterile males). Other zoonotic diseases are still present in the Mediterranean basin: rickettsiosis, leishmaniasis and brucellosis are well-known diseases, while others are emerging and rare [1-3,14-19].

Furthermore, the multidimensional phenomenon of migration, climate change and the increase in national and international transport are changing the geographical distribution of the zoonosis or the zoonotic agent, as well as the distribution range of potential vectors [20-24].

Rickettsiosis

Rickettsiosis is a globally widespread zoonosis caused by gram-negative bacteria belonging to the genus *Rickettsia* and *Orientia*, which are transmitted by arthropod vectors (lice, fleas, ticks and mites). In Europe only the species belonging to the genus *Rickettsia* are responsible for Rickettsiosis, and typically fall into two general groups: the spotted fever group and the typhus group.

The endemic typhus (murine typhus) caused by *Rickettsia typhi* and transmitted by the rat flea (*X. cheopis*) is common in Mediterranean countries (Spain, Croatia, Greece, Cyprus). *Rickettsia felis*

is found in the cat flea (*C. felis*) with high vector prevalence in Europe [25].

However in Europe the predominant rickettsioses belong to the spotted fever group.

Mediterranean spotted fever (MSF), one of the oldest-recognized vector-borne infectious diseases, is an endemic zoonosis in the Mediterranean basin. MSF is caused by *Rickettsia conorii* subsp. *conorii* and it is endemic in southern Europe with sporadic documented cases in some regions of northern and central Europe; *Rhipicephalus sanguineus* is the principal vector and reservoir. The clinical presentation includes fever, maculopapular rash, and the presence of a tache noir on the tick bite site. Most of the time the clinical course is positive, with a mortality rate of 3-7% in hospitalized patients. Most cases are reported in the summer months, when ticks are very active. In the southern European region the higher temperatures permit the persistence and abundance of ticks and for this reason cases can be reported during the spring. In Italy, each year about 300 MSF cases are reported, of which more than half are in Sicily [26,27]. *Rickettsia conorii* subsp. *conorii* has long been considered the only pathogen rickettsia in Europe. However, in recent decades the use of molecular biology techniques allowed the identification of new rickettsia species and subspecies as new emerging pathogens responsible for new clinical diseases different from the classical form of FBM: *Rickettsia conorii* subsp. *israelensis*, *Rickettsia conorii* subsp. *caspia*, *Rickettsia conorii* subsp. *indica*, *Rickettsia slovacica*, *Rickettsia raoultii*, *Rickettsia monacensis*, *Rickettsia massiliae*, *Rickettsia aeschlimannii*, *Rickettsia helvetica*, *Rickettsia sibirica* subsp. *sibirica* and *Rickettsia sibirica* subsp. *mongolitimonae*. In 1996 and 1997 in France the first case of human infection with *Rickettsia slovacica* and *Rickettsia sibirica* *mongolitimonae* was described; *Rickettsia massiliae* was identified for the first time in Marseille and was recognized as a disease agent in Sicily in 2006 [28]. In 2003 in Sicily, *Rickettsia conorii* subsp. *israelensis* was identified, agent of the Israeli spotted fever (ISF), documented in Italy (Sicily and Sardinia) and in other Mediterranean

areas (e.g. Portugal). The distribution of ISF has proved to be wider than previously thought, and it is now possible to assert that many of the MSF severe clinical pictures described in the literature should be attributed to *R. rickettsii* subsp. *Israelensis* [29]. Lastly, TIBOLA / DEBONEL / SENLAT, acronyms for respectively, 'Tick-Borne Lymphadenopathy' / 'Dermacentor-Borne Necrosis Erythema Lymphadenopathy' / 'Scalp ESCHAR and Neck Lymphadenopathy After Tick Bite', are caused by *Rickettsia slovaca*, *Candidatus Rickettsia* and *Rickettsia raoultii*. The main vector in these cases is *Dermacentor marginatus* and the major symptoms are necrotic eschar on the scalp associated with painful cervical lymphadenopathy [28].

Although MSF is mostly a self-limited disease characterized by fever, skin rash, and a dark eschar at the site of the tick bite called a 'tache noire', serious complications have been described, mainly in adult patients [30-35].

Tetracyclines are considered standard treatment for MSF, even though they can cause significant adverse effects like staining of the teeth and bone toxicity, especially in children. For this reason, macrolides have emerged as a potential alternative therapy in children [36-38].

Leishmaniasis

Leishmaniasis are a group of vector-borne parasitic diseases caused by protozoa belonging to the genus *Leishmania*. Generally, *Leishmania* infection is transmitted to humans and to other mammals by the bite of an infected sand fly vector. Rarely, the infection can be transmitted through blood transfusions, by needle sharing, or from mother to child during pregnancy. The World Health Organization (WHO) has stated that leishmaniasis is one of the most neglected diseases, with 350 million people considered at risk of contracting the disease, a burden of about 12 million people currently infected in 98 countries, and two million new cases estimated to occur annually. Among these, visceral leishmaniasis (VL) accounts for about 500 000 cases each year. The clinical spectrum includes cutaneous, mucocutaneous and visceral forms.

Asymptomatic infections have also been demonstrated, but their role has yet to be clarified.

Parasite, vector and host, and their complex interplay, determine the different clinical forms of VL [40]. It has been estimated that in endemic areas the proportion of asymptomatic infections is 5–10 times greater than the number of clinically apparent VL cases in immunocompetent hosts. Cryptic infection can be detected in people without a previous history of clinical VL by serological evidence of anti-*Leishmania* antibodies, by detection of parasite DNA in blood samples, or by a positive reaction to the leishmanin skin test (LST) [41,42]. Asymptomatic infection has also been demonstrated in HIV-infected patients. Also in this population, the percentage of asymptomatic infection could be higher than symptomatic cases. *Leishmania* parasitemia was found to be significantly higher in patients with higher viral loads. A high parasitemic burden could possibly be related to a higher risk of developing symptomatic disease, due to the reciprocal effects that enhance the multiplication of both pathogens [42-44].

In the immunocompetent host, VL is caused by a primary infection with *Leishmania* parasites transmitted by the bite of a phlebotomine sand fly. VL is the result of a chronic infection; the incubation period ranges from 10 days to 1 year. Clinical features of typical forms are fever, weight loss, hepatosplenomegaly, and pancytopenia. Fever can be intermittent in the first period and successively continuous. Non-tender splenomegaly and hepatomegaly are caused by infection of the reticuloendothelial system. Pancytopenia caused by parasites invading the bone marrow is responsible for pallor due to anaemia, and can subsequently cause haemorrhages due to thrombocytopenia and concurrent infections due to leukopenia. Anorexia and weight loss can lead to wasting syndrome in misdiagnosed cases [44-47]. PCR on peripheral blood allows a rapid and non-invasive parasitologic diagnosis of VL [48,49].

Leishmaniasis has been reported among transplant recipients, HIV patients and among other immunosuppressed patients [41,50,51].

Anti-leishmanial treatment is based on the systemic administration of one or a combination of effective drugs. Alongside pentavalent antimonials, which have been the standard first-line medicines for many decades, new anti-leishmanial drugs such as lipid formulations of amphotericin B, miltefosine, and paromomycin are currently used for the treatment of VL. Lipid formulations of amphotericin B, and in particular liposomal amphotericin B, are considered to be the drugs of choice for the treatment of VL [52,53].

Case reports and small series have anecdotally reported the successful use of liposomal amphotericin B or pentavalent antimony in VL complicated by secondary hemophagocytic lymphohistiocytosis [49].

Conditions of depression of the immune system, such as HIV infection or immunosuppressive treatments in transplant recipients and in patients with autoimmune diseases impair the capability of the immune response to resolve the infection and allow the reactivation of the disease from sites of latency of the parasite. Reactivation of chronic infection can occur long after primary contact with the parasite [52-55]. Appropriate screening for leishmaniasis before beginning immunosuppressive treatments could be useful for calling to attention the potential risk for VL in the immunocompromised host. The LST (Montenegro test), an intradermal injection of a suspension of killed promastigotes, measures delayed hypersensitivity reactions and appears to be valuable for detecting asymptomatic *Leishmania* infections [56].

Brucellosis

Brucellosis is a zoonotic infection in domestic and wild animals that is caused by organisms of the *Brucella* genus. Humans become infected by ingesting unpasteurized dairy products, being in direct contact with infected animals, or inhaling infectious aerosols. Brucellosis is one of the world's most widespread zoonoses. The distribution of this disease is worldwide, and areas of high endemicity include the Mediterranean, the Middle East, Latin America and Asia [7,57].

Domestic animals harbouring *Brucella* spp. are raised where adequate control measures are lacking and where the population has the custom of ingesting unpasteurized milk or its products. In Sicily, the largest Italian island, brucellosis is highly endemic and has shown a marked resurgence in the last few years (14 cases per 100,000).

However, fewer than 200 cases are reported per year (0.04 cases per 100,000 population) in the United States.

Brucella spp. are intracellular pathogens that can survive and multiply within mononuclear phagocytes (monocytes and macrophages) of the reticuloendothelial system (RES). Localization within organs of RES may explain some of the clinical manifestations of systemic brucellosis, such as hepatosplenomegaly and the propensity for involvement of the skeletal system. Lack of feasible microbiological methodologies [58] or effective treatment may result in serious and sometimes life-threatening complications such as spondylitis, endocarditis and encephalitis [59-62].

Patients frequently experience relapse, even with treatment, and the disease often becomes chronic (i.e., a clinical manifestation of >6 months duration).

Tolomeo et al. have shown that a high level of apoptosis resistance among monocytes and lymphocytes during and after therapy may therefore represent an index of chronic illness, suggesting to the clinician, in some cases, that a change in therapy is required [63].

The treatment of choice for acute brucellosis is considered a 6-week regimen of tetracycline administered orally in combination with streptomycin (1 g/day intramuscularly for 2-3 weeks). Although other antibiotics have been used, no substantial improvement in relapse rates has been reported in association with any new treatment regimen in the past 45 years [4]. In 1986, the World Health Organization (WHO) recommended therapy with the combination of doxycycline (200 mg/day) plus rifampin (600-900 mg/day) both administered once daily by mouth for 6 weeks [64,65].

Cascio et al. [63] showed that the combination of intravenous rifampin plus oral minocycline administered for 3 weeks obtained the lowest relapse rate (1.7%). This combination therapy, like those reported by WHO (i.e. doxycycline plus rifampin) versus streptomycin plus doxycycline, showed a relapse rate of 16% and 5.3%, respectively [66].

Experience with different treatments in childhood brucellosis is sparse and generally tetracyclines are not used in children ≤ 8 years of age [67,68]. Cascio et al. have published a study suggesting that minocycline could be used (for a maximum of 3 weeks) to treat infections in pediatric patients when indicated [69].

Other rare zoonotic diseases

Erysipelothrix rhusiopathiae is a common commensal or pathogen of many vertebrate and invertebrate species. Pigs are a major reservoir. Human disease is mainly an occupationally acquired zoonosis. The portal of entry is typically a puncture wound or abrasion on the hand; however, it can also be acquired from eating contaminated food [70].

The global burden of leptospirosis remains enormous and new aspects of the disease are constantly being recognized [71].

Of note, animal rotaviruses, astrovirus, and picobirnavirus might be able to cross species barriers, and lack of systematic surveillance of rotavirus infection in small animals hinders the ability to establish firm epidemiological connections [14,71-75].

Echinococcosis and other helminthic diseases are endemic in Southern Italy [8,9,18].

Over the last ten years, studies have confirmed that besides mammals, birds too are responsible for arthropod human infestation. Migratory wild birds play an important role in the ecology and circulation of potential zoonotic pathogens in Sicily [76,77].

Castelli et al. showed how the bite of an avian arthropod, such as *Ornithonyssus* species of bird parasite, could be responsible for dermatitis in a Sicilian patient [77].

In conclusion, Southern Italy and Sicily in particular could be affected by emerging diseases caused by the passage of animals carrying arthropods other than well-known ones such as ticks, and this might be the consequence of climate tropicalization, which carries unpredictable epidemiological and ecological implications.

References

1. Zeppelini CG, de Almeida AM, Cordeiro-Estrela P. Zoonoses As Ecological Entities: A Case Review of Plague. *PLoS Negl Trop Dis*. 2016.
2. Cascio A, Bosilkovski M, Rodriguez-Morales AJ, Pappas G. The socio-ecology of zoonotic infections. *Clin Microbiol Infect*. 2011 Mar;17(3):336-42.
3. Pappas G, Cascio A, Rodriguez-Morales AJ. The immunology of zoonotic infections. *Clin Dev Immunol*. 2012;2012:208508.
4. Cascio A, Gradoni L, Scarlata F, Gramiccia M, Giordano S, Russo R, Scalone A, Camma C, Titone L. Epidemiologic surveillance of visceral leishmaniasis in Sicily, Italy. *Am J Trop Med Hyg*. 1997 Jul;57(1):75-8.
5. Blanda V, Torina A, La Russa F, D'Agostino R, Randazzo K, Scimeca S, Giudice E, Caracappa S, Cascio A, de la Fuente J. A retrospective study of the characterization of *Rickettsia* species in ticks collected from humans. *Ticks Tick Borne Dis*. 2017 Jun;8(4):610-614.
6. Otranto D, Dantas-Torres F, Giannelli A, Latrofa MS, Cascio A, Cazzin S, Ravagnan S, Montarsi F, Zanzani SA, Manfredi MT, Capelli G. Ticks infesting humans in Italy and associated pathogens. *Parasite Vectors*. 2014 Jul 14;7:328.
7. Iaria C, Ricciardi F, Marano F, Puglisi G, Pappas G, Cascio A. Live nativity and brucellosis, Sicily. *Emerg Infect Dis*. 2006 Dec;12(12):2001-2.
8. Minciullo PL, Cascio A, David A, Pernice LM, Calapai G, Gangemi S. Anaphylaxis

- caused by helminths: review of the literature. *Eur Rev Med Pharmacol Sci*. 2012 Oct;16(11):1513-8.
9. Minciullo PL, Cascio A, Isola S, Gangemi S. Different clinical allergological features of *Taenia solium* infestation. *Clin Mol Allergy*. 2016 Dec 7;14:18.
 10. Cascio A, Pernice LM, Barberi G, Delfino D, Biondo C, Beninati C, Mancuso G, Rodriguez-Morales AJ, Iaria C. Secondary hemophagocytic lymphohistiocytosis in zoonoses. A systematic review. *Eur Rev Med Pharmacol Sci*. 2012 Oct;16(10):1324-37.
 11. Mansueto P, Vitale G, Cascio A, Seidita A, Pepe I, Carroccio A, di Rosa S, Rini GB, Cillari E, Walker DH. New insight into immunity and immunopathology of Rickettsial diseases. *Clin Dev Immunol*. 2012;2012:967852.
 12. Cascio A, Giordano S, Dones P, Venezia S, Iaria C, Ziino O. Haemophagocytic syndrome and rickettsial diseases. *J Med Microbiol*. 2011 Apr;60(Pt 4):537-42.
 13. Colomba C, Rubino R, Di Carlo P, Mammaia C, Bonura C, Siracusa L, Titone L, Saporito L. Probable disseminated *Mycobacterium abscessus* subspecies *bolletii* infection in a patient with idiopathic CD4+ T lymphocytopenia: a case report. *J Med Case Rep*. 2012 Sep 4;6:277.
 14. Mendell NL, Bouyer DH, Walker DH. Murine models of scrub typhus associated with host control of *Orientia tsutsugamushi* infection. *PLoS Negl Trop Dis*. 2017.
 15. De Grazia S, Martella V, Giammanco GM, Gòmara MI, Ramirez S, Cascio A, Colomba C, Arista S. Canine-origin G3P[3] rotavirus strain in child with acute gastroenteritis. *Emerg Infect Dis* 2007; 13:1091-1093.
 16. Cascio A, Stassi G, Cacciola I, Saitta C, Squadrito G. Fever and rhomboid target lesion in decompensated cirrhosis *Lancet Infect Dis*. 2012 Jul;12(7):576. doi: 10.1016/S1473-3099(12)70063-4.
 17. Pappas G, Cascio A. Optimal treatment of leptospirosis: queries and projections. *Int J Antimicrob Agents*. 2006 Dec;28(6):491-6.
 18. Colomba C, Scarlata F, Di Carlo P, Giammanco A, Fasciana T, Trizzino M, Cascio A. Fourth case of louse-borne relapsing fever in Young Migrant, Sicily, Italy, December 2015. Mini Review Article. *Public Health*. 2016 Oct;139:22-26.
 19. Salamone G, Licari L, Randisi B, Di Carlo P, Tutino R, Falco N, Augello G, Raspanti C, Cocorullo G, Gulotta G. A primary subcutaneous hydatid cyst in the thigh. A case report. *Ann Ital Chir*. 2016 Apr 8;87(ePub).
 20. Casuccio A, D'Angelo C, Casuccio N, Di Carlo P, Immordino P. Visiting Friends and Relatives (VFRs) travelers and imported malaria in the Palermo district (Sicily). *Ann Ist Super Sanita*. 2014;50(4):369-74.
 21. Di Carlo P, Guadagnino G, Immordino P, Mazzola G, Colletti P, Alongi I, Adamoli L, Vitale F, Casuccio A. Behavioral and clinical characteristics of people receiving medical care for HIV infection in an outpatient facility in Sicily, Italy. *Patient Prefer Adherence*. 2016, 25;10:919-27.
 22. Stefanelli P, Fazio C, Neri A, Rezza G, Severoni S, Vacca P, Fasciana T, Bisbano A, Di Bernardo F, Giammanco A. Imported and Indigenous cases of Invasive Meningococcal Disease W:P1.5,2:F1-1:ST-11 in migrants' reception centers. Italy, June-November 2014. *Adv Exp Med Biol*. 2016;897:81-3.
 23. Usuelli FG, Maccario C, Ursino C, Serra N, D'Ambrosi R. The Impact of Weight on Arthroscopic Osteochondral Talar Reconstruction. *Foot Ankle Int*. 2017 Jun;38(6):612-620.
 24. Giacchino R, Zancan L, Vajro P, Verucchi G, Resti M, Barbera C, Maccabruni A, Marcellini M, Balli F, Cascio A, Nebbia G, Crivellaro C, Bortolotti F, Clemente MG, Bragetti P, Valentini P, Mazzoni N,

25. Losurdo G, Cristina E. Hepatitis B virus infection in native versus immigrant or adopted children in Italy following the compulsory vaccination. *Infection*. 2001 Aug;29(4):188-91.

Rickettsiosis

26. Mendell NL, Bouyer DH, Walker DH. Murine models of scrub typhus associated with host control of *Orientia tsutsugamushi* infection. *PLoS Negl Trop Dis*. 2017.
27. Colomba C, Saporito L, Polara VF, Rubino R, Titone L. Mediterranean spotted fever: clinical and laboratory characteristics of 415 Sicilian children. *BMC Infect Dis*. 2006 Mar 22;6:60.
28. Colomba C, Saporito L, Siracusa L, Giammanco G, Bonura S, Titone L. Mediterranean spotted fever in paediatric and adult patients: two clinical aspects of the same disease. *Infez Med*. 2011 Dec;19(4):248-53.
29. Parola P, Paddock CD, Socolovschi C, Labruna MB, Mediannikov O, Kernif T, Abdad MY, Stenos J, Bitam I, Fournier PE, Raoult D. Update on tick-borne rickettsioses around the world: a geographic approach. *Clin Microbiol Rev*. 2013 Oct;26(4):657-702. doi: 10.1128/CMR.00032-13. Review. Erratum in: *Clin Microbiol Rev*. 2014 Jan;27(1):166.
30. Colomba C, Trizzino M, Giammanco A, Bonura C, Di Bona D, Tolomeo M, Cascio A. Israeli Spotted Fever in Sicily. Description of two cases and mini review. *Int J Infect Dis*. 2017 Apr 10;61:7-12. [Epub ahead of print] Review.
31. Cascio A, Torina A, Valenzise M, Blanda V, Camarda N, Bombaci S, Iaria C, De Luca F, Wasniewska M. Scalp eschar and neck lymphadenopathy caused by *Rickettsia massiliae*. *Emerg Infect Dis*. 2013 May;19(5):836-7.
32. Colomba C, Siracusa L, Trizzino M, Gioè C, Giammanco A, Cascio A. Myocarditis in Mediterranean spotted fever: a case report and a review of the literature. *JMM Case Rep*. 2016 Aug 30;3(4):e005039.
33. Saporito L, Giammanco GM, Rubino R, Ingrassia D, Spicola D, Titone L et al. Severe Mediterranean spotted fever complicated by acute renal failure and herpetic oesophagitis. *J Med Microbiol* 2010;59(Aug (Pt 8)):990–2.
34. Colomba C, Imburgia C, Trizzino M, Titone L. First case of Mediterranean spotted fever-associated rhabdomyolysis leading to fatal acute renal failure and encephalitis. *Int J Infect Dis*. 2014 Sep;26:12-3.
35. Colomba C, Siracusa L, Madonia S, Saporito L, Bonura C, De Grazia S, Giammanco GM. A case of spotted fever rickettsiosis in a human immunodeficiency virus-positive patient. *J Med Microbiol*. 2013 Sep;62(Pt 9):1363-4.
36. Colomba C, Saporito L, Colletti P, Mazzola G, Rubino R, Pampinella D, Titone L. Atrial fibrillation in Mediterranean spotted fever. *J Med Microbiol*. 2008 Nov;57(Pt 11):1424-6.
37. Cascio A, Colomba C. Macrolides in the treatment of children with Mediterranean spotted fever. *Infez Med*. 2002 Sep;10(3):145-50.
38. Cascio A, Colomba C, Antinori S, Paterson DL, Titone L. Clarithromycin versus azithromycin in the treatment of Mediterranean spotted fever in children: a randomized controlled trial. *Clin Infect Dis*. 2002 Jan 15;34(2):154-8.
39. Cascio A, Colomba C, Di Rosa D, Salsa L, di Martino L, Titone L. Efficacy and safety of clarithromycin as treatment for Mediterranean spotted fever in children: a randomized controlled trial. *Clin Infect Dis*. 2001 Aug 1;33(3):409-11. Epub 2001 Jun 21. Erratum in: *Clin Infect Dis* 2001 Sep 1;33(5):749.

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40. Saporito L, Giammanco GM, De Grazia S, Colomba C. Visceral leishmaniasis: host-parasite interactions and clinical presentation in the immunocompetent and

- in the immunocompromised host. *Int J Infect Dis.* 2013 Aug;17(8):e572-6.
41. World Health Organization. Control of the leishmaniasis. *World Health Organ Tech Rep Ser* 2010;(949):xii–xiii, 1–186.
 42. Antinori S, Schifanella L, Corbellino M. Leishmaniasis: new insights from an old and neglected disease. *Eur J Clin Microbiol Infect Dis* 2012;31:109–18.
 43. Colomba C, Saporito L, Vitale F, Reale S, Vitale G, Casuccio A et al. Cryptic *Leishmania infantum* infection in Italian HIV infected patients. *BMC Infect Dis* 2009;9:199.
 44. Colomba C, Saporito L, Polara VF, Barone T, Corrao A, Titone L. Serological screening for *Leishmania infantum* in asymptomatic blood donors living in an endemic area (Sicily, Italy). *Transfus Apher Sci.* 2005 Nov;33(3):311-4. Epub 2005 Oct 4.
 45. Cascio A, Colomba C, Antinori S, Orobello M, Paterson D, Titone L. Pediatric visceral leishmaniasis in western Sicily, Italy: a retrospective analysis of 111 cases. *Eur J Clin Microbiol Infect Dis* 2002; 21:277–82.
 46. Cascio A, Colomba C. Childhood Mediterranean visceral leishmaniasis. *Infez Med* 2003; 11:5–10.
 47. Colomba C, Scarlata F, Salsa L, Frasca Polara V, Titone L. Mediterranean visceral leishmaniasis in immunocompetent children. Report of two cases relapsed after specific therapy. *Infez Med* 2004;12:139–43.
 48. Mansueto P, Seidita A, Vitale G, Cascio A. Leishmaniasis in travelers: a literature review. *Travel Med Infect Dis.* 2014 Nov-Dec;12(6 Pt A):563-81.
 49. Cascio A, Calattini S, Colomba C, Scalamogna C, Galazzi M, Pizzuto M et al. Polymerase chain reaction in the diagnosis and prognosis of Mediterranean visceral leishmaniasis in immunocompetent children. *Pediatrics* 2002;109:E27.
 50. Antinori S, Calattini S, Longhi E, Bestetti G, Piolini R, Magni C, Orlando G, Gramiccia M, Acquaviva V, Foschi A, Corvasce S, Colomba C, Titone L, Parravicini C, Cascio A, Corbellino M. Clinical use of polymerase chain reaction performed on peripheral blood and bone marrow samples for the diagnosis and monitoring of visceral leishmaniasis in HIV-infected and HIV-uninfected patients: a single-center, 8-year experience in Italy and review of the literature. *Clin Infect Dis.* 2007 Jun 15;44(12):1602-10. Epub 2007 May 7. Review.
 51. Simon I, Wissing KM, Del Marmol V, Antinori S, Rimmelink M, Nilufer Broeders E, Nortier JL, Corbellino M, Abramowicz D, Cascio A. Recurrent leishmaniasis in kidney transplant recipients: report of 2 cases and systematic review of the literature. *Transpl Infect Dis.* 2011 Aug;13(4):397-406.
 52. Antinori S, Cascio A, Parravicini C, Bianchi R, Corbellino M. Leishmaniasis among organ transplant recipients. *Lancet Infect Dis* 2008;8:191–9.
 53. Di Masi F, Ursini T, Iannece MD, Chianura L, Baldasso F, Foti G, Di Gregorio P, Casabianca A, Storaci N, Nigro L, Colomba C, Marazzi MG, Todaro G, Tordini G, Zanelli G, Cenderello G, Acone N, Polilli E, Migliore S, Almi P, Pizzigallo E, Sagnelli E, Mazzotta F, Russo R, Manzoli L, Parruti G. Five-year retrospective Italian multicenter study of visceral leishmaniasis treatment. *Antimicrob Agents Chemother.* 2014;58(1):414-8.
 54. Cascio A, di Martino L, Occorsio P, Giacchino R, Catania S, Gigliotti AR, Aiassa C, Iaria C, Giordano S, Colomba C, Polara VF, Titone L, Gradoni L, Gramiccia M, Antinori S. A 6 day course of liposomal amphotericin B in the treatment of infantile visceral leishmaniasis: the Italian experience. *J*

- Antimicrob Chemother. 2004 Jul;54(1):217-20. Epub 2004 May 18.
55. Colomba C, Di Carlo P, Scarlata F, Iaria C, Barberi G, Famà F, Cama V, Cascio A. Visceral leishmaniasis, hypertriglyceridemia and secondary hemophagocytic lymphohistiocytosis. *Infection*. 2016 Jun;44(3):391-2.
56. Colomba C, Adamoli L, Trizzino M, Siracusa L, Bonura S, Tolomeo M, Cajozzo M, Giammanco GM. A case of visceral leishmaniasis and pulmonary tuberculosis in a post-partum woman. *Int J Infect Dis*. 2015 Apr;33:5-6.
57. Cascio A, Iaria M, Iaria C. Leishmaniasis and biologic therapies for rheumatologic diseases. *Semin Arthritis Rheum* 2010; 40:e3-5.
- Brucellosis*
58. Ministero della Sanità. Bollettino Epidemiologico. Available from: URL: http://www.sanita.it/sanita/malinf/BollEpid/illnes_all/.
59. Geraci D, Locorotondo G, Parlato A, Cocchiara R, Caracappa S, Scarlata F, Cascio A. Enzyme-linked immunosorbent assay for *Brucella melitensis*-associated antigens. *Microbiologica*. 1988 Jul;11(3):213-8.
60. Colomba C, Siracusa L, Rubino R, Trizzino M, Scarlata F, Imburgia C, Titone L. A case of *Brucella* endocarditis in association with subclavian artery thrombosis. *Case Rep Infect Dis*. 2012;2012:581489.
61. Cascio A, Iaria C. Brucellar aortitis and brucellar spondylitis. *Lancet Infect Dis*. 2015 Feb;15(2):145-6.
62. Cascio A, Iaria C. *Brucella* aortitis: an underdiagnosed and under-reported disease. *Int J Rheum Dis*. 2014 Sep;17(7):825.
63. Cascio A, De Caridi G, Lentini S, Benedetto F, Stilo F, Passari G, Iaria C, Spinelli F, Pappas G. Involvement of the aorta in brucellosis: the forgotten, life-threatening complication. A systematic review. *Vector Borne Zoonotic Dis*. 2012 Oct;12(10):827-40.
64. Tolomeo M, Di Carlo P, Abbadessa V, Titone L, Miceli S, Barbusca E, Cannizzo G, Mancuso S, Arista S, Scarlata F. Monocyte and lymphocyte apoptosis resistance in acute and chronic brucellosis and its possible implications in clinical management. *Clin Infect Dis*. 2003 Jun 15;36(12):1533-8.
65. Cascio A, Scarlata F, Giordano S, Antinori S, Colomba C, Titone L. Treatment of human brucellosis with rifampin plus minocycline. *J Chemother*. 2003 Jun;15(3):248-52.
66. Joint Food and Agriculture Organization, World Health Organization. FAO-WHO Expert Committee on brucellosis. 6th report. WHO Technical Report Series, no. 740. WHO Geneva 1986.
67. Lubani MM, Dudin KI, Sharda DC et al. A multicenter therapeutic study of 1100 children with brucellosis. *Pediatr Infect Dis J* 1989;8(2):75-78.
68. Street MAJL Jr, Grant WW, Alva JD. Brucellosis in childhood. *Pediatrics* 1975; 55(3):416-421.
69. Llorens-Terol J, Busquests RM. Brucellosis treated with rifampin. *Arch Dis Child* 1980;55(6):486-488.
70. Cascio A, Di Liberto C, D'Angelo M, Iaria C, Scarlata F, Titone L, Campisi G. No findings of dental defects in children treated with minocycline. *Antimicrob Agents Chemother*. 2004 Jul;48(7):2739-41.
- Other rare zoonotic diseases*
71. Cascio A, Stassi G, Cacciola I, Saitta C, Squadrito G. Fever and rhomboid target lesion in decompensated cirrhosis *Lancet Infect Dis*. 2012 Jul;12(7):576. doi: 10.1016/S1473-3099(12)70063-4.
72. Pappas G, Cascio A. Optimal treatment of leptospirosis: queries and projections. *Int J Antimicrob Agents*. 2006 Dec;28(6):491-6.

73. Arista S, Giammanco Gm, De Grazia S, Migliore Mc, Martella V, Cascio A. Molecular characterization of the genotype G9 human rotavirus strains recovered in Palermo, Italy, during the winter of 1999-2000. *Epidemiol Infect* 2004;132:343-349.
74. Arista S, Vizzi E, Migliore Mc, Di Rosa E, Cascio A. High incidence of G9P181 rotavirus infections in Italian children during the winter season 1999-2000. *Eur J Epidemiol* 2003;18:711-714.
75. Arista S, Giammanco GM, De Grazia S, Colomba C, Martella V, Cascio A, Iturriza-Gòmara M. G2 rotavirus infections in an infantile population of the South of Italy: variability of viral strains over time. *J Med Virol.* 2005 Dec;77(4):587-94.
76. Arista S, Vizzi E, Alaimo C, Palermo D, Cascio A. Identification of human rotavirus strains with the P[14] genotype by PCR. *J Clin Microbiol* 1999;37:2706-2708.
77. Foti M, Rinaldo D, Guercio A, Giacopello C, Aleo A, De Leo F, Fisichella V, Mammina C. Pathogenic microorganisms carried by migratory birds passing through the territory of the island of Ustica, Sicily (Italy). *Avian Pathol.* 2011 Aug;40(4):405-9.
78. Castelli E, Viviano E, Torina A, Caputo V, Bongiorno MR. Avian mite dermatitis: an Italian case indicating the establishment and spread of *Ornithonyssus bursa* (Acari: Gamasida: Macronyssidae) (Berlese, 1888) in Europe. *Int J Dermatol.* 2015 Jul;54(7):795-9.